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12 **Efficacy of modified atmospheres on *Trogoderma granarium* and *Sitophilus zeamais***

13

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21

22 **Abstract**

23 We investigated the efficacy of two types of modified atmospheres (MA) against adults  
24 of *Trogoderma granarium* and *Sitophilus zeamais* under laboratory conditions. Adults of  
25 the above species were exposed at increased carbon dioxide (CO<sub>2</sub>) of 70% or at low  
26 oxygen (O<sub>2</sub>) of 0.1% for 0.67 (16 h), 1, 2, 4 and 6 d. After each exposure interval,  
27 mortality (immediate mortality) and knockdown was recorded and the surviving or  
28 knocked down individuals were transferred to normal conditions, where mortality was  
29 recorded again 7 d later (delayed mortality). Additionally, after the immediate and  
30 delayed mortality counts, all adults were removed from the treated substrate, and the  
31 number of progeny production was recorded 60 d later. Both MA condition totally  
32 controlled the adults of *T. granarium* and *S. zeamais* after 6 d of immediate exposure, or  
33 4 d when delayed exposure was taken into account, revealing the post exposure effect of  
34 the MA. Moreover, high CO<sub>2</sub> was more effective than low O<sub>2</sub> for *S. zeamais*, while the  
35 reverse was true for *T. granarium*. The 4 d exposure period was crucial for the progeny  
36 production of both species, since in that period the transferred knocked down insects,  
37 after their exposure to MA, did not produce progeny. From the results of the present  
38 study, we can conclude that both MA can be used with success against the two target  
39 species.

40

41 **Key words:** modified atmospheres, khapra beetle, maize weevil, carbon dioxide, low  
42 oxygen; short exposures; post exposure.

43

44

## Introduction

45           The khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae) and  
46 the maize weevil, *Sitophilus zeamais* Motschulsky (Coleoptera: Curculionidae) are major  
47 stored product beetle species that infest a variety of stored grains and processed  
48 commodities, causing enormous quantitative and qualitative losses. *Trogoderma*  
49 *granarium* is a quarantine species in many parts of the world, has a remarkable tolerance  
50 to many of the currently used control methods, while its larvae can remain at diapause for  
51 extremely long intervals (EPPO 2017, Athanassiou et al. 2019). Moreover, *S. zeamais* is  
52 an important primary colonizer of grains, capable of a rapid population growth  
53 (Athanassiou et al. 2017). As in the case of all stored product insect species, chemical  
54 control is the main method used for the control of both species, such as the application of  
55 the fumigant phosphine, or the use of sprayable formulations of pyrethroids,  
56 organophosphates and other active ingredients. Nevertheless, both species have  
57 developed resistance to various traditional insecticides such as phosphine (Bell and  
58 Wilson 1995, Pimentel et al. 2009, Athanassiou et al. 2019) and some pyrethroids  
59 (Guedes et al. 1995, Kumar et al. 2010), while many novel active ingredients have been  
60 proved ineffective (Letellier et al. 1995, Haddi et al. 2015, Kavallieratos et al. 2016).

61           Modified atmospheres (MA) is an environmentally-friendly alternative control  
62 method, which can provide disinfestation of stored products from a wide range of pests  
63 without leaving toxic residues (Banks and Fields 1995). The application of MA results in  
64 the change of the concentration of gases like carbon dioxide (CO<sub>2</sub>), oxygen (O<sub>2</sub>) and  
65 nitrogen (N<sub>2</sub>) in the stored product environment, which results on an atmosphere that is  
66 toxic for the target species (Jayas and Jeyamkondan 2002, Navarro et al. 2012). The toxic

67 effect of MA can be achieved by adding CO<sub>2</sub> or N<sub>2</sub> in order to produce an atmosphere  
68 with high CO<sub>2</sub> or low O<sub>2</sub> concentrations or a combination of both. The toxic effect of the  
69 MA on insects depends on the concentration of the gases, the insect species, the  
70 development stage, the insect age and the exposure time (Fleurat-Lessard 1990, Navarro  
71 2006). The reduction of exposure time without loss in efficacy levels is essential to  
72 enhance their use (Ruidavets 2014), although MA are considered as slow-acting control  
73 methods because exposed insects are able to gradually reduce their metabolic rate in  
74 order to overcome the toxic effect of changes in the ratio of gases in the atmosphere  
75 (Navarro 2006, Mitcham et al. 2006). The exposure of the insects to MA may result in  
76 their immobilization, which, after their removal from the treated substrate, may lead to  
77 either death or recovery. Important factors that will determine the outcome of this  
78 delayed effect after the return of the insects to normal conditions, are the exposure time  
79 and the toxicity of the MA environment, which increase with the decrease of the O<sub>2</sub> and  
80 the increase of the CO<sub>2</sub> to toxic levels (Fleurat-Lessard 1990, Navarro 2006, Mitcham et  
81 al. 2006).

82           The efficacy of MA is well documented for several stored product pests, with a  
83 variety of O<sub>2</sub> or CO<sub>2</sub> contents (Krishnamurthy et al. 1986, Banks and Annis 1990, White  
84 et al. 1995, Riudavets et al. 2009, Navarro 2012, Iturralde-Garcia et al. 2016). For *T.*  
85 *granarium*, mostly in the case of its larvae, older works mainly examined different  
86 percentages of low O<sub>2</sub> or high CO<sub>2</sub> alone or in combination, at various temperatures,  
87 relative humidity and pressure levels (Navarro 2006, 2012). In contrast, to our  
88 knowledge, the data for the control of *S. zeamais* are mostly focused on the application of

89 high levels of CO<sub>2</sub> (Banks and Annis 1990, Carvalho et al. 2012, Noomhorm et al. 2013),  
90 while the data for low O<sub>2</sub> are extremely few (Bailey and Banks 1975, Haojie et al. 2014).

91 Any application of MA should take into account the time that is needed for a  
92 satisfactory level of efficacy, as long intervals may allow insects to continue to cause  
93 damage on the commodity. Furthermore, delayed mortality may not deter progeny  
94 production, which will further increase the infestation after the termination of the  
95 application. This is particularly important, as, in many cases, eggs and larvae are less  
96 susceptible than adults in MA (Banks and Annis 1990, Riudavets 2009). In this context,  
97 we have tested the use of either low O<sub>2</sub> or high CO<sub>2</sub> for the control of *T. granarium* and  
98 *S. zeamais* adults, at various exposure intervals, by taking into account immediate and  
99 delayed mortality, as well as progeny production capacity.

100

101

## Materials and Methods

102

103 **Insects.** *Trogoderma granarium* was reared on rice, while *S. zeamais* was reared  
104 on wheat at 28 ± 2 °C, 70 ± 5% relative humidity (r.h.) and 16:8 light:dark photoperiod.  
105 All insects were obtained from cultures maintained in IRTA climatic chambers. Adults of  
106 mixed age and sex were used in the tests for both species.

107 **Bioassays.** Bioassays were carried out by exposing the adults of each species to  
108 two types of MA, i.e. either 70% CO<sub>2</sub>, 6% O<sub>2</sub> and 24% N<sub>2</sub> or 99.9% N<sub>2</sub> and 0.1% O<sub>2</sub>, for  
109 16 h (0.67 d), 1, 2, 4 and 6 d. Plastic cups of 400 ml capacity with holes on their covers  
110 were used as experimental units. For each species and exposure interval one cup was  
111 prepared by adding 10 g of wheat and 10 adults (separate cups for each species). After

112 that, the cups were placed into a 300 X 210 mm and 59 mm-thick plastic vacuum  
113 Cryovac Bag (Sealed Air, USA), in order to apply the desired type of MA.

114 The desirable MA conditions were prepared with a gas mixer (KM 100-3M, Witt,  
115 Germany) and were applied to the plastic vacuum bags by using a vacuum packing  
116 machine (EVT-10, Tecnotrip, Spain). Afterwards, a second sealing took place by using a  
117 portable sealer (Sealboy, Audion Elektro BV, The Netherlands) in order to avoid leakage  
118 of gasses. Then, the verification of the CO<sub>2</sub>, O<sub>2</sub> and N<sub>2</sub> containment in the bags, was  
119 carried out by using a gas analyzer (Oxybaby, Witt, Germany). After the application of  
120 the gases, the bags containing the plastic cups were stored in climatic chamber set at 28 ±  
121 2 °C, 70 ± 5% r. h. and 16:8 light:dark photoperiod for the tested exposure intervals.

122 Following each exposure interval, the bags were taken from the climatic chamber  
123 in order to measure the gas containment, through the gas analyzer. Subsequently, the bags  
124 and the cups were opened and dead, alive and knocked down adults (not having the  
125 ability to move, but showing a minimum movement of tarsi or antennae) were counted  
126 under stereoscope. Dead adults were removed, while the alive and knocked down adults  
127 were placed in new cups containing 10 g of new wheat and returned in the climatic  
128 chambers at the same conditions for 7 d more, in order to evaluate the post exposure  
129 effect. After the termination of the 7 d interval the cups were opened and dead and alive  
130 adults were counted and removed and then, the cups were returned to the climatic  
131 chambers and remained for additional period of 60 d in order to measure the progeny  
132 production capacity. This was also done with the cups after the termination of the initial  
133 exposure period for measuring the progeny production. For each exposure interval there





157 differences between the two MA treatments were noted only at the 2 and 4 d exposure  
158 intervals. After 4 d of exposure, mortality in high CO<sub>2</sub> was 100%, while in low O<sub>2</sub>  
159 reached merely 70%. For this species, mortality for both treatments was 100% after 6 d  
160 of exposure (Fig. 1). The calculated LT<sub>50</sub> and LT<sub>99</sub> values also indicate that *T. granarium*  
161 adults were more sensitive to low O<sub>2</sub> conditions while *S. zeamais* adults were more  
162 sensitive to high CO<sub>2</sub> conditions (Table 2).

163 Knockdown effect after exposure to high CO<sub>2</sub> was stronger to *S. zeamais*  
164 compared to *T. granarium* (Fig. 2A and 3). Similar results have been also recorded after  
165 exposure to low O<sub>2</sub>, although the knockdown effect was weaker (Fig. 4A and 5A).

166 **Delayed mortality.** For delayed mortality all factors were significant with the  
167 exception of treatment and treatment X exposure for *T. granarium* (Table 1). For both  
168 species, delayed mortality was increased with the increase of the initial exposure in both  
169 treatments. For *T. granarium* mortality increased and reached 100% from the 4 d  
170 exposure in both MA treatments, while there were no significant differences among MA  
171 treatments in all exposure intervals (Fig. 6). In the case of *S. zeamais*, significant  
172 differences between treatments were noted only for the 1 and 2 d of exposure, while  
173 mortality was generally higher at high CO<sub>2</sub> compared to low O<sub>2</sub>. For example for the 2 d  
174 exposure interval at low O<sub>2</sub> mortality was 14% while at high CO<sub>2</sub> 100%. Moreover,  
175 mortality reached 100% at low O<sub>2</sub> after 4 d of exposure (Fig. 6).

176 The number of knocked down or alive adults after the initial exposure, did not  
177 affect their delayed response (7 d later) (Fig. 2, 3, 4 and 5) dead or alive condition. For  
178 the first three exposure intervals, all adults that had been initially recorded as knocked  
179 down, were either alive or dead 7 d later. Nevertheless, for longer intervals all adults that

180 had been recorded as knocked down were dead at the post exposure period (Fig. 2, 3, 4  
181 and 5).

182 **Progeny Production.** For both species and treatments, no progeny was produced  
183 after the initial exposure of the adults (Table 3). From the adults that had survived after  
184 the high CO<sub>2</sub> treatment, those that were classified as alive were able to produce progeny  
185 only at the 0.67 d exposure interval for *T. granarium*. Similarly, at the low O<sub>2</sub> treatment  
186 for both species, alive adults were able to produce progeny only at the 0.67 and 1 d  
187 exposure intervals. Moreover, for the adults of both species that had been classified as  
188 knocked down, no progeny production was recorded after 4 and 6 d of exposure in both  
189 MA treatments (Table 3).

190

## 191 **Discussion**

192

193 Our results indicate that both treatments were highly effective against the adults  
194 of *T. granarium* and *S. zeamais*, and totally controlled the adults of both species after 6 or  
195 4 d of exposure, in terms of immediate and delayed mortality, respectively. For 60% CO<sub>2</sub>  
196 on *T. granarium*, Spratt et al. (1985) reported that all adults died within 5 or 6 days at 20  
197 °C and in less than a 4 d at 30 °C, which is in accordance with our findings. Nevertheless,  
198 our data set shows that this species was more susceptible to reduced O<sub>2</sub> than to increased  
199 CO<sub>2</sub>. Still, given that we only considered only two treatments, generalizations should be  
200 avoided, as more levels of either O<sub>2</sub> or CO<sub>2</sub> are needed to understand which technique is  
201 faster for the control of *T. granarium*. However, this difference in efficacy between high  
202 CO<sub>2</sub> and low O<sub>2</sub> treatments was also reported by Banks and Annis (1990) for *T.*

203 *granarium* pupae and diapausing larvae, where the authors calculated the LT<sub>95</sub> for pupae  
204 in 4d at 0.0% O<sub>2</sub> and in 5.5d at 60% CO<sub>2</sub>. Since both techniques are compatible with  
205 organic food production, they should be further examined, especially in the case of  
206 quarantine and pre-shipment treatments.

207 The two methods have been also compared in the past for the granary weevil  
208 *Sitophilus granarius* (L.) (Coleoptera: Curculionidae) by Adler (1994) as well as for the  
209 rice weevil, *Sitophilus oryzae* (L.) (Coleoptera: Curculionidae) by Banks and Annis  
210 (1990). Both of these works reported that high CO<sub>2</sub> kills adults more rapidly compared to  
211 low O<sub>2</sub> (through the increase of N<sub>2</sub>). This observation is in accordance with our findings,  
212 since after exposure to high CO<sub>2</sub>, *S. zeamais* adults were totally controlled after 4 d of  
213 exposure, while the exposure time needed at low O<sub>2</sub> was 6 d. Our findings for the  
214 efficacy of low O<sub>2</sub> to *S. zeamais* is close to what was reported by Haojie et al. (2014),  
215 where the authors found that LT<sub>99.9</sub> for the adults of *S. zeamais* at 98-100% N<sub>2</sub> was 5.57  
216 d. For the efficacy of high CO<sub>2</sub> to adults of *S. zeamais*, in large scale applications on rice,  
217 Carvalho et al. (2012) reported 100% mortality at an exposure interval of 10 d. Under  
218 laboratory conditions for *S. oryzae*, Riudavets et al. (2009) reported that for complete  
219 control of the adults, 4 and 8 d are needed, at 50 and 90% CO<sub>2</sub>, respectively. Our results  
220 show that *S. zeamais* adults are more susceptible to high CO<sub>2</sub> compared to low O<sub>2</sub>, but, as  
221 noted above, this conclusion is applicable only for the conditions tested here, and any  
222 data towards this direction are not transferable to other condition combinations. However,  
223 this observation is in agreement with previous findings reported by Spratt et al. (1985),  
224 where *Sitophilus* spp. are more susceptible to high CO<sub>2</sub> (60%) than to low O<sub>2</sub>, in contrast  
225 with other major stored product insects, such as the rusty grain beetle, *Cryptolestes*

226 *ferrugineus* (Stephens) (Coleoptera: Laemophloeidae) and the red flour weevil, *Tribolium*  
227 *castaneum* (Herbst) (Coleoptera: Tenebrionidae), which are more susceptible to low O<sub>2</sub>.

228         Speed of kill is an essential factor for the selection of the appropriate control  
229 method of the stored grain pests, as the longer the insects remain active in the grains, the  
230 higher the losses are in the commodity. In addition, the need for longer exposures in  
231 controlled environments will increase the overall cost of the method, while keeping the  
232 insects alive for long intervals may pose certain risks for tolerance development (Boyer et  
233 al. 2011). In our study, we have found that for both species, all adults that had been  
234 survived after the 4 d of exposure eventually died, which clearly underlines the  
235 irreversible delayed effects of high CO<sub>2</sub> or low O<sub>2</sub>. Both methods caused, in certain  
236 exposures, narcosis on *T. granarium* and *S. zeamais* adults, as a result of the metabolic  
237 changes that take place on the insects to overcome hypoxia or hypercarbia (Fleurat-  
238 Lessard 1990, Mitcham et al. 2006). Moreover, our results clearly suggest that this  
239 narcotic effect resulted in the elimination of progeny production, which can be  
240 considered more as a consequence of adult immobilization, thus inability for egg laying,  
241 rather than a direct effect of high CO<sub>2</sub> or low O<sub>2</sub> on immature life stages. The proportion  
242 of the adults that had been characterized as knocked down after the treatment was  
243 increased with the increase of the exposure interval, for both methods used. This state of  
244 the insects is critical, as, after their removal from the treated area and their return to  
245 normal atmosphere, knockdown may lead to either recovery or delayed mortality  
246 (Fleurat-Lessard 1990, Mitcham et al. 2006). In our study we found that knockdown is  
247 more likely to lead to delayed mortality than to recovery, even at exposures that were <4  
248 d. Moreover, >2 d of exposure are needed to completely eliminate progeny production

249 capacity of the insects that had survived both treatments. It is well established that  
250 sublethal exposure periods affect the reproduction potential of the insects (Fleurat-  
251 Lessard 1990, Dawson 1995, Navarro et al. 2012). In our experimental protocol we saw  
252 that both treatments worked well in progeny production suppression, either on wheat that  
253 had been treated along with the parental adults, or through the post-exposure effects on  
254 these adults. The scenario of post-exposure effects is considered as a reliable indicator for  
255 the application of other gases as well. For example, the basic protocol for the evaluation  
256 of resistance to phosphine, i.e. the Food and Agriculture Organization (FAO) protocol  
257 (FAO Plant Protection Bulletin 1975), is based on mortality levels that are recorded  
258 during a certain post-exposure period (Daglish 2004, Opit et al. 2012).

259         The overall results, provide data for the critical exposure intervals that are  
260 essential for the control of *T. granarium* and *S. zeamais*, after exposure to either high CO<sub>2</sub>  
261 or low O<sub>2</sub>. Based on our findings, this exposure is 4 d, for both treatments. Despite the  
262 fact it is generally regarded that controlled and/or modified atmospheres are slow-acting  
263 (Navarro 2012), the critical exposure proposed here is directly comparable with the  
264 application of phosphine fumigations, and thus, these levels of CO<sub>2</sub> and O<sub>2</sub> can be further  
265 evaluated as alternatives to conventional phosphine fumigations. Moreover, we found  
266 that low O<sub>2</sub> was preferable than low CO<sub>2</sub> for the control of *T. granarium*, while the  
267 reverse was true for *S. zeamais*. This fact should be taken into account in order to build a  
268 species-mediated application strategy. However, the experimental scenario tested here  
269 was based on adults and their progeny production capacity. Further testing with  
270 immatures is necessary to provide a complete “algorithm” on the critical exposures that  
271 are required to control the most difficult-to-kill life stages of each species.

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## Tables and Figures

**Table 1.** ANOVA parameters for immediate and delayed mortality of the species tested (error df=40)

Immediate Mortality					
	<i>T. granarium</i>			<i>S. zeamais</i>	
	df	<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>
Treatment	1	22.17	<0.01	138.46	<0.01
Exposure	4	75.94	<0.01	396.60	<0.01
Treatment * exposure	4	5.46	<0.01	61.44	<0.01
Delayed Mortality					
	<i>T. granarium</i>			<i>S. zeamais</i>	
	df	<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>
Treatment	1	0.07	0.79	124.26	<0.01
Exposure	4	27.44	<0.01	253.28	<0.01
Treatment * exposure	4	0.54	0.71	50.51	<0.01

**Table 2.** Lethal Time (LT) estimates for 50 and 99% mortality of *T. granarium* and *S. zeamais* adults exposed to either 70% of CO<sub>2</sub> or 0.1% or O<sub>2</sub>

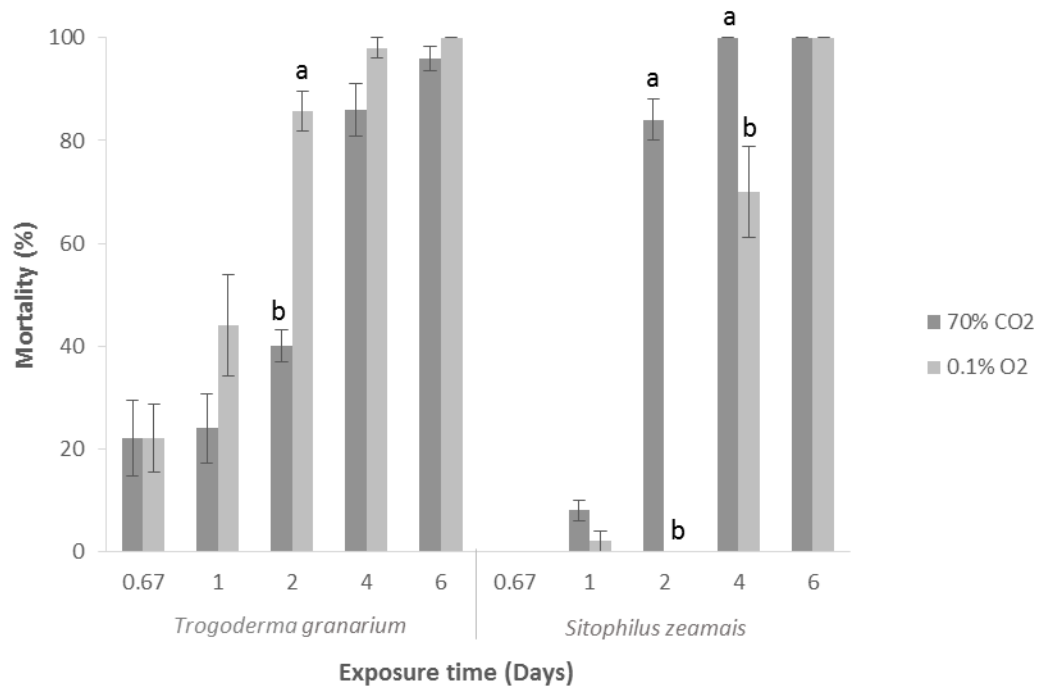
	<i>T. granarium</i>		<i>S. zeamais</i>	
	LT <sub>50</sub> (days)	LT <sub>99</sub> (days)	LT <sub>50</sub> (days)	LT <sub>99</sub> (days)
70% CO <sub>2</sub>	1.6	7.2	1.9	3.7
Probit model	$y=3.43\log(x)+4,35$		$y=4.8\log(x)+4,6$	
0.1% O <sub>2</sub>	1.2	3.7	3.5	6.6
Probit model	$y=8.5\log(x)+2,5$		$y=8.5\log(x)+0,4$	

y value can be expressed to % by using the Probit Transformation Table (Finney's table)

**Table 3.** Mean progeny production (adults per cup) of the tested species after immediate exposure for 0.67, 1, 2, 4, and 6 d to either 70% of CO<sub>2</sub> or 0.1% or O<sub>2</sub> and the corresponding 7-d post-exposure period for each exposure interval, i.e. 7 d after the removal of the alive and knocked down adults from the substrate

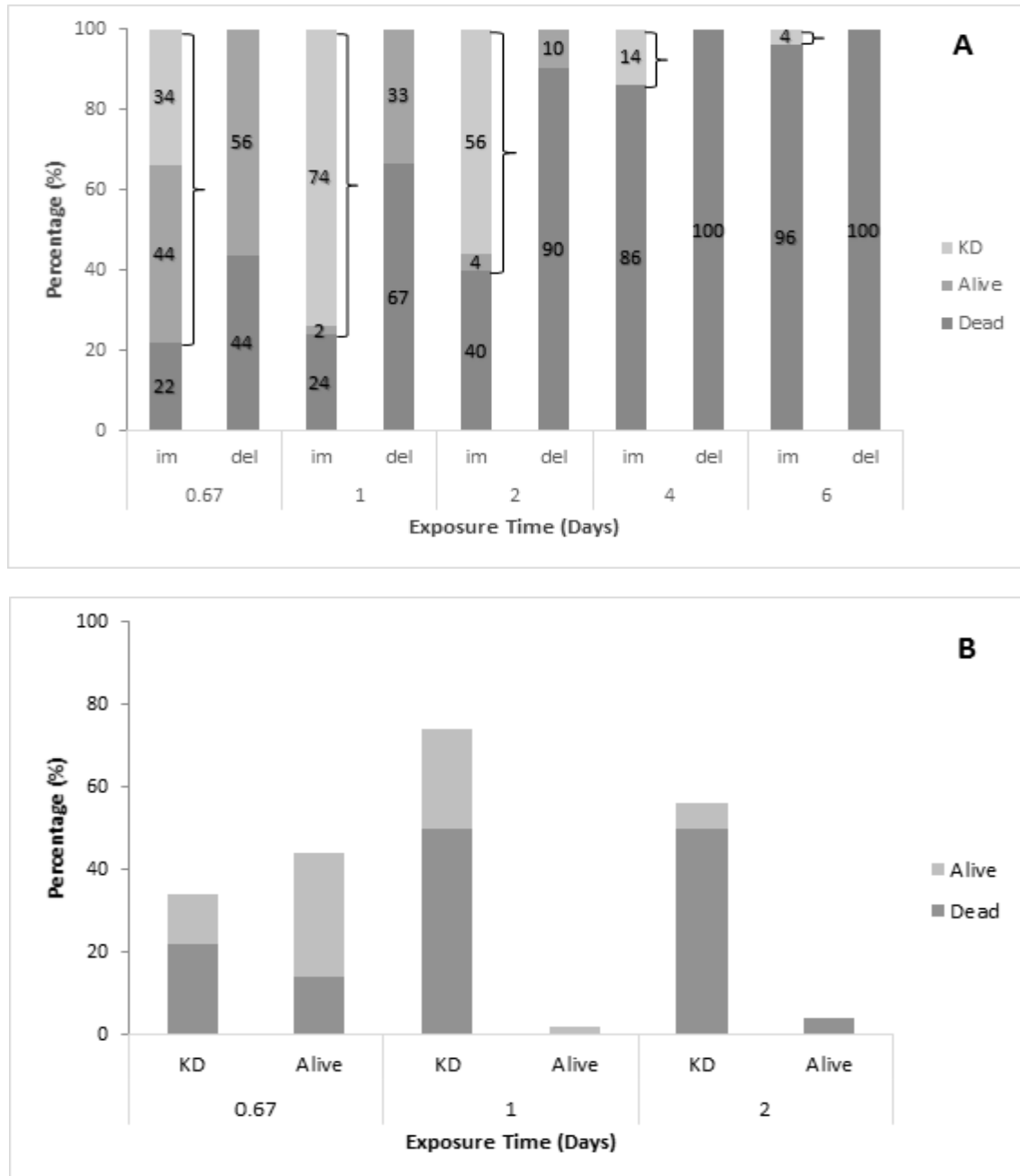
Progeny production after initial exposure										
Exposure (d)	<i>T. granarium</i>					<i>S. zeamais</i>				
	0.67	1	2	4	6	0.67	1	2	4	6
Control	3.5	7.0	48.0	34.0	70.0	8.5	6.5	18.5	48.0	65.0
CO <sub>2</sub>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Control	0.5	40.5	22.5	24.0	59.0	4.5	15.5	22.5	12.0	57.5
O <sub>2</sub>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Progeny production at the 7 d post-exposure period by adults that had been classified as "alive"										
Exposure (d)	<i>T. granarium</i>					<i>S. zeamais</i>				
	0.67	1	2	4	6	0.67	1	2	4	6
Control	38.0	34.0	36.0	52.5	24.0	156.5	120.5	87.5	78.5	67.0
CO <sub>2</sub>	23.4	0.0	0.0	-	-	-	-	-	-	-
Control	38.0	76.0	35.0	19.0	1.5	72.5	102.5	92.0	38.0	98.5
O <sub>2</sub>	10.0	9.6	0.0	-	-	61.2	43.6	-	-	-
Progeny production at the 7 d post-exposure period by adults that had been classified as "knocked down"										
Exposure	<i>T. granarium</i>					<i>S. zeamais</i>				
	0.67	1	2	4	6	0.67	1	2	4	6
Control	38.0	34.0	36.0	52.5	24.0	156.5	120.5	87.5	78.5	67.0
CO <sub>2</sub>	0.0	8.2	3.4	0.2	0.0	32.4	18.0	0.0	-	-
Control	38.0	76.0	35.0	19.0	1.5	72.5	102.5	92.0	38.0	98.5
O <sub>2</sub>	0.8	0.6	0.5	0.0	-	15.0	0.0	41.8	0.0	-

Fig. 1.



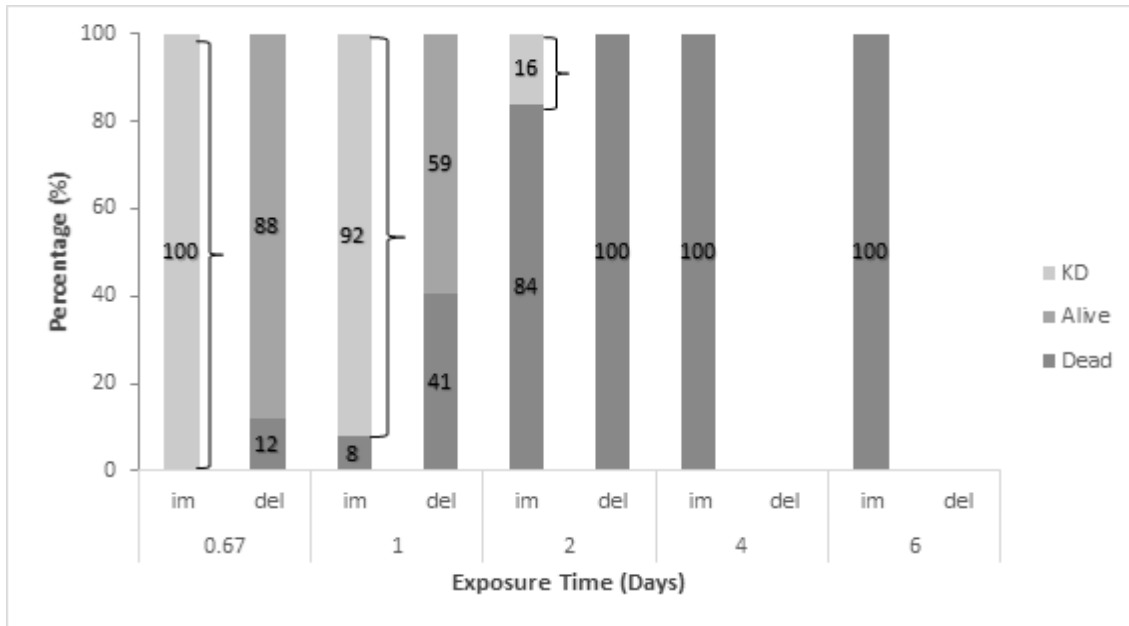
Mean ( $\% \pm$  SE) immediate mortality of *T. granarium* and *S. zeamais* adults exposed to either 70% of CO<sub>2</sub> or 0.1% of O<sub>2</sub>, at five exposure intervals (means among treatments within each species and exposure interval followed by the same lowercase letter are not significantly different; where no letters exist, no significant differences were noted; Tukey-Kramer HSD test at 0.05)

Fig. 2.



Mean (%) immediate (im) percentages of dead, alive and knocked down adults of *T. granarium* exposed to 70% CO<sub>2</sub> for five exposure intervals, and the delayed (del) percentages (%) of alive or dead adults at the post exposure period (7d) of the alive and knockdown (KD) adults (inside brackets) that had been transferred after each exposure interval to normal conditions (A). Mean (%) percentages of dead and alive adults at the post exposure period of Alive or KD adult after their immediate exposure (B).

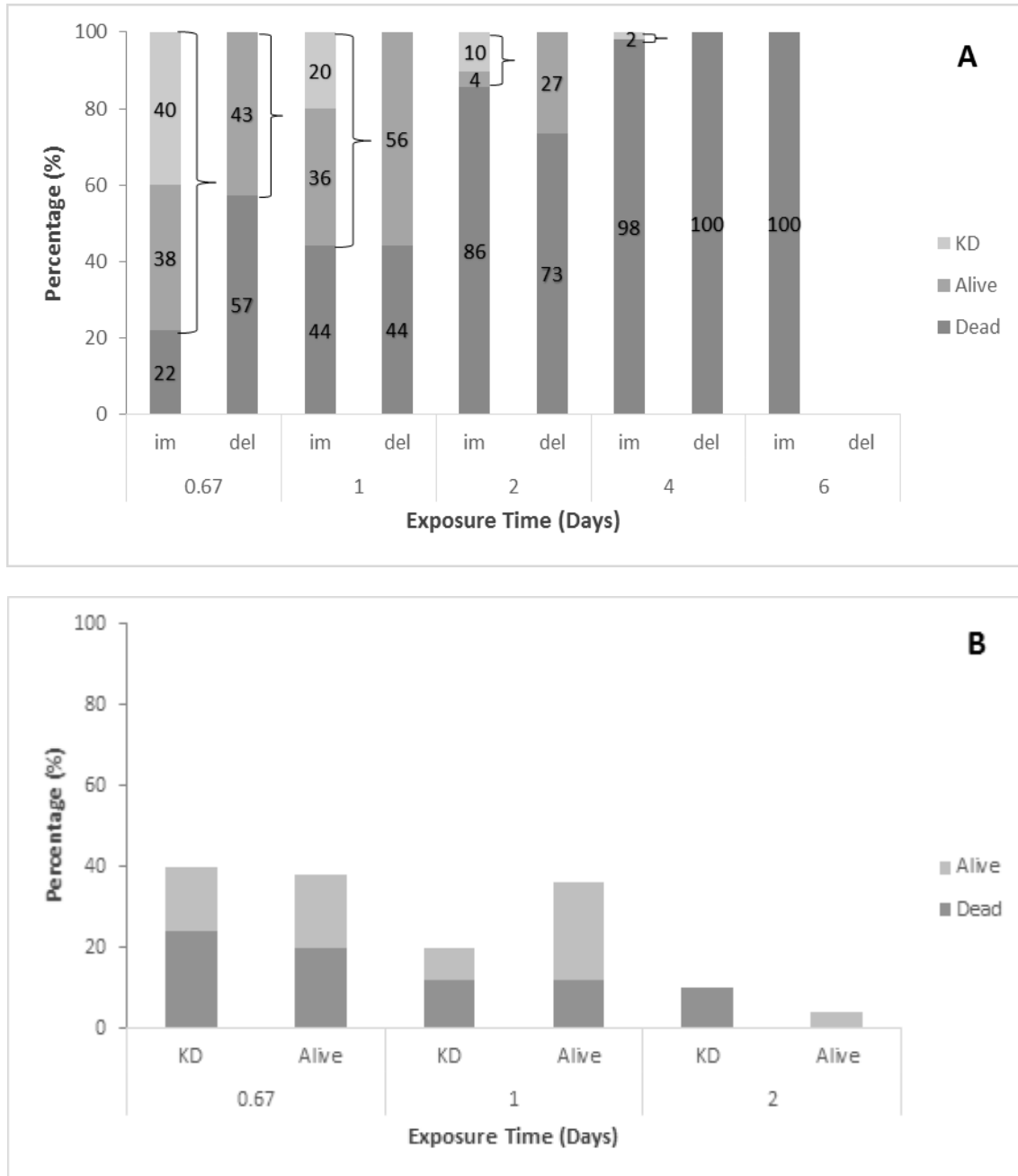
**Fig. 3.**



Mean (%) immediate (im) percentages of dead, alive and knocked down adults of *S. zeamais* exposed to 70% CO<sub>2</sub> for five exposure intervals, and the delayed (del) percentages (%) of alive or dead adults at the post exposure period (7d) of the alive and knockdown (KD) adults (inside brackets) that had been transferred after each exposure interval to normal conditions.

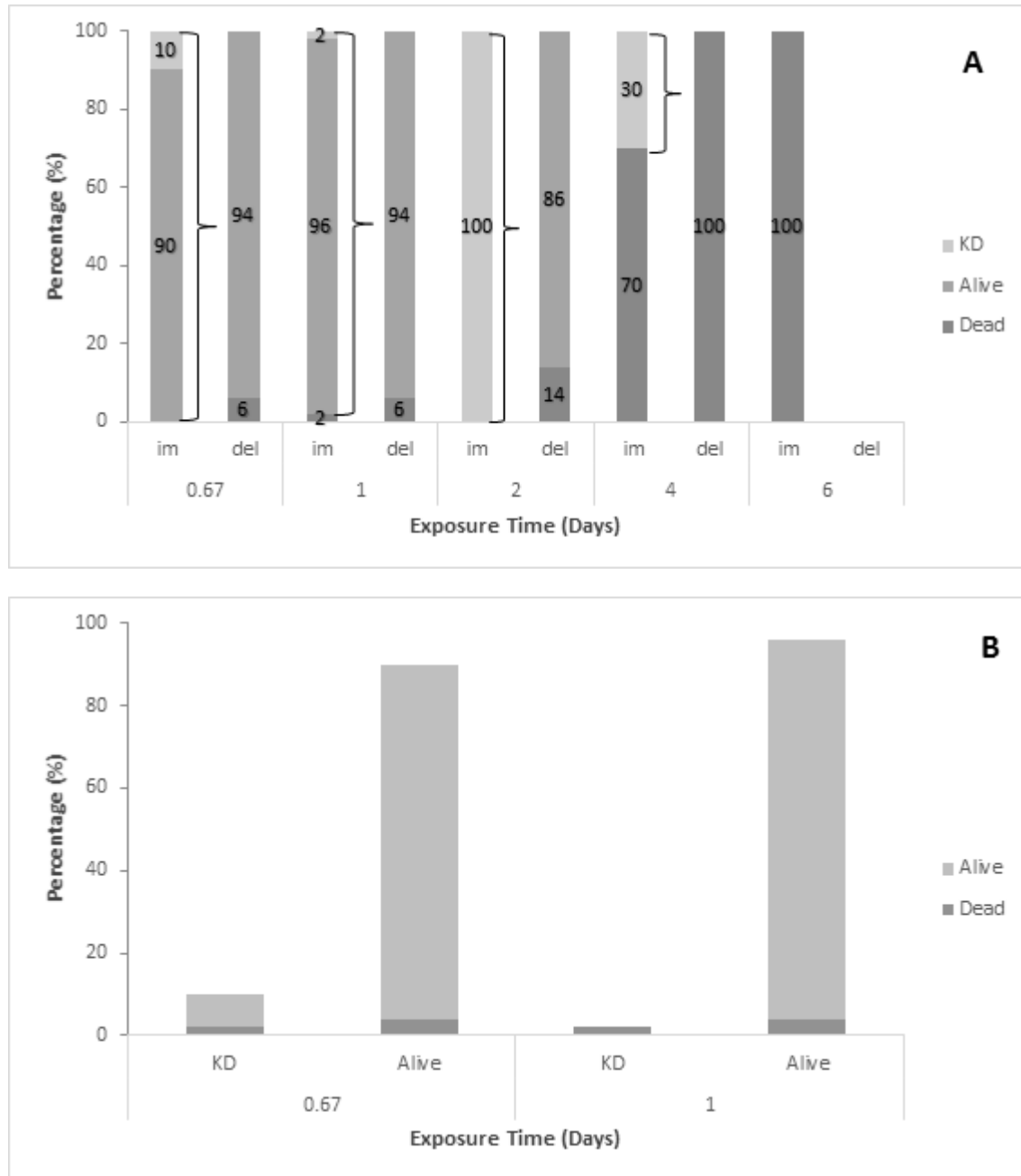


Fig. 4.



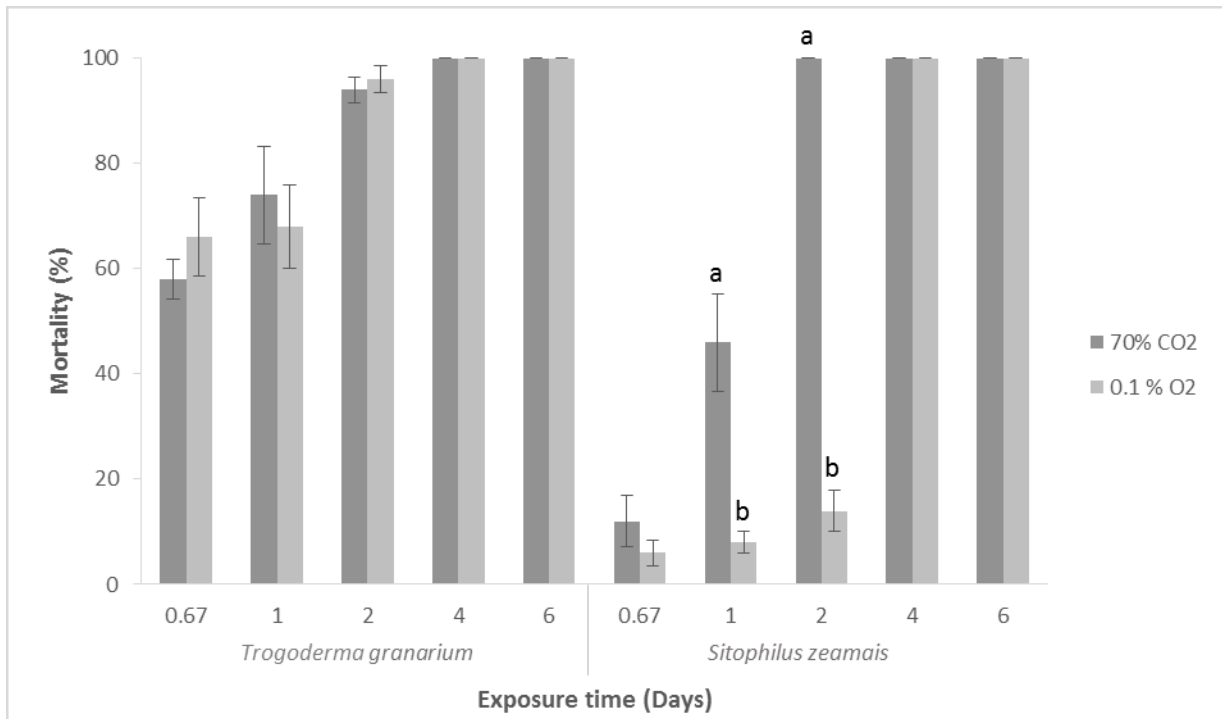
Mean (%) immediate (im) percentages of dead, alive and knocked down adults of *T. granarium* exposed to 0.1% O<sub>2</sub> for five exposure intervals, and the delayed (del) percentages (%) of alive or dead adults at the post exposure period (7d) of the alive and knockdown (KD) adults (inside brackets) that had been initially transferred after each exposure interval to normal conditions (A). Mean (%) percentages of dead and alive adults at the post exposure period of Alive or KD adult after their immediate exposure (B).

Fig. 5.



Mean (%) immediate (im) percentages of dead, alive and knocked down adults of *S. zeamais* exposed to 0.1% O<sub>2</sub> for five exposure intervals, and the delayed (del) percentages (%) of alive or dead adults at the post exposure period (7d) of the alive and knockdown (KD) adults (inside brackets) that had been initially transferred after each exposure interval to normal conditions (A). Mean (%) percentages of dead and alive adults at the post exposure period of Alive or KD adult after their immediate exposure (B).

Fig. 6.



Mean (%  $\pm$  SE) delayed mortality of *T. granarium* and *S. zeamais* adults exposed to either 70% of CO<sub>2</sub> or 0.1% of O<sub>2</sub>, at five exposure intervals (means among treatments within each species and exposure interval followed by the same lowercase letter are not significantly different; where no letters exist, no significant differences were noted; Tukey-Kramer HSD test at 0.05)