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**Co-extruded alginate as an alternative to collagen casings in the production of dry-fermented sausages: Impact of coating composition**

Begonya Marcos, Pere Gou, Jacint Arnau, M<sup>a</sup> Dolors Guàrdia, Josep Comaposada\*

Food Technology. IRTA. Finca Camps i Armet, E-17121 Monells, Catalonia (Spain)

\* Corresponding author. Tel.: +34 972 630 052; fax: +34 972 630 980. *E-mail:* josep.comaposada@irta.cat

Keywords: co-extrusion, composite coating, casing, alginate, dry-fermented sausage

60  
61  
62 **17 Abstract**  
63

64 18 The performance of co-extruded alginate coatings containing no extra additives (A),  
65 19 polyglycerol esters of fatty acids (EA), or pea protein (PA) was assessed as an alternative to  
66 20 collagen casings (C) for the manufacturing of dry-fermented sausages (fuet) with no  
67 21 inoculation of moulds and without a fermentation step (NMNF) and fuet inoculated with  
68 22 *Penicillium candidum* and fermented (MF).

69 23 Stuffing into collagen casings resulted in slower sausage drying kinetics compared with alginate  
70 24 coating. No significant differences in  $a_w$  were observed among the studied casing types for  
71 25 NMNF and MF fuets and for the evolution of the technological and spoilage microorganisms.  
72 26 Fuets coated with A, EA, and PA showed lower pH values than fuets stuffed in collagen casings.  
73 27 No significant differences on sensory properties between casing types were observed.  
74 28 Therefore, alginate coatings would be a feasible alternative to collagen casing from a  
75 29 technological and safety point of view.  
76 30

77 31 **1. Introduction**  
78

79 32 Dry-fermented sausage manufacturing involves grinding of meat and back fat, mixing with salt,  
80 33 spices and other ingredients and additives, and stuffing into casings. After stuffing, sausages  
81 34 are fermented to the desired pH and dried to the target water content, at the appropriate  
82 35 temperature and air humidity. Traditionally, dry-fermented sausages have been stuffed into  
83 36 natural casings made from animal intestine (sheep, pork, and beef) and into artificial casings  
84 37 mainly consisting of collagen, cellulose, or plastic. Natural casings are still a very popular choice  
85 38 all over the world. However, they can be contaminated by enteric bacteria, as well as  
86 39 exogenous microorganisms, mainly due to the lack of hygiene during slaughter, post-  
87 40 processing handling and due to high storage temperatures (Trigo & Fraqueza, 1998). Artificial  
88 41 casings offer several advantages over their natural counterparts including a more uniform size,  
89 42 strength and flexibility for varying processing conditions, a lower variability in the product  
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119  
120  
121 43 weight, and a cleaner, more hygienic product (Kutas, 1987). However, like natural casings,  
122  
123 44 artificial casings are of a finite length and thus sausage stuffing remains a batch process.  
124  
125 45 The sausage casings sector has a global market share of more than €4.2 billion per year, with  
126  
127 46 continuous growth of artificial casings (Adzaly, Jackson, Villalobos-Carvajal, Kang, & Almenar,  
128  
129 47 2015). The importance of the sector has led to innovations in the sector, such as new highly  
130  
131 48 productive casing technology: co-extrusion. A co-extrusion system is a fully automated system  
132  
133 49 that simultaneously extrudes a continuous flow of meat batter and a thin layer of casing  
134  
135 50 material. The most commonly used co-extruded casing materials are composed of calcium  
136  
137 51 alginate. The principle of the co-extrusion process takes advantage of the solubility of alginates  
138  
139 52 in cold water and the ability of sodium alginate to develop strong and elastic gels on exposure  
140  
141 53 to calcium ions. Higher productivity, and lower raw material and production costs in  
142  
143 54 comparison with traditional processes are the main advantages highlighted by the suppliers of  
144  
145 55 this technology (Kamenik, 2014). Additionally, alginate coatings are a vegetable-based  
146  
147 56 alternative to animal-based and artificial casings. They are edible, and suitable for vegetarian,  
148  
149 57 halal and kosher products.  
150  
151 58 Alginates are polysaccharides obtained from the cell walls of brown seaweed, from which they  
152  
153 59 are extracted in the form of alginic acid (Lamkey, 2009). They are composed of  $\beta$ -D-  
154  
155 60 mannuronic acid (M) and  $\alpha$ -L-guluronic acid (G) in varying proportions, sequence, and  
156  
157 61 molecular weight. Alginate gellation takes place when divalent or multivalent ions (usually  
158  
159 62  $\text{Ca}^{2+}$ ) interact ionically with blocks of guluronic acid residues from two different chains  
160  
161 63 resulting in a three-dimensional network (Braccini & Perez, 2001; King, 1983). The  
162  
163 64 performance of alginate films is highly dependent on the processing conditions (e.g. mixing  
164  
165 65 speed), as well as on the alginate coating composition (e.g. presence of ions, pH)  
166  
167 66 (Comaposada, Gou, Marcos, & Arnau, 2015; Harper, Barbut, Smith, & Marcone, 2015; Marcos,  
168  
169 67 Gou, Arnau, & Comaposada, 2016; Senturk Parreidt, Schott, Schmid, & Müller, 2018). The  
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171 68 application of alginate co-extrusion for the manufacture of dry fermented sausages has  
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69 focused on the prevention of surface efflorescence formation (Hilbig, Hartlieb, Gibis,  
70 Herrmann, & Weiss, 2020; Hilbig, Murugesan, Gibis, Herrmann, & Weiss, 2019; Walz et al.,  
71 2018). However, to the knowledge of the authors, the impact of alginate coatings on the  
72 drying process and final quality of dry fermented sausages hasn't been studied. In a previous  
73 study, Comaposada et al. (2018) reported modifications of water and oxygen transfer in  
74 composite films consisting of alginate, protein and surfactant compared to alginate films with  
75 no additives in *in vitro* tests. The authors concluded that there is a need to evaluate the  
76 industrial performance of composite coatings to improve the properties of standard casings.  
77 Thus, the objective of the present work was to fill the gap of knowledge about the impact of  
78 co-extruded composite alginate coatings on the manufacture of dry-fermented sausages. With  
79 this purpose, the performance of different compositions of alginate coatings as an alternative  
80 to artificial casings for the manufacture of small calibre dry-fermented sausages (fuet) was  
81 evaluated.

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238  
239 82 **2. Materials and methods**  
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241 83 *2.1. Dry sausage (fuet) manufacturing*  
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243 84 Two types of dry sausages (fuet type) were produced: non-mould non-fermented fuet (NMNF)  
244  
245 85 produced without addition of starter culture nor mould to surface, and mould-fermented fuet  
246  
247 86 (MF) produced with addition of starter culture and surface inoculation of mould.  
248

249 87 Fuets were manufactured with 70% of pork shoulder and 30% of pork belly using the following  
250  
251 88 formula (g/kg of shoulder and belly blend): sodium chloride (18), sodium nitrite (0.15),  
252  
253 89 potassium nitrate (0.15), sodium ascorbate (0.5), dextrose (2), lactose (20) (all additives from  
254  
255 90 Merck KGaA, Darmstadt, Germany) and black pepper (3). Formula of MF fuet also included 0.1  
256  
257 91 g/kg of starter culture TEXEL® SA 306 (Danisco® DuPont™, Copenhagen, Denmark).  
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259

260 92 Lean pork meat and fat were minced in a mincer with an adjustable plate set at a hole  
261  
262 93 diameter of 5 mm. The minced meat was mixed using a vacuum mixer model 35P (Tecnotrip  
263  
264 94 S.A., Terrassa, Spain) with the ingredients at 0 °C for 3 minutes in a vacuum mixer. At this  
265  
266 95 point, the meat batter was either stuffed into collagen casings using a vacuum stuffer or co-  
267  
268 96 extruded with an alginate coating using the vacuum stuffer with a co-extruding unit developed  
269  
270 97 in a mechanical workshop nearby for this specific activity (Figure 1). Four different casings  
271  
272 98 were used for each type of fuet (NMNF and MF): Collagen (C), stuffed in 40 mm diameter  
273  
274 99 collagen casings (Fibran S.A., Sant Joan de les Abadesses, Spain); Alginate (A), co-extruded with  
275  
276 100 a coating containing 2 % of sodium alginate (Alcogel 6061, Cargill France S.A.S, Saint-Germain-  
277  
278 101 en-Laye, France); E 475 alginate (EA), co-extruded with a coating containing 2 % of sodium  
279  
280 102 alginate and 1% of polyglycerol esters of fatty acids (E 475, Lasenor Emul, S.L., Olesa de  
281  
282 103 Montserrat, Spain); and pea protein alginate (PA), co-extruded with a coating containing 2 % of  
283  
284 104 sodium alginate and 1 % of pea protein (Provital Group, Barcelona, Spain). The concentrations  
285  
286 105 of A, EA and PA were selected according to the results obtained in Comaposada et al. (2018).  
287  
288 106 One percent of EA reduced mass transfer properties, and PA was limited to 1% due to its  
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290 107 negative effect to the mechanical properties of the film. After the co-extrusion, the sausages  
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298 108 coated with alginate solutions (A, EA, and PA) were cross-linked by immersion in a 30 %  
299  
300 109 calcium chloride (Cargill Inc., Minneapolis, MN, USA) solution.  
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302 110 After stuff casing/coating MF fuets were surface inoculated by immersion into a solution of  
303  
304 111 *Penicillium candidum* spores (CHOOZIT™ Cheese Cultures, Danisco) and subjected to a  
305  
306 112 fermentation process (36 h at  $21.5 \pm 0.1$  °C and  $86.9 \pm 3.2$  % RH). Afterwards, the sausages  
307  
308 113 were hung in a drying chamber until an average weight loss of 40 % was achieved. Average  
309  
310 114 drying conditions were:  $12.7 \pm 1.0$  °C and  $74.5 \pm 2.6$  % air relative humidity (RH) for NMNF  
311  
312 115 fuets and  $13.1 \pm 0.5$  °C and  $71.6 \pm 6.3$  % RH for MF fuets. The dryer needed about 20 % of the  
313  
314 116 daily time to maintain the drying conditions with an air velocity of  $0.15 \pm 0.08$  m/s.  
315  
316 117 A minimum of 25 sausages per casing type (C, A, EA, PA) were produced for each fuet type  
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318 118 (NMNF, MF). Two independent manufacturing batches were carried on.  
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### 321 119 2.2. Drying process monitoring

322  
323 120 During the drying process, five sausages per casing type (C, A, EA, PA) for each fuet type  
324  
325 121 (NMNF, MF) were monitored for weight control until an average weight loss of 40 % was  
326  
327 122 achieved. Weight loss (%), weight loss rate (% weight loss/day) and drying time (days) were  
328  
329 123 calculated at the end of the drying process. The calibre of sausages were measured using a  
330  
331 124 Vernier caliper.  
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333 125 Temperature, relative humidity and velocity of air were monitored in each batch with a data  
334  
335 126 logger Testo 400 (Testo SE & Co. KGaA, Lenzkirch, Germany).  
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### 339 127 2.3. Effective diffusivity

340  
341 128 Effective diffusivity ( $D_e$ ) was determined using IRTAsim (IRTAsim, 2018), an online tool  
342  
343 129 developed on Matlab, which implements mass transfer equations and heat transfer equations  
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345 130 (Trigo & Fraqueza, 1998). Mass and heat transfer is modelled approximating stuffed products  
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347 131 to a porous infinite vertical cylinder.  
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357 132 IRTAsim tool allows to estimate the  $D_e$  coefficients as a function of water content when the  
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359 133 drying curve of a product is given together with the drying air conditions (temperature, relative  
360  
361 134 humidity, air speed and time in which air is in forced movement), sausage initial water content  
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363 135 and size(diameter, length) (<http://irtasim.irta.cat>).  
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366 136 The  $D_e$  was calculated considering the sausage water content changes during the drying  
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368 137 process (Equation 1):  
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372 138 
$$De = \exp \left( ax + b - \frac{53467}{RT} \right) \quad (1)$$
  
373  
374

375 139 where  $D_e$  is the effective diffusivity ( $m^2/s$ ),  $a$  and  $b$  are the  $D_e$  equation coefficients,  $x$  is the  
376  
377 140 water content (kg/kg dry mater),  $R$  is the gas constant ( $J \cdot K^{-1} \cdot mol^{-1}$ ) and  $T$  is temperature (K).  
378

379 141 For comparative purposes,  $D_e$  was calculated for each sausage by Equation 1 and it was  
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381 142 averaged in a water content range between 0.5 and 1.5 kg water / kg dm.  
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#### 384 143 2.4. Physico-chemical analysis

385

386 144 Three sausages per casing type (C, A, EA, PA) for each fuet type (NMNF, MF) were sampled for  
387  
388 145 physico-chemical analysis before drying and at the end of the drying process. The fat content  
389  
390 146 of meat paste before drying was determined by triplicate using a FoodScan™ Lab (Foss  
391  
392 147 Analytical, Hillerød, Denmark) device. The water content of fuets was determined in duplicate  
393  
394 148 by drying the minced samples at  $103 \pm 2^\circ C$  until reaching a constant weight (AOAC, 1990). The  
395  
396 149 pH values were measured in triplicate using a portable Crison penetration electrode connected  
397  
398 150 to a Crison pH meter PH25 (Crison Instruments S.A., Alella Spain). Water activity ( $a_w$ ) was  
399  
400 151 measured in duplicate at  $25^\circ C$  with an Aqualab equipment (Decagon Devices, Inc. Pullaman,  
401  
402 152 Washington, USA). The average of replicates were recorded for statistical analysis.  
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#### 405 153 2.5. Instrumental colour analysis

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416 154 Colour was measured using a Minolta CR-410 Chroma Meter (Minolta, Co., Ltd., Japan), with  
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418 155 illuminant D65, 2° standard observer angle, and 50 mm port size. Colour measurements were  
419  
420 156 taken on the surface and interior (transversal section) of fuetts and averaged over three zones  
421  
422 157 (upper, middle, and bottom). Colour coordinates were determined using the 1976 CIEAB  
423  
424 158 system and the results expressed as L\* (lightness), a\* (redness), and b\* (yellowness). Five  
425  
426 159 sausages per casing type (C, A, EA, PA) for each fuet type (NMNF, MF) were used for surface  
427  
428 160 colour measurements and three sausages for internal colour measurements at the end of the  
429  
430 161 drying process. The average of three readings was used for analysis.  
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#### 434 162 *2.6. Microbiological analysis*

435  
436 163 Sampling was performed at days 0 (after stuff casing/coating), 3, 7, and 15 of the drying  
437  
438 164 process. At each selected time, 25 g of fuet were 10-fold diluted in sterile BHI broth (Brain  
439  
440 165 heart infusion, DB, NJ, USA). The solution was homogenised for 1 minute in a Masticator  
441  
442 166 Classic (IUL S.A., Barcelona, Spain). After appropriate dilutions, the following determinations  
443  
444 167 were carried out: lactic acid bacteria (LAB) were enumerated by pour plating in MRS agar  
445  
446 168 (Merck KGaA, Darmstadt, Germany) incubated anaerobically at 30°C for 72 h;  
447  
448 169 *Enterobacteriaceae* were enumerated by pour plating in violet red bile glucose agar (Merck) at  
449  
450 170 30°C for 24 h. Three sausages per casing type (C, A, EA, PA) for each fuet type (NMNF, MF)  
451  
452 171 were measured. Two replicates per sausage were performed.  
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#### 455 172 *2.7. Sensory analysis*

456  
457 173 Quantitative Descriptive Analysis (Stone, Sidel, Oliver, Wooley, & Singleton, 1974) was carried  
458  
459 174 out to assess the appearance, taste/ flavour and texture of fuet. The panel consisted of six  
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461 175 trained assessors (ISO 8586-1, 1993; ISO 8586-2, 1994). The generation and selection of  
462  
463 176 descriptors was carried out by open discussion in three sessions prior to the experiments. The  
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465 177 descriptors retained for the sensory profile are described in Table 1. As detailed in Table 1,  
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467 178 some descriptors were assessed on the whole fuet piece and others on 5 mm-thick slices. In  
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475 179 addition, some attributes (odour: mould and ripened; flavour: ripened) were only evaluated in  
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477 180 mould fermented (MF) fuets because they were not present in non-mould non-fermented  
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479 181 (NMNF) fuets. A non-structured scoring scale (Amerine, Pangborn, & Roessler, 1965) was used,  
480  
481 182 where 0 and 10 meant absence and the highest intensity of the descriptor, respectively.  
482  
483 183 Sensory evaluation was undertaken in 6 sessions for each fuet type (3 sessions for each  
484  
485 184 manufacturing batch) using a complete block design (Steel, Torrie, & Dickey, 1997), where  
486  
487 185 each assessor evaluated one fuet from each casing type in each session. Samples were  
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489 186 presented to the assessors balancing the first order and the carry over effects according to  
490  
491 187 MacFie, Bratchell, Greenhoff, & Vallis (1989).  
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## 495 188 *2.8. Statistical analysis*

496  
497 189 Data obtained from non-mould non-fermented (NMNF) fuet and mould fermented (MF) fuet  
498  
499 190 were analysed separately with the General Linear Models procedure in the SAS program,  
500  
501 191 version 9.4 (SAS Institute Inc., Cary, NC, USA). The linear model included the casing type as a  
502  
503 192 fixed effect and the manufacturing batch as a block effect. The model also included as fixed  
504  
505 193 effects, the sampling time and its interaction with the casing type for the microbiological  
506  
507 194 indicators, and the assessor and the tasting session (nested to manufacturing batch and  
508  
509 195 assessor) for the sensory data. Least square means were calculated and the differences were  
510  
511 196 tested with the Tukey test ( $p < 0.05$ ).  
512

513 197

## 515 198 **3. Results and discussion**

### 517 199 *3.1. Drying parameters*

518  
519 200 The drying parameters measured during the manufacturing process of non-mould non-  
520  
521 201 fermented (NMNF) and mould-fermented (MF) fuets are summarized in Tables 2 and 3. Weight  
522  
523 202 losses in a range of 39.6% to 42.7% were obtained among the different casing types in both  
524  
525 203 fuet types (NMNF and MF). Stuffing into collagen casings (C) resulted in slower drying of NMNF  
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527 204 fuets ( $p < 0.05$ ) compared to coating with alginate solutions. The  $D_e$  of NMNF fuets with C  
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534 205 casings also tended to be lower than the ones with alginate coatings. These results confirm in  
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536 206 real production conditions those obtained by Comaposada, Marcos, Bou, & Gou (2018) using *in*  
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538 207 *vitro* conditions where non fermented salami slices covered with collagen films without  
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540 208 moulds showed lower weight loss rate than those covered with alginate films. As suggested by  
541  
542 209 Walz et al. (2018), differences observed in sausages stuffed in collagen and alginate casings  
543  
544 210 could be related to differences in the pore size of the casings. The pore size of collagen casings  
545  
546 211 have been reported to be around 5 nm (Gong et al., 2016), while the pore size of alginate films  
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548 212 (1.5-3.5% alginate) have been found to be in a range of 158-243  $\mu\text{m}$ . This fact could explain the  
549  
550 213 lower weight loss rate observed in C fuets. The weight loss rate of MF fuets with C casing was  
551  
552 214 also lower, while the  $D_e$  became similar among the fuets with the different coatings. This is  
553  
554 215 because the calculation of  $D_e$  integrates the effect of the initial diameter, which is bigger with  
555  
556 216 the C casing fuets. Moreover, the presence of moulds can influence the water vapour  
557  
558 217 transmission through the casing/coatings making it more uniform. In agreement with this  
559  
560 218 results, Comaposada et al. (2018) reported a significant decrease of weight loss rates of salami  
561  
562 219 slices covered with alginate films containing surfactant E 475. Senturk Parreidt et al.(2018)  
563  
564 220 related the decreased rate of moisture loss in alginate coatings containing surfactants with the  
565  
566 221 reduced superficial water activity. However, no significant differences in weight loss rate  
567  
568 222 between the different alginate coating types used (A, EA, and PA) were observed. The present  
569  
570 223 study showed that in real production conditions, the addition of 1% of polyglycol esters of  
571  
572 224 fatty acids (E 475) was not enough to significantly modify water transfer through alginate  
573  
574 225 coatings.  
575  
576 226 Although NMNF and MF fuets achieved a similar weight loss they had differences in the  
577  
578 227 moisture content due to the differences in fat content of the meat paste before drying:  $16.7 \pm$   
579  
580 228  $2.6 \%$  in NMNF and  $21.3 \pm 4.1 \%$  in MF. Similarly, the lower weight loss rates and  $D_e$  of MF fuets  
581  
582 229 were also attributed to their higher fat content. Palumbo et al. (1977) and Mulet et al. (1992)  
583  
584 230 also reported lower  $D_e$  coefficients in their meat products when fat content increased.  
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593 231 NMNF fuets coated with alginate presented higher moisture ( $p < 0.05$ ) than fuets stuffed in  
594  
595 232 collagen casings at the beginning and at the end of processing (Table 2). Walz et al. (2018) also  
596  
597 233 reported higher moisture content on sausages coated with alginate compared to those stuffed  
598  
599 234 in collagen casings, mainly due to the high moisture content of the alginate coating itself.  
600  
601 235 Comaposada et al. (2018) described similar differences in moisture content between both  
602  
603 236 casing types, which were attributed to the different type and concentration of salt present in  
604  
605 237 alginate coatings (calcium chloride) and in the collagen casings (sodium chloride). No  
606  
607 238 significant differences in moisture content at the end of the drying process were detected  
608  
609  
610 239 (Table 3).  
611  
612 240 No significant differences in the  $a_w$  of fuets were observed among the studied casing types for  
613  
614 241 any of the fuet types (NMNF and MF). It is important to highlight that all the studied samples  
615  
616 242 of fuet presented  $a_w$  values lower than 0.92. Thus, according to Regulation (EC) 2073/2005  
617  
618 243 these type of fuets are considered ready-to-eat foods unable to support the growth of *Listeria*  
619  
620 244 *monocytogenes* (European Commission, 2005). These results indicate that the use of co-  
621  
622 245 extruded alginate as an alternative casing system to commercial collagen casings would not  
623  
624 246 compromise the food safety of these products with regard to *L. monocytogenes*.  
625  
626  
627 247

### 628 248 3.3. Colour parameters

630 249 The colour of the product is a very important quality factor for dry-cured meat products  
631  
632 250 (Morales, Guerrero, Claret, Guàrdia, & Gou, 2008). Instrumental colour measurements  
633  
634 251 revealed significant differences among the studied casing types (Tables 2 & 3) on the product  
635  
636 252 surface, but not in the internal part (transversal section).

638 253 NMNF fuets coated with alginate (A, EA and PA) showed a slight increase of the superficial  $a^*$   
640  
641 254 value compared to those stuffed in collagen casings (C). A fuets also showed a slight decrease  
642  
643 255 of the superficial  $b^*$  value. MF fuets, coated with A and EA showed a slight decrease of the  
644  
645 256 superficial  $a^*$  values with respect to C and PA fuets. A and EA fuets also showed higher

650  
651  
652 257 superficial L\* values ( $p < 0.05$ ), mainly due to the higher growth of white mould on the surface  
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654 258 of those fuets (Figures 2 & 3). Except for the L\* value at the surface in MF, the rest of the  
655  
656 259 differences between C fuets and those coated with alginates were not important, including the  
657  
658 260 internal colour. Therefore, the obtained results confirm that colour would not be a drawback  
659  
660 261 for the use of alginate coatings as an alternative to traditional casings for this type of product.  
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#### 664 263 3.4. Evolution of microbiological indicators and pH values

666 264 The statistical analysis revealed that the interaction between the sampling time and the casing  
667  
668 265 type was not significant. The type of casing effect was not significant for the selected microbial  
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670 266 indicators (LAB and *Enterobacteriaceae*), but the sampling time effect was significant.  
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674 267 The evolution of LAB, *Enterobacteriaceae* and the pH values was very different depending on  
675  
676 268 the product type (Figures 4 and 5). NMNF fuets presented initial LAB counts of 2.0-2.3 log  
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678 269 CFU/g. Endogenous LAB population increased gradually throughout the drying process, with a  
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680 270 maximum increase of 3.3-3.4 log units detected at the end of the process (day 15). MF fuets,  
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682 271 inoculated with a starter culture of LAB at levels of 5.4-5.5 log CFU/g, showed similar increases  
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684 272 (3.2-3.5 log units), but the maximum growth was achieved much earlier, at the 3rd day of  
685  
686 273 drying. MF fuet showed higher LAB counts ( $p < 0.05$ ) than NMNF fuet throughout the drying  
687  
688 274 process, resulting in significantly lower pH values in MF fuets, as previously reported. No  
689  
690 275 significant growth of *Enterobacteriaceae* was observed in NMNF fuets, while a decrease of 1  
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692 276 log unit ( $p < 0.05$ ) was observed in MF fuets (Figures 4 and 5). The control of *Enterobacteriaceae*  
693  
694 277 growth is essential to prevent quality and safety defects such as the formation of off-flavours  
695  
696 278 and biogenic amines (Garriga et al 1996; Maijala et al, 1995). The pH drop within the first days  
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698 279 of the process contributed to reduce endogenous *Enterobacteriaceae* population in fuets  
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700 280 inoculated with starter culture (MF).  
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711 281 Although no significant differences among casing types were observed on the evolution of the  
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713 282 technological (LAB) and spoilage (*Enterobacteriaceae*) microorganisms, fuets stuffed in A, EA,  
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715 283 and PA coatings showed lower pH values than the ones stuffed in collagen casings (C) for both  
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717 284 fuet types (NMNF and MF) which could contribute to improve the food safety of these type of  
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719 285 products. These results confirm that the use of alginate coatings would be a feasible  
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721 286 alternative to the use of collagen casing from a technological and safety point of view.  
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### 723 287 3.5. Sensory analysis

724  
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726 288 Visual assessment of NMNF revealed that alginate coated fuets showed significantly lower  
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728 289 brightness and colour intensity than C fuets (Table 4). While in MF fuets no colour differences  
729  
730 290 were detected between A and C fuets (Table 5).  
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732  
733 291 No significant differences in global odour or flavour intensities between casing types were  
734  
735 292 detected for NMNF and MF fuets. However, some specific odour and flavour differences were  
736  
737 293 detected in MF fuets. Mould and ripened odour, and flavour were more intense in alginate  
738  
739 294 coated MF fuets than in C fuets, probably due to the higher presence of white mould on their  
740  
741 295 surface (Figure 2). Moulds are known to provide a pleasant appearance and contribute to the  
742  
743 296 development of the taste and aroma of fermented meat products (Cook, 1995).  
744

745 297 Calcium chloride used to cross-link alginate coatings can produce bitter taste (Frye, Hand,  
746  
747 298 Calkins, & Mandigo, 1986; Hand, Terrell, & Smith, 1982; Lawless, Rapacki, Horne, & Hayes,  
748  
749 299 2003). However, no bitterness was noticed in any A fuets.

750  
751 300 Collagen casings were removed before the sensory analysis, while alginate coated fuets were  
752  
753 301 evaluated with the coating. Consequently, the sensory analysis revealed that the alginate  
754  
755 302 coating was clearly perceived by the assessors (data not shown).  
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757  
758 303 Both NMNF and MF fuets stuffed in collagen casings showed lower values of hardness and  
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760 304 tactile waxy feeling than alginate coated fuets. Similarly, the assessors found fuets stuffed in  
761  
762 305 collagen casings to be less hard and more crumbly in the mouth. NMNF fuets stuffed in  
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764 306 collagen casings also showed lower elasticity and gumminess than those coated with alginate.  
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770 307 The higher pH in C fuets (Tables 2 and 3) could partly explain the differences in texture. The  
771  
772 308 consistency of fermented sausages increases due to acidification and drying, as the pH decline  
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774 309 during fermentation results in myofibrillar protein aggregation to form a gel (Arnau, Serra,  
775  
776 310 Comaposada, Gou, & Garriga, 2007).

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#### 780 312 **4. Conclusions**

781  
782 313 The reported results have shown that the differences observed during the drying process of  
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784 314 alginate coated and collagen stuffed fuets did not affect the product stability ( $a_w$  and pH).  
785  
786 315 Moreover, no remarkable differences on the internal colour and sensory properties of A fuets  
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788 316 compared to C ones would support the use of co-extruded alginate as an alternative to  
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790 317 artificial casings. These results confirm that the use of alginate coatings would be a feasible  
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792 318 alternative to collagen casing for the production of fuet type small-calibre dry-fermented  
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794 319 sausages from a technological and safety point of view. The potential of producing other types  
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796 320 of dry-fermented meat products should be explored.

797  
798 321 However, the additives used to produce composite coatings (polyglycerol esters of fatty acids  
799  
800 322 and pea protein) did not improve the properties of alginate coatings. Further research in order  
801  
802 323 to improve alginate based composite coatings for the production of dry-fermented meat  
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804 324 products should be performed.

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814 329 de Catalunya.

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416

Figure 1. Pilot equipment for co-extrusion of the meat batter with the alginate coatings.



Figure 2. Image of fuets MF at the beginning of the drying process (day 3). From left to right C, A, EA, PA.



Figure 3. Image of fuets MF at the end of the drying process. From left to right C, A, EA, PA.



Figure 4. LAB (a), *Enterobacteriaceae* (b) levels, and pH values (c) during the drying process of non-mould non-fermented fuet (NMNF). C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate.

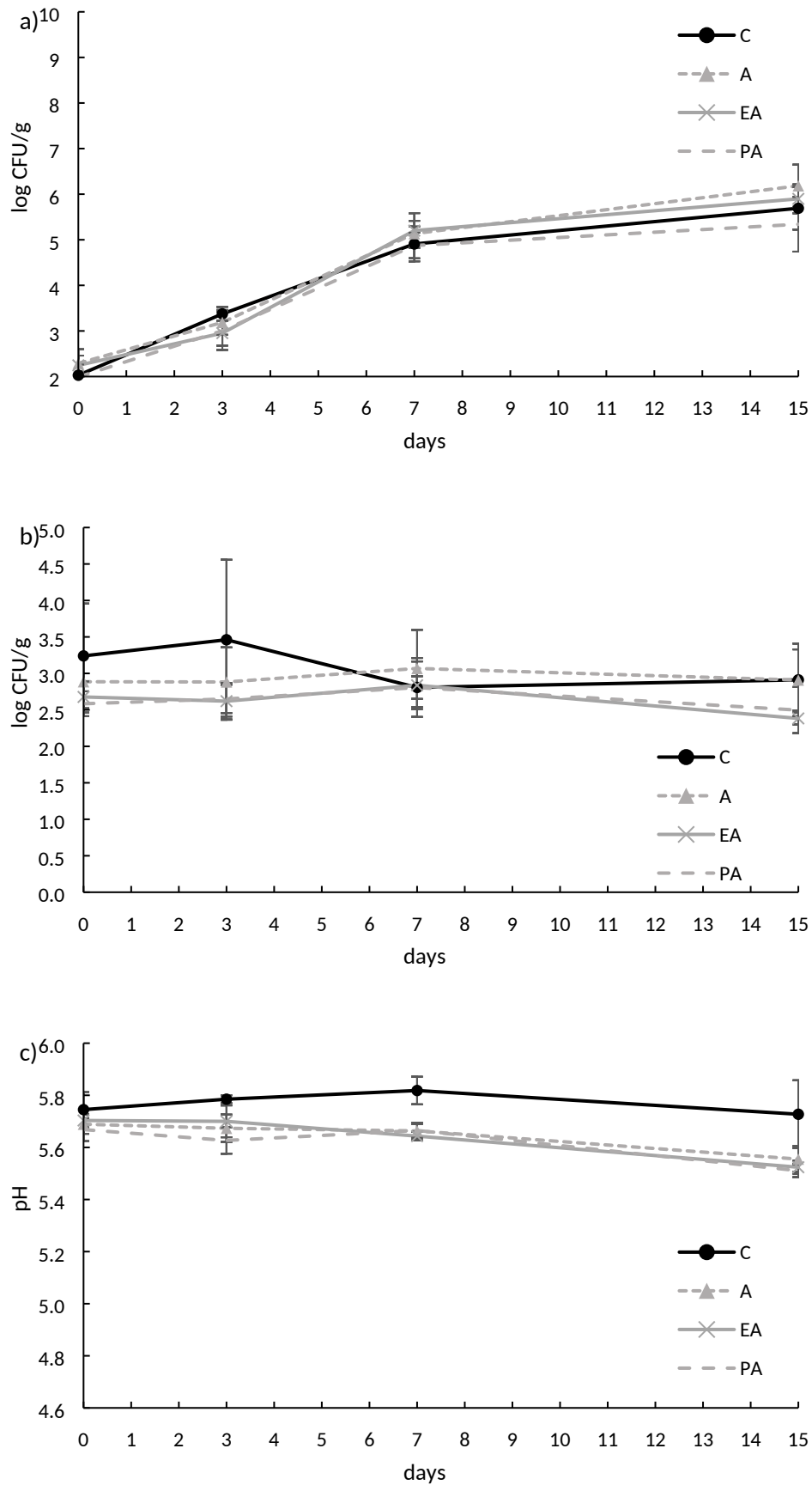


Figure 5. LAB (a), *Enterobacteriaceae* (b) levels, and pH values (c) during the drying process of mould-fermented fuet (MF). C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate.

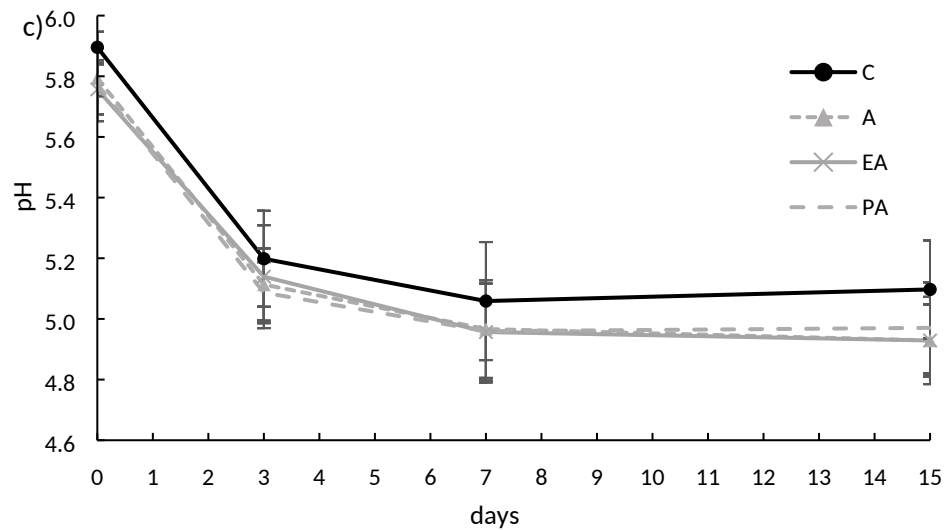
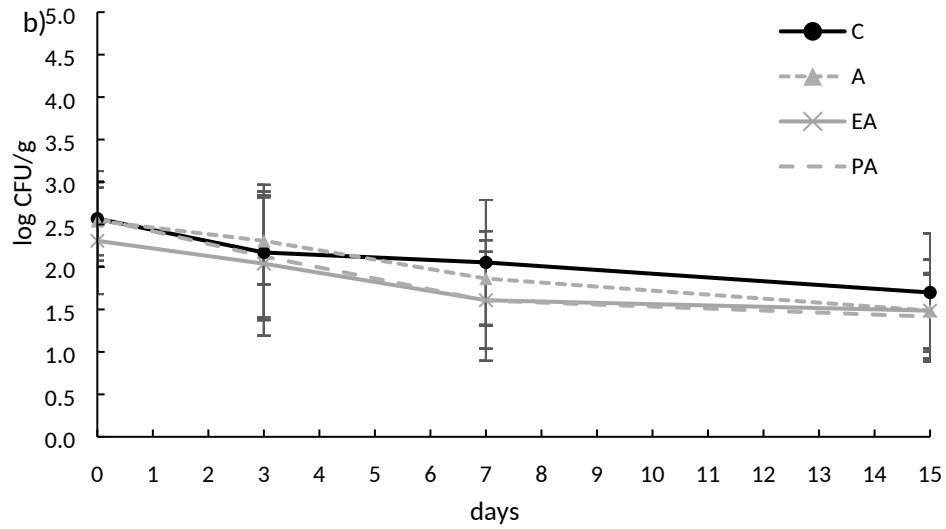
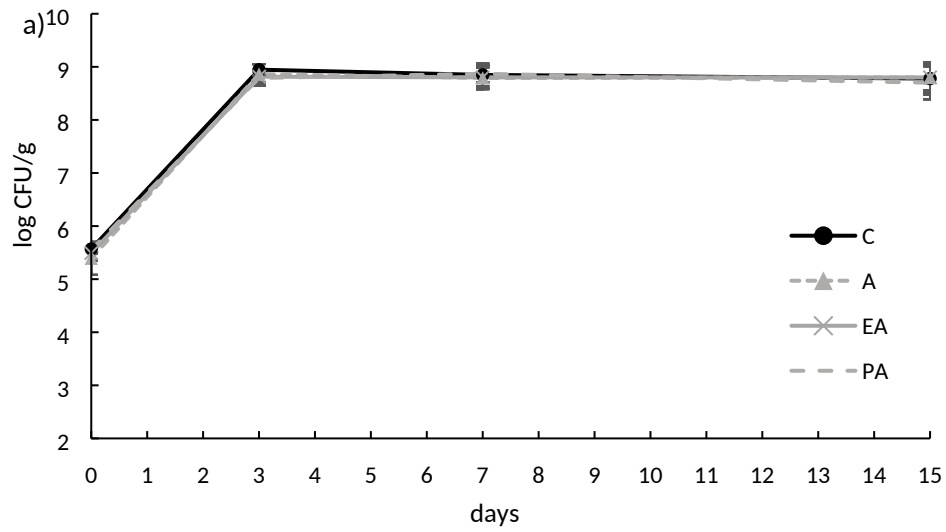


Table 1. Definition of sensory parameters used in the Quantitative Descriptive Analysis of fuet.

<b>Descriptors<sup>1</sup></b>	<b>Definition</b>
<i>Appearance (slice)</i>	
Colour intensity	Red colour intensity on a transversal section (on a freshly cut)
Brightness	Brightness intensity evaluated on a transversal section (on a freshly cut)
<i>Odour (slice)</i>	
Intensity	Overall odour intensity of fuet
Mould	Intensity of mould odour
Ripened	Pleasant odour characteristic of dry-fermented sausages
<i>Taste and flavour (slice)</i>	
Flavour Intensity	Overall flavour intensity of fuet
Sourness	Basic flavour sensation elicited by lactic acid
Bitterness	Basic taste sensation elicited by L-Tryptophan
Ripened	Pleasant flavour characteristic of dry-fermented sausages
<i>Texture (tactile; whole piece)</i>	
Hardness	Amount of pressure required to completely compress the sample
Waxy	Resembling wax when touching
<i>Texture (slice)</i>	
Hardness	Force required to bite through the sample
Elasticity	Degree of return to the original position of the sample when a compression force is applied between molars
Crumbliness	Texture property characterized by the ease with which the product can be separated into smaller particles during chewing
Gumminess	Texture property of a sample which resembles or of the consistency of gum

Table 2. Drying, physico-chemical, and colour parameters of non-mould non-fermented (NMNF) fuet.

Parameters	C	A	EA	PA	RMSE
Weight loss (%) <sup>1</sup>	39.6 <sup>b</sup>	41.0 <sup>a</sup>	40.4 <sup>ab</sup>	40.0 <sup>ab</sup>	1.2
Drying time (day) <sup>1</sup>	17.8 <sup>a</sup>	17.3 <sup>b</sup>	17.3 <sup>b</sup>	17.3 <sup>b</sup>	0.2
Weight loss rate (% weight loss/day) <sup>1</sup>	2.25 <sup>b</sup>	2.41 <sup>a</sup>	2.38 <sup>a</sup>	2.36 <sup>a</sup>	0.06
$D_e$ ( $\times 10^{-11}$ )	2.42	3.03	2.33	2.28	0.87
Calibre <sub>0</sub> (mm) <sup>1</sup>	36.5	36.1	36.4	36.0	0.6
Calibre <sub>f</sub> (mm) <sup>1</sup>	33.4	33.1	33.5	34.3	1.1
Moisture <sub>0</sub> (%) <sup>2</sup>	62.2 <sup>b</sup>	63.6 <sup>a</sup>	63.7 <sup>a</sup>	63.7 <sup>a</sup>	0.7
Moisture <sub>f</sub> (%) <sup>2</sup>	36.9 <sup>b</sup>	39.5 <sup>a</sup>	39.0 <sup>a</sup>	38.9 <sup>a</sup>	1.0
$a_{w,0}$ <sup>2</sup>	0.975	0.975	0.977	0.975	0.001
$a_{w,f}$ <sup>2</sup>	0.912	0.919	0.918	0.915	0.004
pH <sub>0</sub> <sup>2</sup>	5.75 <sup>a</sup>	5.69 <sup>ab</sup>	5.68 <sup>b</sup>	5.67 <sup>b</sup>	0.04
pH <sub>f</sub> <sup>2</sup>	5.73 <sup>a</sup>	5.56 <sup>b</sup>	5.52 <sup>c</sup>	5.51 <sup>c</sup>	0.07
$L^*_{f, \text{surface}}$ <sup>2</sup>	41.8 <sup>a</sup>	38.4 <sup>b</sup>	41.0 <sup>ab</sup>	40.7 <sup>ab</sup>	2.4
$a^*_{f, \text{surface}}$ <sup>2</sup>	6.4 <sup>b</sup>	8.1 <sup>a</sup>	7.9 <sup>a</sup>	8.7 <sup>a</sup>	1.0
$b^*_{f, \text{surface}}$ <sup>2</sup>	1.3 <sup>a</sup>	0.3 <sup>b</sup>	1.0 <sup>ab</sup>	1.0 <sup>ab</sup>	0.8
$L^*_{f, \text{internal}}$ <sup>2</sup>	45.0	46.1	45.1	46.7	2.3
$a^*_{f, \text{internal}}$ <sup>2</sup>	9.9	9.8	9.7	9.5	0.8
$b^*_{f, \text{internal}}$ <sup>2</sup>	3.0	2.8	2.4	2.7	0.7

C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate; RMSE: root mean square error; <sup>1</sup>n=5; <sup>2</sup>n=3; <sub>0</sub>: day 0 (before drying); <sub>f</sub>: end of drying process; surface: colour measured on the surface of fuet; interior: colour measured on the transversal section of fuet. Different letters within a row indicate significant differences between batches (p < 0.05).

Table 3. Drying, physico-chemical, and colour parameters of mould-fermented (MF) fuet.

Parameters	C	A	EA	PA	RMSE
Weight loss (%) <sup>1</sup>	39.9 <sup>c</sup>	41.7 <sup>b</sup>	42.4 <sup>ab</sup>	42.7 <sup>a</sup>	0.8
Drying time (day) <sup>1</sup>	20.3 <sup>a</sup>	19.8 <sup>b</sup>	19.8 <sup>b</sup>	19.8 <sup>b</sup>	0.2
Weight loss rate (% weight loss/day) <sup>1</sup>	2.03 <sup>c</sup>	2.16 <sup>b</sup>	2.20 <sup>ab</sup>	2.21 <sup>a</sup>	0.04
$D_e$ (x10 <sup>-11</sup> )	2.14	2.02	1.97	1.97	0.34
Calibre <sub>0</sub> (mm) <sup>1</sup>	36.6 <sup>a</sup>	34.1 <sup>c</sup>	35.2 <sup>b</sup>	35.2 <sup>b</sup>	0.8
Calibre <sub>f</sub> (mm) <sup>1</sup>	31.2	29.9	29.8	29.6	1.6
Moisture <sub>0</sub> (%) <sup>2</sup>	58.6 <sup>b</sup>	58.9 <sup>b</sup>	60.9 <sup>a</sup>	60.7 <sup>a</sup>	0.8
Moisture <sub>f</sub> (%) <sup>2</sup>	33.0	32.8	32.2	32.5	1.1
$a_{w,0}$ <sup>2</sup>	0.974 <sup>ab</sup>	0.973 <sup>b</sup>	0.974 <sup>ab</sup>	0.975 <sup>a</sup>	0.001
$a_{w,f}$ <sup>2</sup>	0.900	0.887	0.888	0.863	0.027
pH <sub>0</sub> <sup>2</sup>	5.89 <sup>a</sup>	5.79 <sup>ab</sup>	5.77 <sup>b</sup>	5.76 <sup>b</sup>	0.07
pH <sub>f</sub> <sup>2</sup>	5.10 <sup>a</sup>	4.93 <sup>c</sup>	4.98 <sup>b</sup>	4.97 <sup>bc</sup>	0.03
$L^*_{f, surface}$ <sup>2</sup>	58.9 <sup>b</sup>	70.8 <sup>a</sup>	69.6 <sup>a</sup>	61.2 <sup>b</sup>	5.8
$a^*_{f, surface}$ <sup>2</sup>	2.6 <sup>a</sup>	0.6 <sup>b</sup>	0.7 <sup>b</sup>	3.2 <sup>a</sup>	1.5
$b^*_{f, surface}$ <sup>2</sup>	3.7	4.5	4.1	3.7	0.8
$L^*_{f, internal}$ <sup>2</sup>	49.7	50.2	48.4	48.2	1.6
$a^*_{f, internal}$ <sup>2</sup>	8.5	9.1	9.1	9.1	0.6
$b^*_{f, internal}$ <sup>2</sup>	2.6	2.7	2.1	2.1	0.5

C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate; RMSE: root mean square error; <sup>1</sup>n=5; <sup>2</sup>n=3; <sub>0</sub>: day 0 (before drying); <sub>f</sub>: end of drying process; surface: colour measured on the surface of fuet; interior: colour measured on the transversal section of fuet. Different letters within a row indicate significant differences between batches ( $p < 0.05$ ).



Table 4. Sensory analysis of non-mould non-fermented (NMNF) fuet.

<b>Descriptors</b>	<b>C</b>	<b>A</b>	<b>EA</b>	<b>PA</b>	<b>RMSE</b>
<i>Appearance</i>					
Colour intensity	6.3 <sup>a</sup>	5.4 <sup>b</sup>	5.0 <sup>b</sup>	5.4 <sup>b</sup>	1.0
Brightness	4.6 <sup>a</sup>	3.8 <sup>b</sup>	3.8 <sup>b</sup>	3.9 <sup>b</sup>	0.8
<i>Odour</i>					
Intensity	5.8	5.6	5.2	5.1	0.5
<i>Flavour</i>					
Intensity	5.8	5.4	5.7	5.3	0.8
Sourness	2.3	2.7	2.8	2.6	1.2
<i>Texture (tactile)</i>					
Hardness	4.0 <sup>c</sup>	5.0 <sup>a</sup>	5.8 <sup>a</sup>	5.4 <sup>ab</sup>	1.1
Waxy	1.5 <sup>b</sup>	3.4 <sup>ab</sup>	3.3 <sup>a</sup>	3.8 <sup>a</sup>	1.5
<i>Texture (mouth)</i>					
Hardness	3.3 <sup>c</sup>	4.1 <sup>bc</sup>	5.1 <sup>a</sup>	4.7 <sup>ab</sup>	1.0
Elasticity	2.5 <sup>c</sup>	2.9 <sup>bc</sup>	3.6 <sup>ab</sup>	3.7 <sup>a</sup>	1.4
Crumblieness	6.1 <sup>a</sup>	5.1 <sup>b</sup>	4.9 <sup>b</sup>	5.0 <sup>b</sup>	1.0
Gumminess	2.1 <sup>b</sup>	2.9 <sup>a</sup>	3.2 <sup>a</sup>	3.1 <sup>a</sup>	1.2

C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate; RMSE: root mean square error; n=6. Different letters within a row indicate significant differences between batches (p <0.05).

Table 5. Sensory analysis of mould-fermented (MF) fuet.

<b>Descriptors</b>	<b>C</b>	<b>A</b>	<b>EA</b>	<b>PA</b>	<b>RMSE</b>
<i>Appearance</i>					
Colour intensity	5.2 <sup>ab</sup>	4.8 <sup>b</sup>	5.0 <sup>ab</sup>	5.3 <sup>a</sup>	0.9
Brightness	3.7	3.5	3.7	4.0	0.7
<i>Odour</i>					
Intensity	6.0	5.9	5.7	5.9	0.9
Mould	1.3 <sup>b</sup>	2.2 <sup>a</sup>	2.2 <sup>a</sup>	2.2 <sup>a</sup>	0.9
Ripened	3.8 <sup>b</sup>	4.5 <sup>a</sup>	4.3 <sup>ab</sup>	4.2 <sup>ab</sup>	0.8
<i>Flavour</i>					
Intensity	6.0	5.5	5.6	5.7	0.8
Sourness	3.3 <sup>a</sup>	3.4 <sup>a</sup>	3.1 <sup>ab</sup>	2.7 <sup>b</sup>	1.0
Ripened	3.1 <sup>b</sup>	3.7 <sup>a</sup>	3.8 <sup>a</sup>	4.2 <sup>a</sup>	0.9
<i>Texture (tactile)</i>					
Hardness	4.7 <sup>b</sup>	5.7 <sup>a</sup>	5.8 <sup>a</sup>	6.2 <sup>a</sup>	1.0
Waxy	2.3 <sup>b</sup>	4.3 <sup>a</sup>	4.0 <sup>a</sup>	4.3 <sup>a</sup>	1.7
<i>Texture (mouth)</i>					
Hardness	4.1 <sup>b</sup>	4.3 <sup>ab</sup>	4.7 <sup>a</sup>	4.8 <sup>a</sup>	0.7
Elasticity	2.6	2.6	2.5	2.8	0.8
Crumbliness	6.0 <sup>a</sup>	5.7 <sup>ab</sup>	5.5 <sup>b</sup>	5.2 <sup>b</sup>	0.8
Gumminess	2.2	1.8	2.1	2.0	0.6

C: Collagen; A: Alginate; EA: E 475 alginate; PA: pea protein alginate; RMSE: root mean square error; n=6. Different letters within a row indicate significant differences between batches (p <0.05).

## **Conflicts of Interest Statement**

The authors Begonya Marcos, Pere Gou, Jacint Arnau, M<sup>a</sup> Dolors Guàrdia and Josep Comaposada certify that they have NO affiliations with or involvement in any organization or entity with any financial interest (such as honoraria; educational grants; participation in speakers' bureaus; membership, employment, consultancies, stock ownership, or other equity interest; and expert testimony or patent-licensing arrangements), or non-financial interest (such as personal or professional relationships, affiliations, knowledge or beliefs) in the subject matter or materials discussed in this manuscript.

## Author Statement

Begonya Marcos, Pere Gou, Jacint Arnau, M<sup>a</sup> Dolors Guàrdia and Josep Comaposada conceived and planned the experiments. Josep Comaposada lead the project and carried out the experiments with Begonya Marcos. M<sup>a</sup> Dolors Guàrdia planned and carried out the sensory analysis. Pere Gou and Jacint Arnau helped supervise the project. Pere Gou performed the statistical analysis and together with Jacint Arnau contributed to the interpretation of the results. Begonya Marcos took the lead in writing the manuscript. All authors provided critical feedback and helped shape the research, analysis and manuscript