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- Ecological and evolutionary drivers of phenotypic and genetic variation in the European
  crabapple (*Malus sylvestris* (L.) Mill.), a wild relative of the cultivated apple
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### 32 Abstract (250 words max with bullet points)

33 Background and Aims

Studying the relationship between phenotypic and genetic variation in populations distributed across environmental gradients can help us understand the ecological and evolutionary processes involved in population divergence. We investigated the patterns of genetic and phenotypic diversity in the European crabapple, *Malus sylvestris*, a wild relative of the cultivated apple (*Malus domestica*) that occurs naturally across Europe in areas subjected to different climate conditions, to test for divergence among populations.

41 Methods

42 Growth rates and traits related to carbon uptake in seedlings collected across Europe 43 were measured under controlled conditions and associated with the genetic status of the 44 seedlings, which was assessed using 13 microsatellite loci and Bayesian clustering 45 method. Isolation-by-distance, -by-climate, and -by-adaptation patterns, which can 46 explain genetic and phenotypic differentiation among *M. sylvestris* populations, were 47 also tested.

48 Key Results

A total of 11.6% of seedlings were introgressed by *M. domestica*, indicating that cropwild gene flow is ongoing in Europe. The remaining seedlings (88.4%) belonged to seven *M. sylvestris* populations. Significant phenotypic trait variation among *M. sylvestris* populations was observed. We did not observe significant isolation-byadaptation; however, the significant association between genetic variation and the climate during the last glacial maximum suggests that there has been local adaptation of *M. sylvestris* to past climates.

56 Conclusions

57 This study provides insight into the phenotypic and genetic differentiation among 58 populations of a wild relative of the cultivated apple. This may help us better utilize its 59 diversity and provide options for mitigating the impact of climate change on the cultivated 60 apple through breeding.

61

Keywords: population structure, isolation-by-distance, isolation-by-ecology, local
 adaptation, climate change, apple tree, crop wild relatives.

64

65

#### 66 Introduction

Knowledge of the spatial phenotypic and genetic variation among populations is essential for 67 68 understanding the ecological (biotic and abiotic factors) and evolutionary (gene flow, selection, 69 drift, and mutation) processes involved in population divergence and adaptation (Savolainen 70 et al. 2013; Sork 2018). Plant species distributed across climatic gradients typically experience 71 spatial variation in selection, genetic drift, and gene flow, which drive genetic and phenotypic 72 divergences among populations (Svenning et al. 2015). Climate indeed influences 73 demographic processes such as population expansion and contraction, the extent of gene flow 74 among populations, and ultimately the degree of genetic divergence among populations 75 (Edwards et al. 2022). For instance, changes in the climate since the last glacial maximum 76 (LGM) 20,000 years ago have driven the genetic composition of the European crabapple and 77 many other tree species (Comes and Kadereit 1998; Kremer et al. 2002; Pyhäjärvi et al. 2008; 78 Cornille, Giraud, et al. 2013; Riordan et al. 2016; Lander et al. 2021; Yamada et al. 2021 p. 79 20221; Parisod 2021). Climate can also shape phenotypic variation among populations. 80 Populations occurring under the same climate may share physiological tolerances to climatic 81 conditions, including plant carbon uptake via photosynthesis. Carbon uptake traits condition 82 plant size and growth, reproduction, and survival under different climatic conditions(Hutyra et 83 al. 2007; Nicotra et al. 2010; Pasho et al. 2012; Way and Montgomery 2015; Mercado et al. 84 2018; Hartmann et al. 2020; Kühn et al. 2021). Variation in plant carbon uptake in response to 85 climatic conditions can result from phenotypic plasticity, i.e., the ability of individual 86 genotypes to produce different phenotypes when exposed to other environmental conditions, 87 in this case, climate (Dusenge et al. 2019). In some cases, local climate can impose divergent 88 selection on carbon uptake traits which leads to reproductive isolation among populations: loci 89 under adaptive divergence act as a local barrier to gene flow (Keller et al. 2011; Franks et al. 90 2014; Aitken and Bemmels 2015; Ramírez-Valiente et al. 2017; Alexandre et al. 2020). 91 Climate can therefore lead to a long-term reduction in gene flow and local adaptation. Whether 92 the phenotypic variation observed in species distributed across large climatic ranges results 93 from their demographic or/and adaptive histories remains an intense topic of investigation (Li 94 et al. 2012; Tiffin and Ross-Ibarra 2014; Briscoe Runquist et al. 2020). Investigating this 95 question can help us predict how plants may respond to climate change and how species adapt 96 to their environment.

97 There are multiple ways to investigate whether genetic and phenotypic variation among 98 populations distributed across climatic gradients results from selection, genetic drift, and/or 99 gene flow. A first step is to use a common garden experiment to investigate the genetic basis 100 of phenotypic variation among populations. Indeed, different populations occurring across a 101 climatic gradient may display clinal variation, i.e., differences in a trait that may result from 102 phenotypic plasticity or local genetic adaptation (Savolainen et al. 2013; de Villemereuil et al. 2016). Measuring candidate traits for adaptation to climate [e.g., phenology (Brachi et al. 2013) 103 104 or traits related to plant carbon uptake (Savolainen et al. 2013; de Villemereuil et al. 2016)] in 105 individuals from different populations under the same environmental conditions can help us 106 elucidate the genetic basis of phenotypic variation across populations without the confounding 107 effects of the environment. Ideally, common garden trials should include the main genetic 108 groups across the species' distribution (de Villemereuil et al. 2020). The association of neutral 109 genotypic variation with phenotypic variation can also be used as evidence of adaptive 110 divergence among populations (Shafer and Wolf 2013; Wang and Bradburd 2014). The 111 correlation between neutral genetic differentiation and environmental or phenotypic 112 divergence among populations, independent of geographic distance, referred to as isolation-113 by-ecology (IBE hereafter), is an extension of the isolation-by-distance (IBD hereafter) model 114 (Wright 1943). IBE patterns have been increasingly used as an indicator of adaptive divergence 115 between populations (Shafer and Wolf 2013). In the IBE model, natural selection, which results 116 from several factors, including climate, can indirectly increase neutral genetic and phenotypic 117 differentiation between populations by promoting general barriers to gene flow (Nosil et al. 2009; Orsini et al. 2013; Shafer and Wolf 2013; Wang and Bradburd 2014). Although it can 118 119 be challenging to map the processes underlying IBE patterns, testing for it is valuable to 120 understand better how natural selection shapes neutral genetic and phenotypic variation. 121 Therefore, evidence from common garden experiments and IBE patterns can contribute to 122 understanding how genotypes, phenotypes, and the environment interact to influence 123 population divergence and potentially local adaptation.

124 Fruit trees are a significant component of terrestrial ecosystems (Petit and Hampe 125 2011). They are grown in managed plantations and orchards to provide a variety of 126 economically important products (Boyd et al. 2013). Recent breeding efforts have involved the 127 repeated use of a limited number of commercial cultivars, leading to a reduction in genetic 128 diversity that can lead to the loss of valuable alleles at genes that are not directly targeted by 129 human selection (Myles et al. 2011; Warschefsky and von Wettberg 2019; Migicovsky et al. 130 2021). Wild relatives of crop fruit trees (hereafter CWR for crop wild relative) harbor 131 phenotypic and genetic diversity that is potentially highly valuable for future breeding 132 programs in the context of climate change (Zhang et al. 2017; Hoban et al. 2018; Hübner and 133 Kantar 2021). However, phenotypic variation in CWRs in relation to climate variation has

134 rarely been studied in fruit trees (Kremer and Hipp 2019). Key traits to study in this context are related to plant carbon uptake. Indeed, climate impacts plant carbon uptake (Aubin et al. 135 136 2016), which impacts fruit quality characteristics and production (Demestihas et al. 2017). 137 These are topical issues as native CWRs can be threatened by crop-to-wild gene flow from 138 nearby domesticated trees (Delplancke et al. 2011; Cornille et al. 2015; Diez et al. 2015; 139 Feurtey et al. 2017; Flowers et al. 2019; Liu et al. 2019). Therefore, studying the genetic and 140 phenotypic variation among CWR fruit tree populations is timely to guide future breeding 141 programs. It may also contribute to understanding the evolutionary and ecological drivers of 142 population divergence, such as climate.

143 The European crabapple, Malus sylvestris (L.) Mill., is a CWR of the cultivated apple, 144 Malus domestica (Cornille et al. 2012, 2014, 2019; Peace et al. 2019). Substantial crop-to-wild 145 gene flow has been observed across M. sylvestris populations in Europe [(up to 23.1% of 146 naturally occurring individuals are introgressed by M. domestica (Cornille et al. 2015)]. Crop-147 wild hybrids sampled in a forest in France and grown in controlled conditions showed higher 148 growth rates compared to wild seedlings (Feurtey et al. 2017). Population genetic analyses also 149 identified five pure (i.e., not introgressed by *M. domestica*) populations in Scandinavia, western 150 France, eastern France, Eastern Europe, and Italy (Cornille et al. 2015). These five populations 151 resulted from past contractions and expansions associated with the last glacial maximum 152 (Cornille, Giraud, et al. 2013; Cornille et al. 2015). Whether these five populations distributed 153 across a large area with different climatic conditions show a phenotypic variation that could 154 result from local adaptation to past and/or present climates is still unknown.

155 We investigated the spatial phenotypic and genetic variation among populations of a 156 wild contributor to the cultivated apple genome, M. sylvestris, to test for adaptive divergence, 157 particularly to climate. Plant growth and traits related to carbon uptake were measured in 584 158 M. sylvestris seedlings grown under controlled conditions and genotyped using 13 159 microsatellite markers. We first assessed the genetic status of each seedling (pure vs. crop-wild 160 hybrid). Then, we compared growth traits and traits related to plant carbon uptake among 161 seedlings from different European genetic groups. We also formally tested the impact of 162 geography (IBD) and ecology (IBE tested with phenotypic traits and climate) on genetic 163 variation observed from 13 microsatellite markers. We investigated the following questions: 164 1) Do growth rates and carbon uptake traits vary between populations of the European 165 crabapple? Are those traits heritable and therefore be good candidates for the responses to 166 selection?; 2) Is there any association between phenotypic variation and genetic variation, 167 taking into account geographic distance, which would suggest adaptive divergence?; and 3) Do

168 we detect a pattern of isolation-by-climate in the European crabapple that could suggest local

- 169 adaptive divergence to climate?
- 170

#### 171 **Materials and Methods**

#### 172 Plant material, experimental design, and trait measurements

173 A total of 584 seeds were collected from 90 M. sylvestris mother trees (three-15 seeds per 174 mother tree, Table S1) from 22 geographical sites in Europe representing the five main genetic 175 groups previously detected in Cornille et al. 2015: Austria (N = 89, two sites), Denmark (N =176 91, three sites), Spain (N = 39, one site), France (N = 220, eight sites), Italy (N = 32, one site) 177 and Romania (N = 117, seven sites) (Table S1). The length, width, and weight of 30 seeds per 178 mother tree were measured with an Opto-Agri (Optomachine, Riom, France).

179 In mid-April 2019, 584 seeds were washed, sterilized (in 0.5% chlorine for 20 min), and 180 stratified for three months at 4°C in the dark in a mix of damp sand and vermiculite. Then, 181 seeds were sowed in jiffy pellets, and each pellet was randomly placed in a 20-hole array. Seeds 182 were grown in controlled conditions for two months (from mid-July to mid-September 2019: 183  $22 \pm 1$  °C,  $60 \pm 5$  % relative humidity, a 16:8 (L:D) photoperiod, and a light level of 40–60 umol m<sup>-2</sup>.s<sup>-1</sup>). Each 20-hole array was rotated daily in the growth chamber to avoid any micro-184 185 environmental variation in plant response, and plants were watered weekly.

186 During the two-month experiment, the number of leaves and height of each seedling was 187 recorded. Due to the low germination rate, some accessions could not be evaluated (i.e., plant 188 height and number of leaves could not be recorded for 19 seedlings out of 584, resulting in N 189 = 565, Table 1). Seedlings were measured every two to three days, starting from days 7-11 190 after the experiment.

191 In the last week of the experiment, the superficial flavonol and chlorophyll contents were 192 measured, and the nitrogen balance index (NBI) was calculated for three leaves per seedling. 193 The superficial chlorophyll content is the chlorophyll concentration in the leaf epidermis 194  $(\mu g/cm^2)$ , and the superficial flavonol content is an index of the flavonoid concentration  $(\mu g/cm^2)$  in this upper layer and is related to phenol accumulation and UV protection. The NBI 195 196 assesses the leaf's nitrogen status by calculating the ratio of chlorophyll to flavonols (related to 197 nitrogen/carbon allocation) and is currently used as a proxy to estimate the foliar nitrogen 198 content (Demotes-Mainard et al. 2008). Leaf chlorophyll, flavonol content, and NBI are 199 parameters correlated with plant carbon uptake via photosynthesis. Flavonol is a phenolic 200 compound that contributes to plant vigor, acclimation, and adaptation to environmental 201 constraints through various mechanisms, including its antioxidant activity. These traits were

- 202 measured using a portable Dualex<sup>®</sup> device (Force-A, Orsay, France), which uses a combination 203 of fluorescence signals at various excitation bands to quantify pigments and chemical 204 compounds, and has already been calibrated on the apple tree (Hamann *et al.* 2018). As carbon 205 uptake-related traits must be measured on the same day, a subsample of 257 seedlings out of 206 the 565 seedlings (Table 1, numbers in brackets) was measured because of time limitations. 207 Seedlings measured for carbon uptake-related traits were selected so that at least one seedling 208 per mother tree and three seedlings per geographical site were sampled.
- 209

### 210 DNA extraction, microsatellite genotyping, and genetic ancestry of the seedlings

At the end of the experiment, the leaves of each seedling were sampled for microsatellite genotyping. Genomic DNA was extracted using the NucleoSpin Plant DNA Extraction Kit II (Macherey & Nagel, Düren, Germany), according to the manufacturer's instructions. Microsatellites were amplified by multiplex PCR with the Multiplex PCR Kit (QIAGEN, Inc.). We used 13 microsatellite markers, Ch01f02, Ch01f03, Ch01h01, Ch01h10, Ch02c06, Ch02c09, Ch02c11, Ch02d08, Ch03d07, Ch04c07, Ch05f06, GD12 and Hi02c07 in four multiplexes (MP01 to MP04; (Cornille *et al.* 2012)).

218 PCR was performed in a final reaction volume of 15 µl (7.5 µl of QIAGEN Multiplex Master 219 Mix, 10–20 µM of each primer with the forward primer labeled with a fluorescent dye, and ten 220 ng template DNA). We used a touch-down PCR program (initial annealing temperature of 221 60°C, decreasing by 1°C per cycle to 55°C). Genotyping was performed at the GENTYANE 222 platform (INRAE Clermont-Ferrand) on an ABI PRISM X3730XL, with 2 ml of GS500LIZ 223 size standard (Applied Biosystems). Alleles were scored with the GENEMAPPER 4.0 software 224 (Applied Biosystems). We retained only multilocus genotypes presenting less than 10% 225 missing data.

Clones or closely related individuals can bias inferences about the population structure. We estimated the kinship coefficient between pairs of individuals (*Fij*) with SPAGeDI 1.5d (Loiselle *et al.* 1995; Hardy and Vekemans 2002), and removed highly genetically related individuals with Fij > 0.5.

The individual-based Bayesian clustering method implemented in STRUCTURE 2.3.3 (Pritchard *et al.* 2000) was used to estimate the admixture between *M. domestica* and *M. sylvestris*, and the population genetic structure of *M. sylvestris*. STRUCTURE uses Markov Chain Monte Carlo (MCMC) simulations to infer the proportion of ancestry of genotypes from *K* distinct clusters. The underlying algorithm minimizes deviations from Hardy–Weinberg, and 235 linkage disequilibria. K ranged from 1 to 10. Ten independent runs were carried out for each K and 500,000 MCMC iterations after a burn-in of 50,000 steps were used. CLUMPAK (Greedy 236 237 algorithm) (Kopelman et al. 2015) was used to identify distinct modes in the ten replicated runs 238 for each K. STRUCTURE analyses were run for the entire dataset (N = 584), plus 40 M. 239 domestica genotypes from Western Europe (Table S2) included as a reference for the cultivated 240 apple gene pool (Cornille, Gladieux, et al. 2013). The R package pophelper v2.3.0 was used 241 (Francis 2016) to visualize bar plots. The amount of additional information explained by 242 increasing K was determined using the  $\Delta K$  statistic (Evanno *et al.* 2005), as implemented in 243 Structure Harvester (Earl and vonHoldt 2012). However,  $\Delta K$  provides statistical support for 244 the strongest but not the finest population structure (Puechmaille 2016). Natural populations 245 can display a hierarchical genetic structure with a fine-scale population structure. Visual inspection of the bar plots was used to identify the K value for which all clusters have well-246 247 assigned individuals, and where additional clusters at higher K values do not have well-248 assigned individuals. Therefore, the K value corresponded to the finest one, which can be 249 higher than the K value of the strongest population structure identified by  $\Delta K$ .

250 We used the membership coefficients at the best K value (inferred with STRUCTURE) for 251 the crop and wild seedlings and defined the genetic status of each seedling. We first separated 252 cultivated apples (seedlings from mother trees that were misidentified in the field, i.e., with a 253 membership proportion to the *M. domestica* gene pool,  $P_{dom}$ , > 0.9, referred to as "dom" 254 hereafter), crop-wild hybrids (seedlings with  $0.1 > P_{dom} > 0.9$ , referred to as "*cw*" hereafter) 255 and pure *M. sylvestris* (seedlings with a membership coefficient to a given wild apple cluster 256 > 0.9 and with  $P_{dom} < 0.1$ , referred to as "*pure*" hereafter). We then separated "pure" wild 257 seedlings (i.e., seedlings with membership coefficients to a given wild gene pool > 0.9) from 258 wild-wild admixed seedlings (i.e., seedlings with a membership coefficient to a given wild 259 apple cluster < 0.9, "*ww*", hereafter). Two effects were then tested using the statistical models 260 described below: the genetic status effect (i.e., dom, cw, ww, pure), and the wild apple population effect (i.e., corresponding to the "pure" populations detected with STRUCTURE). 261 A seedling that could not be assigned to any cluster (with a membership coefficient to any 262 263 cluster < 0.5) was removed for further analysis.

We computed descriptive population genetic estimates for each population (i.e., each cluster inferred with STRUCTURE, excluding admixed individuals with a membership coefficient < 0.9). Heterozygosity (expected and observed), Weir and Cockerham *F*-statistics, and deviations from Hardy–Weinberg equilibrium were calculated with Genepop v4.2 (Raymond and Rousset 1995; Rousset 2008). 269

#### 270 **Phenotypic traits and correlations**

Height, leaf growth rate, and carbon-related traits were estimated for each seedling. The absolute height and leaf growth rates (*AGR*, (Radford 1967)), relative growth rates (*RGR*,

273 (Briggs *et al.* 1920)), and whole *AGRs* were estimated as follows:

274 
$$AGR(cm/day) = \frac{(trait_{t+1} - trait_t)}{(date_{t+1} - date_t)}$$
(1)

275 
$$RGR(cm/day/day) = \frac{AGR}{date_t}$$
 (2)

276  $WholeAGR(cm/day) = \frac{trait_{end} - trait_{beginning}}{date_{end} - date_{beginning}}$ (3)

Note that for whole AGRs, the beginning of the experiment corresponded to days 7 and 11 for
leaf and height measurements, respectively, while the last measurement was performed on day
60. The *internode* ratio, which represents the ratio between the number of leaves and the height
of the seedling at day 60, was also considered a fitness trait, as this value plays an essential
role in apple tree architecture (Ripetti *et al.* 2008).

Seven phenotypic traits were therefore calculated for the entire dataset (565 seedlings, Table 1): *height\_AGR*, *height\_RGR*, *whole\_height\_AGR*, *leaf\_AGR*, *leaf\_RGR*, *whole\_leaf\_AGR*, and *internode*. In addition, chlorophyll (*Chl*) content, flavonol (*Flav*) content, and *NBI* were measured in a subsample of 257 seedlings (Table 1).

A preliminary exploration of the variation among phenotypic traits was performed. We used the cor R function to assess the correlation 1) between phenotypic traits and seed sizes (length, width, and weight) of the mother trees and 2) between phenotypic traits. The corrplot R package to visualize the correlations (Wei and Simko 2017). Principal component analyses of the phenotypic traits of the seedlings (PCA) were conducted with the FactoMineR R package (Lê *et al.* 2008).

292

# 293 Statistical analyses of phenotypic variation

A previous study demonstrated that crop-to-wild gene flow impacts early-stage growth rate (Feurtey *et al.* 2017). The effect of the genetic status of seedlings (*dom, cw, ww, pure*; Table 1) on fitness variation among seedlings was therefore tested. A linear mixed model was fitted to the data as follows:

298

299  $Y_{ijkl} \sim \mu + \text{wild population of origin}_i + \text{genetic status}_j + \text{wild population of origin}_i * \text{genetic status}_j$ 300  $+ \text{mother}_k + e_{ijkl}(4)$  302 Where  $Y_{iik}$  is the phenotypic trait (height or leaf growth rate or a carbon uptake-related trait) of 303 a seedling from mother k from population i with the genetic status j, wild population of origin 304 is the fixed effect of the population of origin *i* of the seedling inferred with STRUCTURE, 305 genetic status is the fixed effect of the genetic status of the seedling (i.e., pure, dom, ww, cw), 306 the interaction between the two fixed effects, mother tree is a normally distributed random 307 effect with its own mean and variance parameters and  $e_{ijkl}$  is the residual. The mother tree of 308 each seedling was used as a random factor to avoid pseudo-replication due to the presence of 309 multiple half-siblings (i.e., from the same mother tree). We ran the model (4), but replaced the 310 genetic status effect with the Pdom fixed effect. We gradually removed interactions and effects 311 depending on their significance. In addition, we evaluated the differences in the effect on trait 312 variation using a contrast analysis. We fitted the data to the model using the *lme4* R package 313 (Bates et al. 2015). The statistical significance of an effect was assessed with a type II Wald 314 chi-square test.

For phenotypic traits estimated from the number of leaves (*leaf\_AGR*, *leaf\_RGR* and *whole\_leaf\_AGR*), a log-link function was used and the residual distribution was fitted to a negative binomial distribution (function *glm.nb* in R package lme4). For phenotypic traits defined from the height of the seedling (*height\_AGR*, *height\_RGR*, *whole\_height\_AGR*), and for chlorophyll content, flavonol content, and NBI, a similar linear mixed model was run, but with a residual term that was assumed to be normally distributed.

321

## 322 Heritability estimates

The level of heritability estimate provides an indication of the potential extent of the response 323 324 to the selection of a trait. Therefore, computing heritability can allow us to determine whether 325 the traits measured in this study could be good candidates to respond to selection. Heritability 326 estimates were calculated using only pure and wild-wild M. sylvestris seedlings (Table 1). We fitted each fitness proxy with a linear mixed model as follows:  $Y_{ijk} = \mu + F_i + C_j + e_{ijk}$  (5), 327 where  $Y_{ijk}$  is the fitness proxy (growth rate or carbon uptake-related trait) of the k<sup>th</sup> seedling 328 from mother tree *i*, member of the  $j^{th}$  genetic cluster,  $\mu$  the overall fixed mean of the population, 329  $F_i$  the random effect of the *i*<sup>th</sup> mother tree,  $C_i$  is the fixed effect of the *j*<sup>th</sup> genetic cluster and  $e_{ijk}$ 330 331 the random error term. The model was fitted using REML (restricted maximum likelihood). 332 Calculations were performed by the *lme*-function of the *R*-library *nlme* (Pinheiro *et al.* 2022). 333 The output of *lme* provides estimates for the variance components, the corresponding standard

- deviations (sd), and the best unbiased linear predictors (BLUP) for random effects. Genetic
  parameters were then calculated as follows:
- 336 the additive genetic variance:  $VA = 4\sigma_F^2$  with  $\sigma_F^2$  representing the between-mother tree 337 variance
- 338 the corresponding coefficient of variation:  $CVA = \frac{\sqrt{VA}}{\mu}$
- 339 the phenotypic variation:  $VP = \sigma_F^2 + \sigma_E^2$ , with  $\sigma_E^2$  representing the residual variance
- 340 the corresponding coefficient of variation:  $CVP = \frac{\sqrt{VP}}{T}$
- 341 Narrow-sense heritability:  $h^2 = \frac{VA}{VP}$
- 342 Dickerson's approximation of standard deviation:  $sd(h^2) \approx \frac{4sd(\sigma_F^2)}{VP}$
- 343

@

# 344 Test for isolation-by-ecology

345 Only pure and wild-wild hybrid *M. sylvestris* seedlings were sampled for the IBE analysis (N 346 = 449, 21 sites, Table 1). The IBE pattern, i.e., the contribution of climate (isolation-by-347 climate) and phenotypic (isolation-by-adaptation) distances to the genetic structure, taking into 348 account geographical distance, was evaluated using distance-based redundancy analysis (db-349 RDA). Db-RDA can be used when the response variable is a distance matrix - here is a genetic 350 distance matrix  $(F_{ST})$  across 21 sampled sites - and the explanatory variables are in vector form 351 as follows. The geographical distance between sampled sites, estimated with SPAGeDI 1.5d 352 (Hardy and Vekemans 2002), that underlies an IBD process, was represented by vectors with 353 positive Eigenvalues of a principal coordinate of a neighbor matrix (PCNM) (Borcard and 354 Legendre 2002). A total of 19 bioclimatic variables, downloaded from the Worldclim2 355 database (30s resolution, https://www.worldclim.org/data/worldclim21.html) and representing 356 annual and seasonal trends and extremes averaged over the years 1970-2000 and averaged for 357 the Pleistocene period (20,000 years ago) (Gamisch 2019), were used to test for an isolation-358 by-climate pattern. Growth rate (N = 551, Table 1) and carbon uptake-related traits (N = 239, Table 1) averaged per site were used to test for an isolation-by-adaptation pattern, referred to 359 360 as IBA hereafter.

To identify the variables that explained the genetic structure of *M. sylvestris*, a db-RDA using the "capscale" function (Oksanen *et al.* 2014) was run on a complete model that included all investigated variables (i.e., PCNM components, growth rates, carbon uptake-related traits, and 19 bioclimatic variables). The best variables were selected for an optimum model with the function "step" based on the Akaike Information Criterion (AIC). Because db-RDA does not provide information on the relative contribution of each variable of the model, a variance
partitioning analysis was run using the "varpart" function from the R-package "vegan" (PeresNeto *et al.* 2006).

- 369
- 370 Results

### 371 Genetic status of seedlings

No clones or closely related individuals were detected (Figure S1); therefore, 584 seedlings
were included in the STRUCTURE analyses.

374 STRUCTURE revealed a clear spatial population genetic structure of M. sylvestris in Europe and crop-wild admixture (Figures 1 and S2). From K=2 to K=8, several clusters 375 376 appeared. When K > 8, STRUCTURE did not reveal any further substructures but only additional clusters with highly admixed individuals (Figure S2). Therefore, although the  $\Delta K$ 377 378 indicated that the most likely K value was five (Figure S3), K = 8 was the finest population 379 structure and was retained in subsequent analyses. For K = 8, we found that the *M. domestica* 380 reference varieties were admixed with Italian and Western French M. sylvestris. Note that 381 cultivars used as references originated from Western Europe (Table S2). Conversely, we 382 detected 68 *M. sylvestris* seedlings with  $P_{dom} > 0.1$  (considered to be *cw* hybrids), 383 corresponding to 11.6% of the seedlings (N = 584, Figures 1, S4, S5, S6, Table 1). We also 384 found 21 seedlings with a membership coefficient to the *M*. domestica gene pool > 0.9, 385 corresponding to 4% of the seedlings. Nearly all Spanish seedlings were assigned to the M. *domestica* gene pool with membership coefficients > 0.1 (i.e., 26 cw hybrids and 13 individuals 386 387 assigned to the *M. domestica* gene pool) and showed admixture only with the wild Italian 388 purple gene pool (Figures S4 and S6). A total of 33 individuals could not be assigned to any 389 cluster (i.e., individuals with a membership coefficient to any cluster < 0.5).

390 We, therefore, identified 68 cw, 21 dom, 167 ww, and 282 pure seedlings (Table 1, N = 551). 391 After removing *cw* hybrids (N = 68), seedlings sampled from misidentified mother trees (N =21), the *M. domestica* reference samples (N = 40), and individuals with a membership 392 coefficient to any cluster < 0.5, seven wild apple populations (i.e., groups of seedlings with a 393 394 membership coefficient > 0.5 to a wild apple cluster) were defined: French Western (FR-W395 hereafter, N = 77), French Eastern (*FR-E*, N = 50), French Lorraine (*FR-Lor*, N = 28), Danish 396 (DA, N = 78), Italian (IT, N = 27), Austrian (AUT, N = 81) and Romanian (RO, N = 108) (Figure 397 S6). Each *M. sylvestris* population exhibited a high level of genetic variation (Table S3). The 398 Romanian population was the most genetically differentiated and was close to the Austrian 399 population; the Danish and French Western populations were the closest genetically (Figure 400 S7), which is congruent with previous results from 26 microsatellite markers (Cornille et *al.*401 2015).

402

## 403 No effect of seedling genetic status on phenotypic variation

404 Variations and correlations among phenotypic traits are presented in Figures S8 to S12. 405 Significant correlations between seed length and weight of the mother tree and height of the 406 seedling were observed (*P*<0.01). Leaf growth rates (*leaf\_RGR*, *leaf\_AGR*, *leaf\_whole AGR*) 407 were significantly correlated, as were height growth rates (height\_RGR, height\_AGR, 408 *height\_whole\_AGR*). Chlorophyll content was positively correlated with NBI, whereas 409 flavonol content was negatively correlated with NBI. Heritability estimates were moderate to 410 high for all traits except growth rates based on leaf number (AGR\_leaf, RGR\_leaf, 411 whole\_AGR\_leaf; Table S4). Given the limited sample size, these estimates must be taken 412 cautiously, as reflected by the large standard deviations.

We did not find any significant effect of the genetic status of the seedlings (i.e., *pure*, *ww*, *cw*, *dom*) (Table S5, Figure S13) or  $P_{dom}$  (Table S6, Figure S14) on phenotypic traits, except a marginally significant effect (P = 0.04, Table S6) of  $P_{dom}$  on the whole height AGR: seedlings with higher levels of introgression by *M. domestica* had higher whole height AGRs. We, therefore, removed the seedling genetic status and *Pdom* effects from the model 4, as well as *dom* and *cw* individuals. Thus, the final model only included wild apple seedlings (i.e., *pure* and *ww*, N = 449) and focused on the *wild population of origin* effect (Table 2).

420

421 Significant variation in growth rates and chlorophyll content among populations 422 The mean height variation during the experiment among seedlings from different populations 423 is shown in Figure 2. There was significant variation among seedlings from other populations 424 in certain growth-related traits (Table 2). On average, seedlings belonging to the Austrian 425 population were taller (+11 cm, P = 0.047) whereas Romanian (-14.9 cm, P = 0.008) and Italian 426 (-18.7 cm, P = 0.044) seedlings were shorter (Figures 2 and S15) than seedlings from other 427 populations. Seedlings from other populations did not show a significant height difference. In 428 addition, the number of leaves and height traits were negatively correlated, r = -0.3, P < 0.001). 429 The Austrian population presented the lowest number of leaves (average = 5, sd = 4), whereas 430 seedlings belonging to the Romanian population had the highest number of leaves (average = 431 8, sd = 7, Figure S16). The Romanian population also had the largest *internode* (+0.02 leaf/cm,432 P = 0.024).

433 Chlorophyll content differed among populations, with seedlings from the Italian population 434 producing, on average, more chlorophyll (+4.14  $\mu$ g/cm<sup>2</sup>, *P* = 0.039, Figure S17) than seedlings 435 from other populations. Flavonol content and NBI did not differ significantly among 436 populations.

437

# 438 Significant IBD and IBC

439 Correlation plots between bioclimatic variables are provided in Figures S18 and S19; however, 440 all variables were included in the analysis, as db-RDA can cope with correlated variables. The 441 optimal model was chosen according to its best AIC value. The optimal model explained up to 442 25.9% of the genetic structure (Adj- $R^2$  = 69.9%, P = 0.001) and contained seven variables (four 443 geographic and three bioclimatic variables) (Table 3): the geographical distance is represented 444 by the 1st, 2nd, 3rd and 6th axis of the PCNM analysis and three past climatic variables (Bio3: 445 isothermality; Bio6: minimum temperature of the coldest month; Bio 9: mean temperature of the driest quarter). In total, IBD explained 47% of the variance of the wild apple tree population 446 447 genetic structure, whereas IBC explained 22% (Figure S20). Taking geographical distance into 448 account, we did not find a pattern of IBA, i.e., covariation between phenotype and genetic 449 divergences.

450

### 451 **Discussion**

452 This study assessed the relationship between phenotypic and genetic variation among 453 populations of a wild contributor to the cultivated apple genome (Cornille et al. 2012), M. 454 sylvestris, a CWR species that occurs naturally along a climatic gradient in Europe. Bayesian 455 clustering revealed a substantial number of seedlings introgressed by *M. domestica*. With the 456 hybrids removed, seven populations of *M. sylvestris* distributed across Europe were detected 457 and showed phenotypic variation in growth and chlorophyll content. Based on the IBA pattern 458 estimated from the phenotypic traits measured in this study, this phenotypic variation was not 459 adaptive. However, the significant association between population genetic variation and the LGM climate suggests that the European crabapple may be locally adapted to the past climate 460 461 conditions of the LGM. The results of this study, therefore, indicate the occurrence of adaptive 462 divergence related to climate in a wild contributor to the cultivated apple genome. This may 463 help us to utilize its diversity better, providing options for mitigating the impact of climate 464 change on the cultivated apple through breeding (Warschefsky et al. 2014; Prohens et al. 2017; 465 Satori et al. 2022).

466

#### 467 **Ongoing crop-to-wild gene flow in the European crabapple**

We showed substantial gene flow from *M. domestica* to the European crabapple gene pool, 468 469 with 11.6% of seedlings, mainly from Western Europe, introgressed by M. domestica. 470 Introgression rates were lower compared to previous studies (i.e., 37% in Cornille et al. (2013b) 471 and 23.1% in Cornille et al. (2015)). However, these studies genotyped more mother trees (i.e., 472 N = 756 and N = 1,889, respectively), which could explain the difference in estimates of crop-473 to-wild gene flow. Crop-to-wild gene flow is therefore still ongoing in the European crabapple. 474 The higher number of seedlings from Western Europe that are introgressed by *M. domestica* 475 can be explained by the use of reference Western European cultivated apple varieties. Alleles 476 specific to Eastern and Northern cultivated apple varieties can be missed and may decrease the 477 probability of detecting crop-to-wild introgression events in non-Western wild populations (from Eastern and Northern Europe). Note that the Spanish seedlings sampled in this study 478 479 were the progeny of trees growing in a location known to have high levels of introgression by 480 *M. domestica* (pers. comment. G. Alins). It is even possible that the mother trees of these 481 seedlings were *M. domestica* and not *M. sylvestris*.

The consequences of crop-wild introgression on phenotypic variation between crop and wild individuals are poorly understood in perennial fruit trees. One study has shown that cropwild hybrid apple seedlings have higher growth rates and germinate earlier than wild apple seedlings (Feurtey *et al.*, 2017). We did not detect any effect of the genetic status of a seedling (*pure*, *ww*, *cw*, *dom*) or the level of introgression (*Pdom*) on growth and carbon uptake-related fitness proxies. This could be due to the low number of samples from the *cw* and *dom* categories.

489

(a)

# 490 Signs of local adaptation to past climate in the European crabapple

491 Under controlled conditions, seedlings from different populations exhibited significantly 492 different growth and physiology. Seedlings belonging to the Austrian population were the 493 tallest, with the highest absolute growth rate and the lowest number of leaves; by contrast, 494 Romanian seedlings were the shortest and had the lowest absolute growth rate and the highest 495 number of leaves. Italian seedlings had the highest chlorophyll content. We therefore tested 496 whether this phenotypic variation was adaptive. For instance, the seedlings from the Austrian 497 population may be fitter in the climate conditions simulated in this controlled condition 498 experiment. However, considering the geographic distance, we found no significant 499 covariation between genetic and phenotypic variation. This suggests a lack of divergent 500 selection on carbon uptake or growth traits, which are yet often associated with plant responses

501 to climate (Bussotti et al. 2015). Therefore, the phenotypic variation we observed among 502 populations under controlled conditions may result from genetic drift alone and not from 503 divergent selection. Alternatively, although the specific traits we selected are among the traits 504 that are generally considered to be related to responses to climate (Kühn et al. 2021), they 505 might not be perfect candidates for investigating divergent selection by climate. Another 506 explanation for the lack of observation of divergent selection on carbon uptake or growth traits 507 could be that we did not phenotype enough seedlings from each genetic group. Indeed, we 508 observed a high variation in each phenotypic trait and their heritability estimates, suggesting 509 that the traits we studied may be relevant but that a larger number of seedlings could be 510 phenotyped and analyzed. However, some studies have found that even with large sample sizes, 511 the standard error of heritability estimates can still be large and vary significantly between 512 experimental designs (Visscher and Goddard 2015). The reasonably high heritability estimates 513 for most traits considered here could be consistent with relatively weak within-population 514 selection, enabling the maintenance of ample additive genetic variation (Wheelwright et al. 515 2014). Furthermore, high variation in seedling traits combined with high heritability estimates 516 could suggest a substantial amount of genetic variation for adaptation to work on. Altogether, 517 the effect of the population of origin on phenotypic traits suggests a genetic basis for this 518 variation. However, we need to increase the sampling size and measure new traits to draw more 519 precise conclusions on the occurrence of adaptive phenotypic variation in the European 520 crabapple.

As M. sylvestris is distributed across various climatic conditions, we further 521 522 investigated the role of climate in shaping the genetic variation among populations of the 523 European crabapple without considering phenotypic trait variation. While the association 524 between phenotypic and genetic variation was not significant (see paragraph above), we 525 observed a significant association between LGM climate and genetic variation in the European 526 crabapple suggesting local adaptation. We tested for an IBE pattern, where the pattern of 527 neutral genetic variation covaries with ecological variables (here, climate). There was no combined effect of geographic and climatic distance (IBD  $\cap$  IBC), which allowed us to assess 528 529 the contribution of these processes separately (Wang and Bradburd 2014). We showed that IBD and IBC played a significant role ( $R^2_{adj} = 47\%$  and  $R^2_{adj} = 22\%$ , respectively) on the genetic 530 531 differentiation of European crabapple populations. Weak but significant IBD has been 532 previously identified in wild apple relatives of the cultivated apple (i.e., M. sylvestris, M. 533 orientalis, and M. sieversii) (Cornille, Giraud, et al. 2013; Cornille, Gladieux, et al. 2013; 534 Cornille et al. 2015). Here we used 13 out of the 26 microsatellite markers used in previous

535 studies; the lack of resolution of the 13 SSRs may explain the lack of IBD. However, this is 536 unlikely as the Bayesian inferences method was previously able to detect genetic clusters in 537 wild apples (Cornille, Giraud, et al. 2013; Cornille, Gladieux, et al. 2013; Cornille et al. 2015). 538 Weak IBD pattern suggests that *M. sylvestris* has high dispersal capacities (Coart *et al.* 2006; 539 Larsen et al. 2006; Cornille, Giraud, et al. 2013; Cornille et al. 2015; Reim et al. 2017; Feurtey 540 et al. 2017). A weak IBD is explained by a self-incompatibility system that prevents self-541 fertilization (Brown 1992), pollen dispersal by bees, beetles, and flies, and endozoochorous 542 seed dispersal by large mammals such as ungulates, wild boars, brown bears or humans (Larsen 543 et al. 2006).

544 We show that in addition to IBD, IBC persisted after considering the geographical 545 distance. Climate can impose divergent selection pressures on different locations and thus 546 reduce gene flow between populations. For instance, divergent selection imposed by climate 547 can limit the reproductive success of individuals moving between different climates from 548 which they are adapted, so that IBC contributes to genetic differentiation among populations 549 (Wang and Bradburd 2014). The main variables explaining genetic differentiation in the 550 European crabapple were related to temperature during the LGM. This suggests that the 551 European crabapple may be adapted to its past climate but not to its current one. In wind-552 dispersed trees, local adaptation to the current climate has been demonstrated (Savolainen et 553 al. 2013; Kremer and Hipp 2019; Pyhäjärvi et al. 2020), but to our knowledge, no study has 554 shown local adaptation to past climate conditions in an insect-pollinated tree species.

555 Factors other than climate can also shape the adaptive divergence between populations. 556 Local adaptation to biotic factors, such as the presence of other species, is possible. Malus 557 sylvestris is a species that needs high levels of light and is not very competitive. Some of the 558 observed variations of the seedlings in growth rate, leaf number, and chlorophyll content may 559 derive from adaptations that would be advantageous within the local niche. In Romania, the 560 populations sampled were in forest edges, middle succession woodlands (with hawthorn and 561 wild pear), or grasslands; no wild apples were found in mature forests. There, M. sylvestris does not compete with other woody species but with grasses and shrubs in the seedling phase; 562 563 this might explain the slower growth in height and the larger number of leaves (leading to a 564 larger leaf area and more shading of the competing grasses). In France, the trees are present at 565 the edge of mature forests, and in Austria, in mature forests. It is hard to draw clear conclusions; 566 further investigations of the ecology of the wild populations *in situ* are needed. The rhizosphere 567 composition among seedlings from different populations can impact the phenotypic variation 568 and potentially plant fitness in response to climate (Trivedi et al. 2022). Here seeds were

569 cleaned with chlorine to avoid the effect of local micro-organism community and seed growth.
570 Still, investigations of phenotypic variation among cleaned and uncleaned seeds can also help
571 assess the role of the rhizosphere in the divergence of the wild apple population. Local
572 adaptation of fruit trees to biotic factors, including parasites (Olvera-Vazquez *et al.* 2021) also
573 deserves further investigation. Besides selection, the role of phenotypic plasticity in enabling
574 growth and optimal fitness in changing environments also needs to be carefully evaluated
575 (Benito Garzón *et al.* 2011).

576

# 577 Further investigations are needed on local adaptation and phenotypic plasticity in 578 response to climate in the European crabapple

579 Our study raises questions regarding the response of wild apple populations to climate change. 580 More data is needed to draw clear conclusions. The adaptation of tree species to climate is 581 complex (Bussotti et al. 2015). For instance, in Eucalyptus camaldulensis, variation in leaf 582 traits and performance was unrelated to the climate of genotype provenance (Asao *et al.*, 2020), 583 while in Australia, the same species displays variation in several photosynthetic traits that were 584 related to the climate of genotype provenance (Dillon *et al.* 2018). By contrast, collective 585 differences in leaf morphology and photosynthetic physiology associated with the length of the 586 growing season, temperature, and the level of insolation in several *Populus* species may be 587 adaptive (Keller et al. 2011; Kaluthota et al. 2015). Further investigations on local adaptation 588 and phenotypic plasticity in response to climate in the European crabapple are needed. 589 Genomic data will help determine the relative influence of adaptive and neutral processes on 590 climate-driven divergence by scanning the genomes of trees from different populations in 591 Europe. Comparing the fitness of seedlings from different populations in reciprocal transplants 592 (under controlled or natural conditions) will also allow us to investigate local adaptation and 593 phenotypic plasticity. Further studies using additional genetic markers (single nucleotide 594 polymorphism) and measuring phenotypic traits in different climate conditions are therefore 595 needed. This study is nevertheless a starting point for future breeding and conservation 596 programs of a CWR of an emblematic temperate fruit tree, as it characterized the phenotypic 597 and genetic variation of seedlings in the wild that can be used as *ex-situ* sources to enrich the 598 crop gene pool. Indeed, some of the seedlings included have sequenced genomes and have been 599 planted in several orchards in France (France, https://www.ideev.universite-paris-600 saclay.fr/en/the-orchard) and are measured each year for several phenotypic traits. Information 601 and samples from these orchards can be requested from the corresponding author. Genetic 602 variation of trees in these *ex-situ* orchards can help enrich the cultivated apple gene pool.

- 603 Certain traits related to climate change adaptation can be introduced into future apple varieties
- 604 (Warschefsky et al. 2014; Prohens et al. 2017; Satori et al. 2022).
- 605

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# 622 Authors contributions

AC, GV, AR, SB, and TU conceived and designed the experiments; AC, GV, AT, TU, and TK
obtained the funding; AC, GV, AT, TU, KAO, SV, RR, XC, TK, CR sampled the material;
XC, CR, AV, AR, GL, KAO, RR, MLG, HB, VC, HC, SV, MF performed the molecular
biology analyses; AC, AF, KomAvi and XC analyzed the data. The manuscript was written by
AC, KomAvi and AF, with essential input from other co-authors.

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### **Figures and Tables**

Figure 1. Bayesian clustering of *Malus sylvestris* seedlings sampled in this study (N = 584) and reference samples of *Malus domestica* (N = 40) inferred with STRUCTURE at K = 8 and its associated map of mean membership per sampled site. Each individual is represented by a vertical bar partitioned into clusters. Visualization was improved by sorting genotypes by country. Countries are separated by a white line. The *M. domestica* reference samples are shown on the far left of the map in the Atlantic. Circle size is proportional to the number of individuals within the cluster (scale shown in the top right-hand corner).

Figure 2. Cumulative height growth over time in apple seedlings under controlled conditions. Seedlings measured were pure and wild-wild hybrid *Malus sylvestris* seedlings, N = 449, and seedlings assigned to the *M. domestica* gene pool, N = 21, as detected with STRUCTURE for *K*=8. The 40 reference *M. domestica* individuals were not measured under controlled conditions and are therefore not shown here. Vertical lines represent the standard deviation. Populations: Austria (AUT, N = 81), Denmark (DA, N = 78), *M. domestica* (DOM, N = 21), which includes 13 Spanish genotypes and seedlings from other countries), Eastern France (FR-E, N = 50), Lorraine in France (FR-Lor, N = 28), Western France (FR-W, N = 77), Italy (IT, N = 27) and Romania (RO, N = 108).

Table 1. Number of *Malus sylvestris* seedlings used in this study for population genetic analyses inferred with STRUCTURE for K = 8, with 13 microsatellite markers and phenotyping (growth and carbon uptake-related traits).

Clusters	N <sub>pure</sub>	$N_{ww}$	$N_{cw}$	N <sub>dom</sub>	Nno cluster	Total measured for phenotypic traits	Population name
Q1 (light green)	32	46	7	0	10	92	FR-W
Q2 (yellow)	0	52	5	0	4	57	FR-E
Q3 (lor)	28	1	0	0		28	FR-Lor
Q4 (blue)	61	21	1	0	6	85	DA
Q5 (purple)	23	4	3	0	4	34	IT
Q6 (dark green)	66	17	1	0	1	83	AUT
Q7 (red)	77	34	5	0	0	113	RO
Q8 (black – M. domestica)	<mark>40</mark>	<mark>0</mark>	<mark>46</mark>	<mark>21</mark>	8	73	DOM
Total	287	175	68	21	33	551 (	(584)

Total measured for height and number of leaves	282	167	63	21	32	533 (565)
Total measured for leaf chlorophyll and flavonol contents, NBI	129	82	22	6	18	239 (257)

 $N_{pure}$ : the number of seedlings assigned to a wild gene pool with a membership coefficient > 0.9;  $N_{ww}$ : the number of wild-wild hybrids (i.e., seedlings with a membership coefficient > 0.1 to a wild gene pool other than its own wild gene pool and a membership coefficient < 0.1 to the *M*. *domestica* gene pool);  $N_{cw}$ : the number of crop-wild hybrids (i.e., seedlings assigned to the *M*. *domestica* gene pool with a membership coefficient > 0.1).  $N_{no\ cluster}$ : seedlings that could not be assigned to any defined gene pool; Total measured for phenotypic traits : the number of individuals measured for each phenotypic trait and included in the statistical analyses, the number in brackets represents the initial sample size before data were filtered for statistical analyses. Wild population name: populations defined with STRUCTURE for *K*=8 excluding crop-wild hybrids and seedlings from misidentified mother trees (i.e., including only wild pure and wild-wild hybrids). Populations: Austria (AUT), Denmark (DA), Eastern France (FR-E), Lorraine in France (FR-Lor), Western France (FR-W), Italy (IT), Romania (RO), *M. domestica* (DOM).

Table 2. Final model depicting the effects of the *Malus sylvestris* population to which each seedling belonged (i.e., cluster inferred with STRUCTURE for *K*=8) on phenotypic traits (*i.e.*, height, number of leaves, internode, chlorophyll and flavonol contents, NBI) measured on 533 individuals.

Explanatory variable	Population			Mother tree			Mod	Mother tree	
Fitness	X <sup>2</sup>	P-value	df	REML	Standard Deviatio n	AIC	R²	Corrected R <sup>2</sup>	R²
Height_AGR	17.863	0.007** *	6	1,229	0.204	1,248	0.047	0.091	0.044
Height_RGR	12.846	0.045*	6	-2,264	0.006	- 2,245	0.041	0.147	0.106
Leaf_AGR	4.302	0.6359	6	134	0.036	152	0.01	0.028	0.018
Leaf_RGR	4.302	0.6359	6	134	0.036	152	0.01	0.028	0.018
Whole height AGR	22.243	1.00e- 03***	6	630	0.175	650	0.074	0.192	0.118

Whole leaf AGR	36.326	2.38e- 06***	6	-1,277	0.009	- 1,258	0.09	0.118	0.028
Height	31.623	1.93e- 05***	6	4,113	11.69	4,131	0.119	0.301	0.182
Number of leaves	22.285	0.001** *	6	-	0.084	2,659	0.052	0.064	0.012
Chlorophyll	14.418	0.025*	6	1,181	1.352	1,199	0.074	0.171	0.097
Flavonol	6.7752	0.342	6	-124	0.091	-105	0.043	0.299	0.256
NBI	1.838	0.934	6	1,735	7.6617	1,754	0.011	0.224	0.213
Internode (nbleaf/heigh t)	17.768	0.007** *	6	-1,328	0.009	- 1,309	0.044	0.073	0.029

\*\*\*: P-value <0.001; \*\*: 0.01< P-value <0.001; \*: 0.05< P-value <0.01; AIC: Akaike Information Criterion; - Models without any significant effect.

 Table 3. Contribution of geography and climate to the genetic variation observed among *Malus sylvestris* seedlings. Distance-based redundancy analyses tested the effects of geography, climate, and phenotype on the genetic differentiation (from 13 microsatellites) among 21 sites in the European crabapple. Only the significant variables are listed.

	db-RDA						
	% of variance explained	d.f.	p-value	Adj-R <sup>2</sup>			
Global analysis	25.9	7	0.001				
Residuals	11.3	13	-				
Geography (IBD, PCNM 1-2-3-6)	14.9	4	< 0.015	<i>c</i> 0 0			
Environment (IBC_LGM: BIO3_LGM, BIO6_LGM, BIO9_LGM)	11.04	3	<0.015	69.9			
Residuals	11.3%	-	-				

BIO3\_LGM: isothermality (BIO2/BIO7) (×100)); BIO6\_LGM: minimum temperature of the coldest month; BIO9\_LGM: mean temperature of the driest quarter; IBD: isolation-by-distance; IBC\_LGM: isolation-by-climate during the last glacial maximum.

# References

Aitken SN, Bemmels JB. 2015. Time to get moving: assisted gene flow of forest trees. *Evolutionary applications* **9**: 271–290.

Alexandre H, Truffaut L, Klein E, *et al.* 2020. How does contemporary selection shape oak phenotypes? *Evolutionary Applications* 13: 2772–2790.

Aubin I, Munson AD, Cardou F, *et al.* 2016. Traits to stay, traits to move: a review of functional traits to assess sensitivity and adaptive capacity of temperate and boreal trees to climate change. *Environmental Reviews* 24: 164–186.

**Bates D, Mächler M, Bolker B, Walker S**. **2015**. Fitting linear mixed-effects models using lme4. *Journal of Statistical Software, Articles* **67**: 1–48.

**Benito Garzón M, Alía R, Robson TM, Zavala MA**. **2011**. Intra-specific variability and plasticity influence potential tree species distributions under climate change. *Global Ecology and Biogeography* **20**: 766–778.

**Borcard D, Legendre P. 2002**. All-scale spatial analysis of ecological data by means of principal coordinates of neighbour matrices. *Ecological Modelling* **153**: 51–68.

**Boyd IL, Freer-Smith PH, Gilligan CA, Godfray HCJ. 2013**. The Consequence of Tree Pests and Diseases for Ecosystem Services. *Science* **342**.

**Brachi B, Villoutreix R, Faure N, et al. 2013**. Investigation of the geographical scale of adaptive phenological variation and its underlying genetics in Arabidopsis thaliana. *Molecular Ecology* **22**: 4222–4240.

Briggs AW, Kidd F, West C. 1920. A quantitative analyses of plant growth: part II. *Annals of Applied Biology* 7: 202–223.

**Briscoe Runquist RD, Gorton AJ, Yoder JB**, *et al.* **2020**. Context Dependence of Local Adaptation to Abiotic and Biotic Environments: A Quantitative and Qualitative Synthesis. *The American Naturalist* **195**: 412–431.

**Brown SK**. **1992**. Genetics of apple In: *Plant breeding reviews vol 9*. John Wiley & Sons New York, 333–366.

**Bussotti F, Pollastrini M, Holland V, Brüggemann W**. **2015**. Functional traits and adaptive capacity of European forests to climate change. *Environmental and Experimental Botany* **111**: 91–113.

**Coart E, Van Glabeke S, De Loose M, Larsen AS, Roldán-Ruiz I**. **2006**. Chloroplast diversity in the genus Malus: new insights into the relationship between the European wild apple (Malus sylvestris (L.) Mill.) and the domesticated apple (Malus domestica Borkh.). *Molecular Ecology* **15**: 2171–2182.

**Comes HP, Kadereit JW**. **1998**. The effect of Quaternary climatic changes on plant distribution and evolution. *Trends in Plant Science* **3**: 432–438.

Cornille A, Antolín F, Garcia E, *et al.* 2019. A Multifaceted Overview of Apple Tree Domestication. *Trends in Plant Science* 24: 770–782.

**Cornille A, Feurtey A, Gélin U,** *et al.* **2015**. Anthropogenic and natural drivers of gene flow in a temperate wild fruit tree: A basis for conservation and breeding programs in apples. *Evolutionary Applications* **8**: 373–384.

**Cornille A, Giraud T, Bellard C,** *et al.* **2013**. Post-glacial recolonization history of the European crabapple (Malus sylvestris Mill.), a wild contributor to the domesticated apple. *Molecular Ecology* **22**: 2249–63.

**Cornille A, Giraud T, Smulders MJM, Roldán-Ruiz I, Gladieux P. 2014**. The domestication and evolutionary ecology of apples. *Trends in Genetics* **30**: 57–65.

**Cornille A, Gladieux P, Giraud T**. **2013**. Crop-to-wild gene flow and spatial genetic structure in the wild closest relatives of the cultivated apple. *Evolutionary Applications* **6**: 737–748.

**Cornille A, Gladieux P, Smulders MJM**, *et al.* **2012**. New insight into the history of domesticated apple: secondary contribution of the European wild apple to the genome of cultivated varieties. *PLoS Genet* **8**: e1002703.

**Delplancke M, Alvarez N, Espíndola A, et al. 2011**. Gene flow among wild and domesticated almond species: insights from chloroplast and nuclear markers. *Evolutionary Applications* **5**: 317-329.

Demestihas C, Plénet D, Génard M, Raynal C, Lescourret F. 2017. Ecosystem services in orchards. A review. *Agronomy for Sustainable Development* 37: 12.

**Demotes-Mainard S, Boumaza R, Meyer S, Cerovic ZG**. **2008**. Indicators of nitrogen status for ornamental woody plants based on optical measurements of leaf epidermal polyphenol and chlorophyll contents. *Scientia Horticulturae* **115**: 377–385.

**Diez CM, Trujillo I, Martinez-Urdiroz N, et al. 2015**. Olive domestication and diversification in the Mediterranean Basin. *New Phytologist* **206**: 436–447.

**Dillon S, Quentin A, Ivković M, Furbank RT, Pinkard E**. **2018**. Photosynthetic variation and responsiveness to CO2 in a widespread riparian tree. *PLOS ONE* **13**: e0189635.

**Dusenge ME, Duarte AG, Way DA**. **2019**. Plant carbon metabolism and climate change: elevated CO2 and temperature impacts on photosynthesis, photorespiration and respiration. *New Phytologist* **221**: 32–49.

**Earl DA, vonHoldt BM. 2012.** STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources* **4**: 359–361.

**Edwards SV, Robin VV, Ferrand N, Moritz C**. **2022**. The Evolution of Comparative Phylogeography: Putting the Geography (and More) into Comparative Population Genomics. *Genome Biology and Evolution* **14**: evab176.

**Evanno G, Regnaut S, Goudet J**. **2005**. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology* **14**: 2611–2620.

**Feurtey A, Cornille A, Shykoff JA, Snirc A, Giraud T**. **2017**. Crop-to-wild gene flow and its fitness consequences for a wild fruit tree: Towards a comprehensive conservation strategy of the wild apple in Europe. *Evolutionary Applications* **10**: 180–188.

Flowers JM, Hazzouri KM, Gros-Balthazard M, *et al.* 2019. Cross-species hybridization and the origin of North African date palms. *Proceedings of the National Academy of Sciences* 116: 1651–1658.

**Francis R**. **2016**. POPHELPER: An R package and web app to analyse and visualise population structure. *Molecular Ecology Resources* **17**: n/a-n/a.

**Franks SJ, Weber JJ, Aitken SN. 2014**. Evolutionary and plastic responses to climate change in terrestrial plant populations. *Evolutionary Applications* **7**: 123–139.

**Gamisch A**. **2019**. Oscillayers: A dataset for the study of climatic oscillations over Plio-Pleistocene time-scales at high spatial-temporal resolution. *Global Ecology and Biogeography* **28**: 1552–1560.

Hamann FA, Czaja S, Hunsche M, Noga G, Fiebig A. 2018. Monitoring physiological and biochemical responses of two apple cultivars to water supply regimes with non-destructive fluorescence sensors. *Scientia Horticulturae* 242: 51–61.

Hardy OJ, Vekemans X. 2002. SPAGeDI: a versatile computer program to analyse spatial genetic structure at the individual or population levels. *Molecular Ecology Notes* 2: 618–620.

Hartmann H, Bahn M, Carbone M, Richardson AD. 2020. Plant carbon allocation in a changing world – challenges and progress: introduction to a Virtual Issue on carbon allocation. *New Phytologist* 227: 981–988.

Hoban S, Volk G, Routson KJ, Walters C, Richards C. 2018. Sampling Wild Species to Conserve Genetic Diversity In: Greene SL, Williams KA, Khoury CK, Kantar MB, Marek LF, eds. *North American Crop Wild Relatives, Volume 1: Conservation Strategies*. Cham: Springer International Publishing, 209–228.

Hübner S, Kantar MB. 2021. Tapping Diversity From the Wild: From Sampling to Implementation. *Frontiers in Plant Science* 12: 38.

Hutyra LR, Munger JW, Saleska SR, *et al.* 2007. Seasonal controls on the exchange of carbon and water in an Amazonian rain forest. *Journal of Geophysical Research: Biogeosciences* 112.

Kaluthota S, Pearce DW, Evans LM, Letts MG, Whitham TG, Rood SB. 2015. Higher photosynthetic capacity from higher latitude: foliar characteristics and gas exchange of southern, central and northern populations of Populus angustifolia. *Tree Physiology* **35**: 936–948.

Keller SR, Soolanayakanahally RY, Guy RD, Silim SN, Olson MS, Tiffin P. 2011. Climate-driven local adaptation of ecophysiology and phenology in balsam poplar, Populus balsamifera L. (Salicaceae). *American Journal of Botany* **98**: 99–108. **Kopelman NM, Mayzel J, Jakobsson M, Rosenberg NA, Mayrose I**. **2015**. Clumpak: a program for identifying clustering modes and packaging population structure inferences across K. *Molecular Ecology Resources* **15**: 1179–1191.

Kremer A, Hipp AL. 2019. Oaks: an evolutionary success story. *New Phytologist* 226: 943–946.

**Kremer A, Kleinschmit J, Cottrell J,** *et al.* **2002**. Is there a correlation between chloroplastic and nuclear divergence, or what are the roles of history and selection on genetic diversity in European oaks? *Range wide distribution of chloroplast DNA diversity and pollen deposits in European white oaks: inferences about colonisation routes and management of oak genetic resources*. **156**: 75–87.

Kühn N, Tovar C, Carretero J, Vandvik V, Enquist BJ, Willis KJ. 2021. Globally important plant functional traits for coping with climate change. Frontiers of Biogeography. *Frontiers of Biogeography* 13.

Lander TA, Klein EK, Roig A, Oddou-Muratorio S. 2021. Weak founder effects but significant spatial genetic imprint of recent contraction and expansion of European beech populations. *Heredity* **126**: 491–504.

Larsen A, Asmussen C, Coart E, Olrik D, Kjær E. 2006. Hybridization and genetic variation in Danish populations of European crab apple (Malus sylvestris). *Tree Genetics & Genomes* 2: 86–97.

Lê S, Josse J, Husson F. 2008. FactoMineR: An R Package for Multivariate Analysis. *Journal of Statistical Software* 25: 1–18.

Li J, Li H, Jakobsson M, Li SEN, Sjödin PER, Lascoux M. 2012. Joint analysis of demography and selection in population genetics: where do we stand and where could we go? *Molecular Ecology* 21: 28–44.

Liu S, Cornille A, Decroocq S, *et al.* 2019. The complex evolutionary history of apricots: species divergence, gene flow and multiple domestication events. *Molecular Ecology* 28: 5299–5314.

Loiselle BA, Sork VL, Nason J, Graham C. 1995. Spatial genetic structure of a tropical understory shrub, Psychotria officinalis (Rubiaceae). *American Journal of Botany* 82: 1420–1425.

Mercado LM, Medlyn BE, Huntingford C, *et al.* 2018. Large sensitivity in land carbon storage due to geographical and temporal variation in the thermal response of photosynthetic capacity. *New Phytologist* 218: 1462–1477.

**Migicovsky Z, Gardner KM, Richards C,** *et al.* **2021**. Genomic consequences of apple improvement. *Horticulture Research* **8**: 1–13.

Myles S, Boyko AR, Owens CL, *et al.* 2011. Genetic structure and domestication history of the grape. *PNAS* 108: 3530–3535.

Nicotra AB, Atkin OK, Bonser SP, *et al.* 2010. Plant phenotypic plasticity in a changing climate. *Trends in Plant Science* 15: 684–692.

Nosil P, Funk DJ, Ortiz-barrientos D. 2009. Divergent selection and heterogeneous genomic divergence. *Molecular Ecology* 18: 375–402.

Oksanen J, Blanchet FG, Kindt R, et al. 2014. Vegan: Community ecology package.

**Olvera-Vazquez SG, Alhmedi A, Miñarro M, et al. 2021**. Experimental test for local adaptation of the rosy apple aphid (Dysaphis plantaginea) to its host (Malus domestica) and to its climate in Europe. *PCI Ecology* **Pre-registration version**.

**Orsini L, Vanoverbeke J, Swillen I, Mergeay J, De Meester L. 2013**. Drivers of population genetic differentiation in the wild: isolation by dispersal limitation, isolation by adaptation and isolation by colonization. *Molecular Ecology* **22**: 5983–5999.

**Parisod C. 2021**. Plant speciation in the face of recurrent climate changes in the Alps. *Alpine Botany*.

**Pasho E, Julio Camarero J, Vicente-Serrano SM**. **2012**. Climatic impacts and drought control of radial growth and seasonal wood formation in Pinus halepensis. *Trees* **26**: 1875–1886.

Peace CP, Bianco L, Troggio M, *et al.* 2019. Apple whole genome sequences: recent advances and new prospects. *Horticulture Research* 6: 59.

**Peres-Neto PR, Legendre P, Dray S, Borcard D**. **2006**. Variation partitioning of species data matrices: estimation and comparison of fractions. *Ecology* **87**: 2614–2625.

Petit RJ, Hampe A. 2011. Some evolutionary consequences of being a tree. *Annu. Rev. Ecol. Evol. Syst.* 37: 187–214.

**Pinheiro J, Bates D, R Core Team**. **2022**. *nlme: Linear and Nonlinear Mixed Effects Models*.

Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155: 945–959.

Prohens J, Gramazio P, Plazas M, *et al.* 2017. Introgressiomics: a new approach for using crop wild relatives in breeding for adaptation to climate change. *Euphytica* 213: 158.

**Puechmaille SJ. 2016**. The program structure does not reliably recover the correct population structure when sampling is uneven: subsampling and new estimators alleviate the problem. *Molecular Ecology Resources* **16**: 608–627.

**Pyhäjärvi T, Kujala ST, Savolainen O**. **2020**. 275 years of forestry meets genomics in Pinus sylvestris. *Evolutionary Applications* **13**.

**Pyhäjärvi T, Salmela MJ, Savolainen O**. **2008**. Colonization routes of Pinus sylvestris inferred from distribution of mitochondrial DNA variation. *Tree Genetics & Genomes* **4**: 247–254.

**Radford PJ**. **1967**. Growth Analysis Formulae - Their Use and Abuse1. *Crop Science* **7**: cropsci1967.0011183X000700030001x.

**Ramírez-Valiente JA, Center A, Sparks JP, et al. 2017**. Population-Level Differentiation in Growth Rates and Leaf Traits in Seedlings of the Neotropical Live Oak Quercus oleoides Grown under Natural and Manipulated Precipitation Regimes. *Frontiers in Plant Science* **8**: 585.

**Raymond M, Rousset F. 1995**. GENEPOP (version 1.2): population genetics software for exact tests and ecumenicism. *Journal of Heredity* **86**: 248–249.

**Reim S, Proft A, Heinz S, et al. 2017**. Pollen movement in a Malus sylvestris population and conclusions for conservation measures. *Plant Genetic Resources* **15**: 12–20.

**Riordan EC, Gugger PF, Ortego J, et al. 2016**. Association of genetic and phenotypic variability with geography and climate in three southern California oaks. *American Journal of Botany* **103**: 73–85.

**Ripetti V, Escoute J, Verdeil JL, Costes E**. **2008**. Shaping the shoot: the relative contribution of cell number and cell shape to variations in internode length between parent and hybrid apple trees. *Journal of Experimental Botany* **59**: 1399–1407.

**Rousset F**. **2008**. Genepop'007: a complete re-implementation of the genepop software for Windows and Linux. *Molecular Ecology Resources* **8**: 103–106.

Satori D, Tovar C, Faruk A, *et al.* 2022. Prioritising crop wild relatives to enhance agricultural resilience in sub-Saharan Africa under climate change. *PLANTS, PEOPLE, PLANET* **4**: 269–282.

Savolainen O, Lascoux M, Merila J. 2013. Ecological genomics of local adaptation. *Nat Rev Genet* 14: 807–820.

Shafer ABA, Wolf JBW. 2013. Widespread evidence for incipient ecological speciation: a meta-analysis of isolation-by-ecology. *Ecology letters* 16 7: 940–50.

**Sork VL**. **2018**. Genomic Studies of Local Adaptation in Natural Plant Populations. *Journal of Heredity* **109**: 3–15.

**Svenning J-C, Eiserhardt WL, Normand S, Ordonez A, Sandel B**. **2015**. The Influence of Paleoclimate on Present-Day Patterns in Biodiversity and Ecosystems. *Annual Review of Ecology, Evolution, and Systematics* **46**: 551–572.

**Tiffin P, Ross-Ibarra J. 2014**. Advances and limits of using population genetics to understand local adaptation. *Trends in Ecology & Evolution* **29**: 673–680.

**Trivedi P, Batista BD, Bazany KE, Singh BK**. **2022**. Plant–microbiome interactions under a changing world: responses, consequences and perspectives. *New Phytologist* **234**: 1951–1959.

**de Villemereuil P, Gaggiotti OE, Goudet J**. **2020**. Common garden experiments to study local adaptation need to account for population structure. *Journal of Ecology* **n**/**a**.

de Villemereuil P, Gaggiotti OE, Mouterde M, Till-Bottraud I. 2016. Common garden experiments in the genomic era: new perspectives and opportunities. *Heredity* 116: 249–254.

**Visscher PM, Goddard ME**. **2015**. A General Unified Framework to Assess the Sampling Variance of Heritability Estimates Using Pedigree or Marker-Based Relationships. *Genetics* **199**: 223–232.

Wang IJ, Bradburd GS. 2014. Isolation by environment. *Molecular Ecology* 23: 5649–5662.

Warschefsky E, Penmetsa RV, Cook DR, von Wettberg EJB. 2014. Back to the wilds: Tapping evolutionary adaptations for resilient crops through systematic hybridization with crop wild relatives. *American Journal of Botany* 101: 1791–1800.

**Warschefsky EJ, von Wettberg EJB**. **2019**. Population genomic analysis of mango (Mangifera indica) suggests a complex history of domestication. *New Phytologist* **222**: 2023–2037.

Way DA, Montgomery RA. 2015. Photoperiod constraints on tree phenology, performance and migration in a warming world. *Plant, Cell & Environment* 38: 1725–1736.

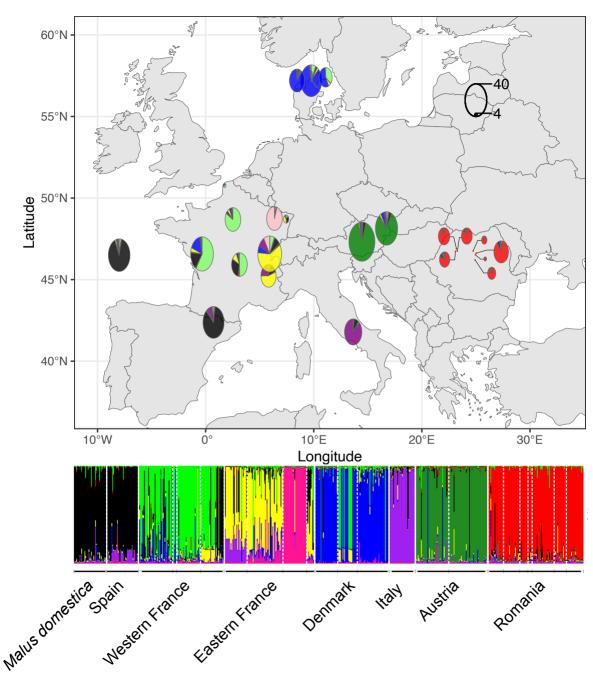
Wei T, Simko V. 2017. R package "corrplot": Visualization of a Correlation Matrix.

**Wheelwright NT, Keller LF, Postma E**. **2014**. The effect of trait type and strength of selection on heritability and evolvability in an island bird population. *Evolution* **68**: 3325–3336.

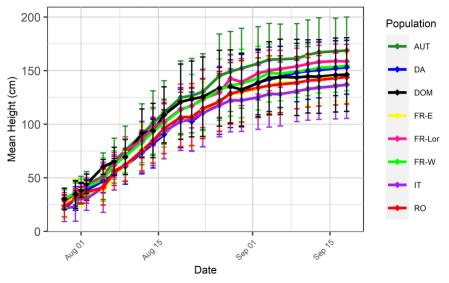
Wright S. 1943. Isolation by distance. Genetics 28: 114–138.

**Yamada T, Kokubugata G, Fujii S,** *et al.* **2021**. Refugia during the last glacial period and the origin of the disjunct distribution of an insular plant. *Journal of Biogeography* **48**: 1460–1474.

**Zhang H, Mittal N, Leamy LJ, Barazani O, Song B-H**. **2017**. Back into the wild—Apply untapped genetic diversity of wild relatives for crop improvement. *Evolutionary Applications* **10**: 5–24.



#### Evolution of height over time



# Variance partitioning of the db–RDA results

