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Pinna nobilis in suboptimal environments are more

tolerant to disease but more vulnerable to severe

weather phenomena

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ABSTRACT

We examined a disease outbreak of the fan mussel, *Pinna nobilis* (L.), in the Alfacs Bay (South Ebro Delta, Spain) during a period of two years in three zones exposed to a summer salinity gradient resulting from agricultural freshwater discharges and distance to the open sea. Long-term monitoring was also conducted in Fangar Bay (North Ebro Delta), featuring lower salinities and no evidence of disease. Results showed that the salinity gradient of Alfacs Bay (37.4 to 35.7) was associated to cumulative mortality (100% near the mouth, 43% in middle regions, and 13% in inner regions), thus hindering the spread of pathogens. Young specimens showed to be more tolerant to disease than large adults but become vulnerable over time. In Fangar Bay, lower salinities (30.5 to 33.5) prevented the disease but individuals were highly vulnerable to Storm Gloria which caused 60% mortality in 3 weeks, and ~100% in 6 weeks.

- Keywords: Confined waters; salinity; suspended solids; MMEs; Haplosporidium; Mycobacteria;
- 36 storm effects; pen shell; Mediterranean

1. Introduction

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The pen shell Pinna nobilis L. is an endemic Mediterranean bivalve distributed across a wide type of coastal environments including coastal and estuarine ecosystems (Butler et al., 1993; Kersting et al., 2019). In open coastal waters, the distribution and abundance of the species is greatly associated to that of seagrass meadows of Posidonia oceanica, which has been indicated as its optimal habitat (Guallart and Templado, 2012). Yet, the species has also been shown to occur in some coastal lagoons (De Gaulejac, 1995; Katsanevakis, 2006; Zakhama-Sraieb et al., 2011; Russo, 2017; Belando et al., 2015) and estuarine bays (Addis et al., 2009; Prado et al., 2014, 2020), often dominated by other soft-bottom habitats such as Cymodocea nodosa and Zostera spp. seagrass meadows, macroalgal beds of Caulerpa prolifera and/or sandy and muddy shores. Populations in both types of environments are exposed to contrasting environmental conditions that may shape their abundance, distribution, age structure and overall vulnerability and recovery from severe weather phenomena. In the open sea, the species distribution and size structure are strongly influenced by wave action and density of seagrass (P. oceanica) meadows that determine the intensity of drag forces (García-March et al., 2007a, b; Hendriks et al., 2011). The species can be found from ca. 5 to 60 m (Butler et al. 1993; García-March et al., 2007b), although higher abundances are usually reported from 10 m to up to 38 m (García-March et al., 2007b; Rouanet et al., 2015). In contrast, confined waters provide a more sheltered environment, and populations have been found to peak at less than 1 m (Zakhama-Sraieb et al. 2011; Russo, 2017; Prado et al., 2014), 2-5 m (Coppa et al., 2013) and up to 11-13 m depth (Katsanevakis 2006, 2007), possibly depending on the influence of local geomorphology in recruitment patterns (see Prado et al., 2020a). Pinna nobilis has been indicated to withstand a wide range of water temperatures from ca. 7 to 28 °C but to have a narrow window of optimal salinities ranging from 34 to 40 (Butler et al., 1993). Open sea populations in the Mediterranean dwell in a relatively stable environment

with small salinity fluctuations of around 36-39, that overlap the optimal range of the species. In contrast, pen shell populations in some coastal lagoons and estuarine bays, may have to persist in salinity conditions that are a priori suboptimal for the development of the species. For instance, in the hypersaline lagoon of the Mar Menor (SW Spain) pen shells endure salinity ranges between 42 and 47 (Fernández-Torquemada and Sánchez-Lizaso, 2011), and similar ranges from 36 to 51 are attained in the Ghar El Melh lagoon (N-E Tunisia) (Zakhama-Sraieb et al., 2011). In the lower salinity range, pen shells in Fangar Bay (Ebro Delta) are exposed to annual values ranging from 20 to 35 associated to important seasonal discharges of freshwater from rice agriculture (Carrasco et al., 2008). Despite these suboptimal conditions, massive, occasional recruitment events during favorable periods may lead to the establishment of thousands of individuals such as populations reported for the Mar Menor, Venice Lagoon, or Alfacs Bay (Prado et al., 2014; Russo, 2017; Giménez-Casalduero et al., 2020). Yet, these populations may also be more vulnerable to changes in the confined water bodies resulting from persistent urban and agricultural pressures (e.g. García-Pintado et al., 2007) and/or great storms and heavy rains, typically reducing salinity and providing large quantities of silts and sediments (Isla, 2009). For instance, mortalities of cultivated bivalves such as mussels and cockles have been reported as a result of burial by storm-eroded biodeposits (Landahl, 1988; Nehls and Thiel, 1993; Dare, 2004), but the impact in other non-commercial species may have passed unnoticed. Recently, Pinna nobilis has been listed as a "critically endangered" species by the Spanish government (BOE 251-14181, 2018), and the International Union for Nature Conservation (UICN; Kersting et al., 2019). Mortalities close to 100% of the populations have been reported along the entire Mediterranean Spanish coast and other Mediterranean countries such as France, Italy, Greece, Tunisia, Croatia, Cyprus, and Turkey (Cabanellas-Reboredo et al., 2019), which are presumably due to the cooperation of a parasitic disease by Haplosporidium pinnae

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(Catanese et al., 2018) and bacterial pathogens such as Mycobacterium sp. (Carella et al.,

2019). At present, only one population along the Spanish Mediterranean coast located in Fangar Bay (North Ebro Delta, containing hundreds of individuals) remains clean of pathogens. Just a few kilometers south, the population in Alfacs Bay (South Ebro Delta) was infected in its more external part in 2018 (García-March et al., 2020a) located next to the open sea and subjected to higher salinities (Cerralbo et al., 2019). In the other remaining Spanish population in the Mar Menor, some individuals were positive for the presence of *H. pinnae* after a long-term (in the order of months) salinity reduction to ca. 39 following a meteorological phenomenon known as "cold drop" or DANA, the Spanish acronym for isolated depression in high layers of the atmosphere, causing heavy rains in September of 2019 (Nebot-Colomer et al., submitted). These infection patterns seem to confirm the disease dispersion model at salinities of ca. 36.5-39.7 proposed by Cabanellas-Reboredo et al., (2019), and suggests the importance of preserving extreme abiotic conditions for protecting the last remaining populations of pen shells.

In this work, we evaluate the magnitude and the causes of two mass mortality events (MME) in the two Ebro Delta populations of *Pinna nobilis* in order to assess the resilience of the populations and the capacity of open estuarine environments to act as refugia for the species. The first MME took place in the summer of 2018 in the external part of Alfacs Bay (South Ebro Delta), while the second one occurred in Fangar Bay (South Ebro Delta) after the pass of Storm Gloria along the Spanish Mediterranean coast. Environmental data was collected at different monitoring times, and in the particular case of Fangar Bay, physicochemical conditions were also followed from two days after Storm until the normalization of conditions ca. 2 weeks later. In both bays, samples of recently dead individuals and survivors were collected in order to investigate a possible infection by *Haplosporidium pinnae* and *Mycobacterium* sp. related to possible lesions at the tissue level. In addition, the overall age structure of the populations from growth records in empty shells (already available for Alfacs

Bay in Prado et al., 2020a) and the size structure of surviving individuals was assessed in order to evaluate the long-term persistence of the populations.

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2. Materials and Methods

2.1. Study areas Alfacs and Fangar Bays are semi-enclosed water bodies with freshwater inputs greater than evaporation, so that their mean salinity is always lower than that of the sea (Delgado, 1987). Both bays have a great economic importance due to the presence of extensive shellfish aquaculture, particularly of oysters and mussels. Alfacs Bay spreads over ca. 49 km² of the south of the Ebro Delta. The northern shore is bordered by rice fields and receives agricultural discharges (ca. 275 x 10⁶ m³ year⁻¹) high in nutrients and organic matter that cause eutrophication (Llebot et al., 2011; Prado, 2018). The bay is characterized by a salinity-dominated stratification, with a superficial layer (0 to 2-3 m deep) of low salinity (30-35) and an outward movement, and a deep salty layer (salinity: 36-38) with an inward movement (Quijano-Scheggia et al., 2008). The bulk of the P. nobilis population is located in the southern shore of the bay -the Banya Sandspit-, which stretches along a shallow platform of 18 km² (ca. 700 m wide) at depths from ca. 30 to 130 cm covered by Cymodocea nodosa seagrass meadows (Prado et al., 2014, 2020). Densities of P. nobilis along the sandspit also follow the superficial summer salinity gradient (Cerralbo et al., 2019) with higher densities in the outer part of the bay closer to the bay mouth, and lower in the inner part (Prado et al., 2014). Fangar Bay is comparatively much smaller, covering an area of ca. 12 km² of the north of the Ebro Delta, but receives a similar amount of freshwater than Alfacs Bay (ca. 250 x 106 m³ year⁻¹; Delgado, 1987; Llebot et al., 2011), resulting in vertical stratification but with lower

superficial salinities of ca. 20-35 (Carrasco et al., 2008). Hence, the pen shell population is also

located in the shallow platform (ca. 30 to 60 cm depth) of the sandspit peninsula, but its

distribution is mostly restricted to the outer part, closer to the mouth of the bay. The submerged platform is composed of silty sediments and dominated by the seagrass *C. nodosa* along with patches of *Zostera noltii*.

2.2. Assessment of mortality and survival patterns

Long-term and large-scale patterns of pen shell mortality in Alfacs and Fangar bays were investigated at increasing distances from the opening of the bay between January 2018 and February 2020. In the much larger Alfacs Bay, three sites were selected, that were representative of outer (Site 1), middle (Site 2) and inner areas (Site 3) of the bay. At each site, three monitoring circles of 8 m of diameter were deployed at ca. 50 m apart in order to capture variability at the smaller spatial scale (Fig. 1a). In Fangar Bay, given the much smaller size of the population, only three single monitoring circles were deployed in the area with highest densities of individuals (Fig. 1b). Monitoring circles in Alfacs Bay contained from 14 to 26 animals and 9 to 22 in Fangar Bay. All individuals were marked with PVC tags for easy identification during following surveys. All circles were in shallow areas (ca. 30 to 90 cm depth) and were monitored by walking with an aquascope and by snorkeling when necessary to verify the tag number.

In January 2020, two and a half years after a first mortality event occurred in the outer part of Alfacs Bay (Site 1), an area of 1 ha was randomly selected within the impacted area and an intensive search of surviving individuals conducted within 40 contiguous transects of 50 x 5 m. In each transect living and dead young specimens (i.e., 3-year-old individuals from the last recruitment event in 2017; see Prado et al., 2020a) and large adults were counted. The same survey was conducted again in late July 2020.

An intensive monitoring was conducted to assess mortality patterns in Fangar Bay, which was severely hit by Storm Gloria on 19th to 22nd of January 2020. A first survey was conducted on the 11th of February 2020 (20 days after Storm), and then again on the 4th of March (42

days after Storm) in order to assess both short-term and medium-term mortality resulting from direct alterations in physicochemical variables, and the capacity of animals to recover normal physiological functions. Despite the restricted pen shell distribution of only ca. 2.28 ha found during in situ evaluation, during the first survey we noticed the occurrence of 4 subareas with contrasting abundances of individuals (young specimens and adults) and survival rates. In each sub-area, we conducted 15 to 21 transects of 50 x 5 m (up to a total of 77 transects) across the total area of distribution (Fig. 1b). On the first survey date, for each transect we counted the number of surviving young specimens and adults, the number of recently dead (still containing some flesh remains and having no epiphytes in the internal shells) young specimens and adults, and the number of old shells from previous mortalities. On the second survey over the same exact areas, only the number of surviving young specimens and adults were counted.

In both bays, transects were conducted simultaneously by two persons walking together and searching within half of the width area.

2.3. Size-age evaluation of individuals

The size-age structure of pen shells from Alfacs Bay has been previously determined (Prado et al., 2020a). For Fangar Bay, twenty shells ranging between 31.8 and 43.6 cm long, — including the entire size variability available in this shallow environment—, were used to determine the age range of the population and to assess the availability of different cohorts indicative of recruitment years. All individuals were collected on February 11th 2020, twenty days after Storm Gloria (19th to 22nd of January 2020) and still contained some tissues in the process of decomposition, as evidence of recent dead, possibly 1 week earlier. A sample size of N= 20 has been shown as sufficient for accurate age estimation in population studies (García-March and Márquez-Aliaga, 2007; García-March et al., 2011; Prado et al., 2020a). Briefly, one valve was processed to study the record of the posterior adductor muscle scar (PAMS) from

the interior of the shell according to García-March et al. (2011). The valve was radially cut through the PAMS and ca. 8 cm dorso-ventral sections of one side were mounted on slides. A thick section (ca. 200 μ m) of the portion glued to the slide was cut using a low-speed Buehler Isomet saw. The free surface of the slide preparation was polished to improve observation of the growth record. From each polished section, the growth record was counted. Missing records were calculated using the width of the calcite layer in the three oldest records of all specimens (García-March et al., 2011).

2.4. Analyses for detection of potential pathogens and lesions

Analyses were aimed at the detection of *Haplosporidium pinnae*, which has been reported as the main cause of mass mortality events of *P. nobilis* across the Mediterranean (Catanese et al., 2018; Carella et al., 2020) and *Mycobacterium* sp. which has also been associated to massive pen shell mortality (Carella et al., 2019; 2020).

In Alfacs Bay within Zone 1, one recently dead individual and three showing disease symptoms (N= 4) (i.e., retracted mantle and difficulty to close the valves) were taken to the laboratory in July 2018 for assessing the presence of pathogens. For Zones 2 and 3, only two recently dead individuals could be found in August 2018, which were also taken to the laboratory for analytical purposes. Later in January 2020, three apparently healthy individuals from Zone 2 and another five from Zone 3 were sacrificed for the evaluation of structural anomalies of the shell and microbiome analysis, and were also used to assess the presence of *H. pinnae* and *Mycobacterium* sp. in both areas. An additional sample from Zone 2 was collected in May 2020 from a recently dead individual found in the area (i.e., final N= 4 in Zone 2 and N= 5 in Zone 3).

For Fangar Bay, two recently dead pen shells and three living individuals without disease symptoms (N= 5) were collected after Storm Gloria. Samples of all individuals were all tested at the LIMIA using the specific primers HPNF3/HPNR3 for *H. pinnae* (Catanese et al. 2018) and

PCR conditions described by López-Sanmartín et al. (2019). For analyses of presence of *Mycobacterium* sp. we used specific primers (mycgen-f/ mycgen-r) described by Böddinghaus et al., (1990) and PCR conditions indicated by Carella et al., (2019).

For all individuals from Alfacs Bay (except that found dead in Zone 2 in May 2020), and the three sacrificed individuals from Fangar Bay, different transverse sections of each tissue, including gonad, digestive gland, kidney, mantle, and gills were fixed in Davidson's solution and preserved for a week at room temperature. Subsequently, tissues were dehydrated in ascendant alcohol series and embedded in paraffin blocks, and 3-4 µm thick sections were cut with a Microm HM330 rotary microtome and stained with Mayer's hematoxylin and eosin (H&E) for routine light microscopic examination. Ziehl-Neelsen (ZN) staining was also performed to detect acid-fast bacteria. Slides were examined under a light microscope (Olympus DP20 video camera on an Olympus BX51microscope) for possible presence of pathogens and detection of structural tissue damage.

2.5. Measurement of physicochemical variables

Physicochemical (FQ) variables including temperature, salinity, dissolved oxygen, and pH were measured monthly at each monitoring circle (N= 3 per zone in Alfacs Bay, and N= 3 in Fangar Bay) during the entire monitoring period from January 2018 to March 2020 using a YSI 6660 multiparametric probe equipped with a 650 MDS data logger.

Furthermore, for Fangar Bay, water transparency was estimated from a Secchi disk, and salinity, temperature, chlorophyll concentrations (spectrophotometer quantification after acetone extraction) were measured at 0 to 0.5 m depth (at approx. 12 to 13 h) 6 days before and 2, 5, 12, 19, 26, 33, 40, and 47 days after Storm Gloria hit the Ebro Delta (19th to 22nd of January 2020) in the three points within the bay that are regularly surveyed as a part of a monitoring program of the Catalonian Government for shellfish safety (see http://www.marinemonitoring.org). Since the mortality of the entire population was

investigated twice after the storm (20 days, and an additional sample 42 days after), the temporal evolution of FQ variables together with the presence/absence of pathogens in the tissues collected, was used to assess the more likely causes of the mortality event. Images of the entire Catalonian coast before, during and after Storm Gloria are available from the Cartographic and Geologic Institute (ICGC) of the Catalonia Government (https://visors.icgc.cat/costa/#9/41.5374/1.8718).

2.6. Analyses of data

Monthly trends in mortality were investigated with RM-ANOVA (repeated measures). For Alfacs Bay, differences among sites were investigated, whereas for Fangar Bay with a much smaller area of distribution, circles were used as replicates with no site (between subjects') effects. Data were log (x+1) transformed and tested for RM-ANOVA assumptions of normality and sphericity. Yet, since the Mauchly's Test of Sphericity indicated that the assumption of sphericity had been violated, a Greenhouse-Geisser correction was used.

Differences in the abundance of dead and living young specimens (DY and LY) and adults (DA, LA) within the surveyed area of 1 ha in Alfacs Bay affected by MME at each of the 2 sampling times were investigated with a two-way RM-ANOVA (Condition and Age, fixed factors with two levels).

Mortality patterns in Fangar Bay at the two sampling times after Storm Gloria were investigated using a one-way RM-MANOVA testing for differences in the abundance of recently dead and living young specimens (RDY and LY) and adults (RDA and LA) and the presence of older shells (OS) from dead animals, across the four observed sub-areas.

Monthly patterns in the physicochemical variables of Alfacs Bay during the two study years were assessed with a two-way factorial ANOVA (Time and Zone fixed factors), and those of Fangar Bay with a one-way ANOVA (Time fixed factor). Significant differences among factor groups were investigated using Student-Newman-Keuls (SNK) post hoc tests. ANOVA

assumptions of normality (chi-squared test) and heteroscedasticity (Cochran's test) of data were not always achieved by transformation, but the ANOVA F-statistic is still able to provide robust results (Underwood, 1997). In those instances, however, the risk of making a Type I error was minimized by setting the level of significance to 0.01. For the remaining analyses, the critical level of significance was fixed at P<0.05. SNK post hoc comparisons were used to identify significant differences.

The association between cumulative mortality patterns across the different zones of Alfacs Bay during the hottest summer months (July-August 2018 and 2019) and salinity was investigated with correlation analyses. Repeated measures analyses were conducted with the SPSS software v. 20, and ANOVAs of FQ variables and correlation analyses were performed with STATISTICA v. 12.

3. Results

3.1. Assessment of mortality and survival patterns

In Alfacs Bay, RM-ANOVA showed significant time differences across the study period (January 2018 to March 2019), as well as a significant effect of zone and its interaction (Fig. 2a, Table 1a). A mortality of up to 63% of the individuals occurred in Zone 1 in July 2018, and 98% were dead one month later (Fig. 2a). Conversely, the lower mortality in Zones 2 and 3 during the summer of 2018 was mostly attributed to impacts with boat propellers in shallow areas around the main sand bar of the bay (8 out of 9 individuals found dead) (Fig. 2a). No mortality was observed in any monitoring circle until the next summer period in 2019, when the last individual remaining in Zone 1 was found dead, and some mortality was also detected in Zone 2 (ca. 21% in July and 13.7% in August). No more mortality was recorded during the fall and winter period of 2019-2020, even after the pass of Storm Gloria in late January 2020.

The evaluation of the 1 ha area affected by mass mortality in the surroundings of Zone 1 in January 2020 showed significant differences in the condition of individuals (dead vs. alive), as

well as a significant time x condition and time x age x condition interactions (Table 1b, Fig. 2b). In January, young specimens (recruitment of 2017) showed higher survival than adults (35.6% vs. 0% of all individuals counted), with a total of 36 individuals found alive within the study area, but only 8 (7.91%) were still alive by late July 2020 (Fig. 2b).

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In Fangar Bay, dead individuals showed no apparent effect of hydrodynamic dislodgement and were all well anchored to the seagrass substrate. RM-ANOVA also indicated significant time differences, although effects were due to a massive mortality of all individuals in the three monitoring circles after Storm Gloria (Fig. 3a, Table 2a), with only minor losses registered in the area during previous monitoring months (3 individuals in two years). Only one individual was recorded as dead by collision, likely due to this bay having less touristic boat traffic.

Monitoring circles were all located in Zone 3, which together with Zone 2 (Fig. 1a), had the largest number of individuals in the area and recorded the largest number of deaths. Results from one-way RM-MANOVA showed significant differences in the numbers of living and dead young specimens and adult individuals among zones, as well as significant differences in time and zone interactions across pen shell age and condition (Fig. 3b, Table 2b). At the first sampling time (20 days after Storm), recently dead adults (RDA) were more abundant in Zone 3 (180 individuals), followed by Zone 2 (101 individuals), and then by Zones 4 and 1 (33 and 3, respectively), which also hosted much lower densities of individuals. Living adults (LA) were mostly present in Zone 2 (a total of 210 individuals), whereas in the rest of the zones only 1 to 4 were recorded. For young specimens, the bulk of the population was concentrated in Zone 1 (a total of 47 individuals vs. 13 in all the other zones). Finally, the presence of old shells (OS), was also much higher in Zones 2 and 3 than in Zones 1 and 4, demonstrating that, in the longterm, those areas have hosted the highest densities of individuals. At the second sampling time (42 days after Storm), only 12 LA were detected in Zone 2. Overall, the mortality of young specimens and adults was ca. 60% at the first sampling time and rose to nearly 100% at the second sampling time.

324 3.2. Size-age evaluation of individuals

The analysis of the growth record (posterior adductor muscle scars) of the empty shells showed that from the 20 shells cut, 75% were 4-5 years of age, followed by 15% of 6 years, and 10% of 7 year, this being the maximal age (Fig. 4). A total of 60 young specimens were also present in the study area and were estimated to be 2-3 years old (shells that were not cut because growth parameter estimation is not possible with such young shells). The growth equation for the Fangar Bay [$L_t=41.96\cdot(1-e^{(-0.39\cdot(t+0.276)})$] shows high K (0.39) and low L ∞ (41.96 cm), concurring with multiple shell fractures and reconstructions that account for small maximum sizes. The total number of individuals in the age range of this equation before Storm Gloria was 533 (RDA and LA).

3.3. Detection of potential pathogens and lesions

PCR analyses for the four investigated individuals (one recently dead and 3 with disease symptoms) collected in Zone 1 (i.e., the region of Alfacs Bay closer to the mouth of the bay) in July 2018, showed positive results for both *H. pinnae*, and *Mycobacterium* sp. (two individuals were only positive to *H. pinnae* by PCR, and the four of them were positive to *Mycobacterium* sp. through both PCR and microscopy). The two recently dead individuals found in Zone 2 and Zone 3 in August 2018 were also positive for *H. pinnae*. Further, in the case of the individual from Zone 2, the parasite could only be detected by histology, not by PCR. Among those individuals collected in January 2020 (N= 3 from Zone 2 and N= 5 from Zone 3), two from Zone 2 resulted positive for *H. pinnae* (one of them only by histology), but not for *Mycobacterium* sp. The dead individual from May 2020 in Zone 2 was also positive for *H. pinnae* by PCR. None of the individuals from Zone 2 or 3 were positive for the presence of *Mycobacterium* sp. either by PCR or histology.

In Fangar Bay, the five individuals investigated (2 recently dead examined by PCR and 3 sacrificed examined by both PCR and histology) after Storm Gloria were all negative for both *H. pinnae*, and *Mycobacterium* sp. The three individuals sacrificed after Storm Gloria showed a normal digestive gland with no evidence of *H. pinnae*, *Mycobacterium* sp., or other bacterial infections (Fig. 5a,b). However, individuals displayed a regressing female phase of gonad maturation, similar to that observed in the post-spawning phase of a normal annual reproductive cycle, with remaining oocytes after their release in late summer (Prado et al., 2020a; Deudero et al., 2017) but, without this actually occurring (Fig. 5c,d). When regression occurred, the first individual was in a previtellogenic stage, whereas the second and third individuals were already in a more advanced vitellogenic stage. Only few oocytes were observed remaining attached to the developing gonadal follicle, and the majority of them were detached and phagocyted by hemocytes (Fig. 5c,d), suggesting a limited reproductive capacity during the current year, at least at the beginning of the reproductive season.

3.4. Physicochemical variables

In Alfacs Bay, all investigated variables showed important variations across monthly study dates, as well as significant Date per Zone interactions (Table 3). For temperature, maximums of ca. 27-30 °C occurred in July-August, and minimum of ca. 9.5 to 14 °C in January-February, with non-significant differences between study zones (Fig. 6a, Table 3a). Salinity also showed important temporal differences which are attributed to seasonal rainfall and to freshwater discharges from rice cultivation, with annual maximums of ca. 37 and minimums of ca. 33, excepting a lower value of 32.16 ± 0.03 recorded in January 2020, after Storm Gloria (Fig. 6b, Table 3b). However, in the case of salinity there was also a significant zone effect, with slightly higher values (by ca. 0.6 over the entire study period) in Z1 closer to the mouth of the bay, and expectedly lower values in Z2 and Z3. Most importantly, the magnitude of those differences was higher during the summer months (June-August 2018 and 2019; see Fig. 6b), when

Haplosporidium pinnae is more infective due to high temperature. Dissolved oxygen was high in the area, with values ranging from ca. 77 to 123% of saturation, but despite significant temporal and temporal x zone variability, no clear seasonal or zone pattern could be observed (Fig. 6c, Table 3c). The pH ranges varied between minimums of ca.7.3 to 7.6 in September 2018 and April 2019, and maximums of ca. 8.3 to 8.5 in June 2018 and October 2019 (Fig. 6d, Table 3d). Significantly higher pH values were detected in Z3 in July 2019, whereas lower values were observed in Z1 in August-September 2019 (Fig. 6d).

A positive association was detected between cumulative patterns of pen shell mortality across the three study zones and the salinity (R^2 = 0.652, df= 35, F= 63.795, p= 2.62·10E⁻⁹) recorded during the months of July and August (Fig. 7).

In Fangar Bay, significant temporal variability was observed for temperature, salinity, and dissolved oxygen, but not for pH (Table 3e-h). For temperature, the temporal profile was very similar to that of Alfacs Bay, although with slightly higher maximums and lower minimums (27 to 30 °C and 7 to 13 °C, respectively; Fig. 6e, Table 3e). The salinity was lower than in Alfacs Bay, with values ranging from ca. 27 to 36, although a minimum of 11.05 ± 0.36 was observed after Storm Gloria (Fig. 6f, Table 3f). Dissolved oxygen showed a clearer seasonal variability compared to Alfacs Bay, with minimums of ca. 88-89% during the summer period and maximums of ca. 100-113% in winter (Fig. 6g, Table 3g).

At the shorter temporal scale (before and after Storm Gloria), the only FQ variables, which experienced a sharp change were salinity and the water transparency given by the Secchi disc measurements (Fig. 8A-F). Salinity was reduced from ca. 28 to 11 just after Storm Gloria with normal values being again recorded 12 days later, although a new minimum was recorded again in March (Fig. 8B). Similarly, the visibility of the Secchi disc was greatly reduced from 3.4 to 0.34 m after Storm and normal values were also recorded 12 days later (Fig. 8E). This result of strong reduction of water transparency confirms satellite images from the ICGC institution showing elevated water turbidity after Storm Gloria.

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4. Discussion

The pen shell populations of the Ebro Delta are, together with that from Mar Menor Lagoon (this study; Giménez-Casalduero et al., 2020), the only ones in Spanish waters that have persisted, -totally or partially-, after MMEs caused by the rapid spread of Haplosporidium pinnae (Vazquez-Luis et al., 2018; Cabanellas-Reboredo et al., 2019). Yet, similar patterns of survival are being observed in confined waters across other Mediterranean regions such as lagoons along the French Occitan Coast and Corse (Simide et al., 2019; García-March et al., 2020a; Foulquié et al. 2020), the Venice lagoon in Italy (Carella, pers. observ.), and the Thermaikos Gulf in Greece (Lattos et al., 2020). In Alfacs Bay, the summer salinity gradient (from 37.4 to 35.7) showed to be significantly associated to observed patterns of cumulative mortality. Although this salinity range is too narrow to prevent the infection of the population, it seems to be effectively hindering the spread of the parasite compared to open water conditions, as predicted by the dispersion model at salinities above 36.5 (Cabanellas-Reboredo et al., 2019). However, it is important to remark that salinities during the rest of the year also range from 34 to 36 in external parts of Alfacs Bay, and from 32 to 36 in intermediate and internal parts of the bay (see also Cerralbo et al., 2019), which might contribute to further obstruct the spread of the parasite and the rate of disease transmission. In fact, although in open waters H. pinnae has been reported to cause disease at temperatures as low as 13.5 °C (García-March et al., 2020a), major mortalities in Alfacs Bay were constricted to periods above 27 °C mostly during July and August. Other extant estuarine fan mussel populations (usually between ca. 30 to 40) such as those in the Diane and Thau Lagoons in France (Simide et al., 2019; Foulquié et al. 2020) might be exposed to a similar seasonal risk as those in Alfacs Bay, although the more restricted connection with the open sea might provide an additional protection. Given this more constricted period for high risk of disease and its apparently lower transmission in waters below salinity of 36.5, the persistence of estuarine populations

confronted with potential infections will be largely dependent on the ratio between mortality and new recruitment. In Alfacs Bay, Prado et al. (2020) showed that the age structure of the population is dominated by individuals of the 8 ± 1 year-old class with a maximum age of 15 years and only one major recruitment event in 11 years followed by periods of slow population growth. Also unsettling is the fact that the natural pattern of distribution and abundance of individuals across the bay is inversely related to the salinity gradient (Cerralbo et al., 2019; Prado et al., 2014), suggesting that larval viability may, to some extent, be hindered by agricultural discharges in areas that are safer from the parasite (Prado et al., 2020a). Yet, there is also genetic evidence that the population in Alfacs Bay is source of larvae for more distant populations in the Balearic Islands (Wesselmann et al., 2018), advocating that the population is capable of successful larval export.

Among other factors conditioning the persistence of the pen shell populations the occurrence of resistant individuals may be a major asset. Usually, large, adult individuals surviving two waves of the parasitic disease caused by *H. pinnae* do not die during further exposure (Vázquez-Luis, pers. comm.). Currently, only one living adult has been observed in the outer impacted area of Alfacs Bay (Prado, pers. observ.), which is consistent with mortality patterns close to 100% of individuals in other regions (Vázquez-Luis et al., 2017). Yet, 36 surviving young specimens of reproductive age (ca. 30 cm; from 2017 recruitment) were observed in January 2020 in only 1 Ha within the most impacted area, and the presence of further numbers in adjacent sites needs to be explored. The cause of such higher survival in young specimens vs. adult individuals (35.6 and 0%, respectively) is unclear, although differences in the expression of immune genes and in the capacity to mitigate oxidative damage to support physiological functions have been observed over the lifetime of bivalves (Philipp and Abele, 2008; Husmann et al., 2014). Unfortunately, individuals might end dying if exposure to the pathogen persists throughout the growth period, as observed in later monitoring (77.78% found dead by May 2020; only 8 of the 36 young specimens were found

alive in late July), suggesting a narrow window of time for reproductive success to perpetuate the species. Also possibly, the abundance of *Mycobacterium* sp. might accumulate in larger individuals (A. Grau et al., unpublished data) and could contribute to increased mortality with age and explain observed patterns over time. Yet, apparently healthy young individuals showed presence of *Mycobacterium* sp. linked to strong inflammatory lesions in different Croatian regions affected by MMEs (Šarić et al., 2020). In fact, animal immune response to intracellular *Mycobacteria* spp. is complex, due to interaction between bacterial virulence and host resistance, two distinct and independent variables, leading to periods of remission followed with disease progression (Davidovich et al. 2020). In Alfacs Bay, *Mycobacterium* sp. was found in animals from the outer region closer to the mouth of the bay (coupled with *H. pinnae*; see also Carella et al., 2020), but not in middle and inner areas, suggesting that it could be even more sensitive than *H. pinnae* to salinity variations occurring in estuarine waters. In fact, *Mycobacterium* sp. has not been found in the Mar Menor lagoon either where salinities above those of the open sea are a common feature (Nebot-Colomer et al., submitted).

In Fangar Bay, where no positives have been found for *H. pinnae* or *Mycobacterium* sp., the salinity range to which the local population of *P. nobilis* is exposed is even lower, with an annual mean of ca. 32.4 (from ca. 27 to 36 during some winter months with low rainfall). In particular, salinities from ca. 30.5 to 33.5 were detected during the summer months (2018 and 2019) with higher risk of disease, confirming that this might be a safe range to prevent infection by these pathogens (Cabanellas-Reboredo et al., 2019). Unfortunately, the availability of populations that exist in habitats with salinities consistently below the lower limit for the parasite (at 36.5) is, to our knowledge, very scarce. One possibility is the Sea of Marmara, where healthy populations are still found, possibly due to low salinities resulting from the mixing of Black Sea (ca. 18) and Aegean Sea waters (ca. 38.5) (Öndes et al., 2020), but more information is necessary about the species' abundance and seasonal fluctuations in salinities.

population in the Venice lagoon (Russo 2017), which appears to be exposed to annual salinity ranges of ca. 32-34 (Zirino et al., 2014). Similarly, there are also few examples such as that of the Mar Menor in Spain (usually 42 to 47; Fernández-Torquemada and Sánchez-Lizaso, 2011; Giménez-Casalduero et al., 2020) that are usually above the upper limit of salinity acceptable for distribution of the parasite, at 39. Even so, exceptional weather events bringing large amounts of rain into the lagoon were able to decrease salinity levels to ca. 39 and might have allowed the entrance of the parasite (Vázquez-Luis, pers. comm.), although no MMEs have yet been detected and the evolution of the disease is yet to be seen (Giménez-Casalduero, pers. comm.). Other populations inhabiting areas with a wider salinity range such as that of the Ghar El Melh lagoon in Tunisia (36 to 51; Zakhama-Sraieb et al., 2011) may also be presumably exposed to low risk due to lower salinity levels occurring during the winter period, when temperatures are less favorable for disease transmission (García-March et al., 2020a).

Although pen shell populations in confined waters appear to have a higher tolerance to disease that could help the species chances of surviving, these environments are also more vulnerable to severe weather phenomena. Twenty days after Storm Gloria, a MME of ca. 60% of the population, equally affecting young specimens and adult individuals, was observed in Fangar Bay. Gloria featured wave heights of up to 8 m along the Spanish Mediterranean coast, and in the Ebro Delta, which caused a major storm surge that entered 3 km inland, devastating rice paddies and coastal features (Amores et al., 2020). The event, however, did not cause any dislodgement of pen shells within Alfacs or Fangar Bays. This suggests that despite hydrodynamic conditions these habitats may have a major structuring role in pen shell populations (García-March et al., 2007a; Hendricks et al., 2011), as both locations are effectively protected from wind-waves. In contrast, rain induced flooding decreased the salinity of Fangar Bay to values around 11 during an approximate period of 10 days. Comparatively, these salinity values are much lower than those observed in Alfacs Bay (approximately 32) where the surface area is ca. 4 times the size and therefore attaining lower

dilution from rainfall. To our knowledge, this is the first time that the species is reported to be exposed to such a low salinity within such a short period following heavy rain. Hernandis et al. (2018), studied salinity variations in three different areas of Boka Kotorska Bay (Montenegro), and found decreases down to ca. 19 following punctual but high inputs of freshwater with no apparent effect on pen shell populations, although the exact duration of the effect is unknown (measures conducted every two weeks). Also possibly, the influence of wave action on the resuspension of fine sediments may have further contributed to observed effects. Episodes of strong weather have been shown to cause large sediment fluctuations and in situ burial of cockle (Dare 2004) and mussel beds even at high biomasses of mussels (Landahl 1988). Further, storms have been identified as a major factor limiting the distribution of mussel beds to sheltered areas of the coast due to scarcity of phytoplankton and competition for food with other communities of filter feeders (Nehls and Thiel 1993). In Fangar Bay, pen shells are located on a shallow seagrass bed of C. nodosa, which might have prevented burial by a sediment layer. However, our measures of water transparency from the Secchi disc and satellite images (ICGC visor) demonstrate increased turbidity in the water column over an estimated time period of around 10 days. Later in March, a further decrease in water transparency was also observed, coinciding with pumping of seawater from flooded rice drainage channels into Fangar Bay (see http://www.saihebro.com), which might have hindered the recovery of the remaining pen shells and contributed to additional mortalities. Pen shells have been shown to filter small-size sediment particles (< 200 μm) without an apparent assimilation for tissue growth (Prado et al., 2020b), thus suggesting a reduced intake during the siltation period. In other species such as the green-lipped mussel Perna viridis, experimental exposure to high loads of suspended solids caused no direct mortality during a period of 96 h (much shorter than Storm Gloria effects), although serious damage was reported in the structure of the gills that may impair long-term feeding and respiration (Shin et al., 2002). There may also have been some feeding and metabolic impairment as also

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suggested by the histological examination of the gonads of the 3 individuals sacrificed after Storm Gloria, in which there were found few oocytes attached to the developing gonadal follicle as described for the post-reproductive period in late summer (Prado et al., 2020a; Deudero et al., 2017). This gonad regression observed 20 days after the storm was very likely a symptom of discomfort and physiological impairment, which ended with the death of nearly 100% of the population (only 12 individuals were found alive in early March 2020) about 1.5 months later.

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Our results for the age structure of the population show that most of the individuals were young; the oldest were in the range of 4 to 7 years (much younger than the Alfacs Bay) and featured an L∞ of 41.9 suggesting that the baseline conditions of the population were already stressing and individuals might have reduced tolerance to additional distress. Such reduced life-expectancy and maximum size compared to other Mediterranean populations reaching over 50 years of age (Rouanet et al., 2015) and greater maximum sizes (García-March et al., 2020b) demonstrate that pen shells in Fangar Bay are subjected to extremely harsh conditions, -reduced salinity and desiccation at shallow depth-, causing continuous shell fractures and reconstructions. Yet, the area also hosted numerous old shells remains (approx. 500), indicating that the species may have dwelled for decades in Fangar Bay. The results indicate there has been some recruitment for most of the years (60 young specimens of 2-3 years old, and shells of 4 to 7 years), and although the effect of external supply cannot be discarded due to the presence of healthy populations in the southern French Mediterranean coast until mid-2018 (García-March et al., 2020a), the histological results indicate that the population was fertile. Now, unless self-recruitment occurs among the few individuals that are still alive or some managerial intervention is conducted with healthy individuals from Alfacs Bay, the population may face extinction in just 2-3 years. Seasonal monitoring is also needed to assess potential (although improbable) recruitment, particularly in zone 1 where an even shallower bathymetry acts as a physical barrier favoring larval settlement while hampering long-term growth (only three adults were found in this zone in February 2020; see also Prado et al., 2020a for similar patterns in Alfacs Bay), thus this population will also require later translocation of individuals.

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5. Conclusions

The Fangar population subjected to salinities of 30.5 to 33.5 during the summer period appears to be in a safe range to avoid the entrance of the pathogens (both H. pinnae and Mycobacterium sp.), but pen shells are rarely found in these extreme conditions. Most of the remaining populations in confined waters fall outside the 36.5 to 39 salinity range during some annual periods and are less vulnerable (Cabanellas-Reboredo et al., 2019), but not protected against infection by H. pinnae as evidenced for the Alfacs Bay population. Aside the degree of connection with the open sea, the chances of infection and the evolution of MMEs appear to be related to the time gap that populations are exposed to favorable salinities for the parasite and its interaction with seasonal temperatures. In Alfacs Bay, attaining even slightly lower salinities during the hottest summer months could be a feasible goal that could be accomplished thanks to the availability of a major network of drainage channels for rice agriculture that may enhance freshwater release, and help to minimize the spread of the parasite. Populations in confined waters may be more exposed to changes in environmental conditions compared to the open sea and are more vulnerable to severe weather phenomena (Fangar Bay, this study) or increased eutrophication (Mar Menor; García-Ayllon, 2018). According to historic data, 30 river floods greater than 2000 m³/s have occurred in the Ebro Delta from 1779 to 2001, with years registering up to 3 storms and gap periods of over 20 years (DMAH, 2008). Therefore, although this might constrict the establishment of a permanent population in Fangar Bay, it should still provide a more secure environment than the open sea in the context of the present pandemics.

Urgent actions are necessary to manage the salinity and conditions of habitat quality of these last remaining populations to prevent, as much as possible, further losses of individuals. Moreover, immunological studies (*in vitro* and *in vivo*) are needed to evaluate pen shells response to pathogens along the salinity gradient. Suboptimal salinity conditions can result in changes in bivalve defense mechanisms, growth, oxygen consumption, heart and filtration rate (Matozzo and Marin, 2011) that act on many aspects of disease pathogenesis. The knowledge of the mechanisms involved in immune modulation is crucial to develop management and conservation programs. The finding and preservation of surviving individuals (preferably adults but also young specimens) is another critical goal requiring intervention, rather than left to chance.

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Fig. 1. Map of the Ebro Delta showing the location of Alfacs Bay in the South and Fangar Bay in the North. A) In Fangar Bay, we indicate the four study zones were mortality patterns were assessed after Storm Gloria. Shallow seagrass beds along the four study area are indicated. Monitoring circles were deployed within zone 3, which had the highest abundance of individuals. B) In Alfacs Bay, the three study zones (Z1, Z2, and Z3) are indicated, as well as the 1 ha adjacent to Z1 where an extensive search for surviving individuals was conducted.

Fig. 2. Mortality patterns in Alfacs Bay. A) Cumulative mortality in monitoring circles located in the three study zones of Alfacs Bay, and (B) Abundance of living and dead individuals (adults and young specimens) in 1 ha of Alfacs Bay located within the mass mortality area next to the mouth of the bay (i.e., adjacent to Zone 1). DY= Dead Young specimens, LY= Living Young specimens, LA= Living Adults, DA= Dead Adults. Error bars are SE.

Fig. 3. Mortality patterns in Fangar Bay. A) Cumulative mortality in the three monitoring circles within Zone 3, and B) abundance of living and recently dead individuals (also young specimens and adults), and old shells in the four contiguous zones of Fangar Bay where the entire population is distributed. DY= Dead Young specimens, LY= Living Young specimens, LA= Living Adults, RDA= Recently Dead Adults; RDY= Recently Dead Young specimens, OS= Old Shells. Error bars are SE.

Fig. 4. Age structure and year of recruitment of Fangar Bay population from shells (N = 20) collected on February 11th 2020. The presence of 60 young specimens too small to be aged using the dating technique (2-3 years age) is also indicated.

Fig. 5. Histological sections of pen shell digestive gland and gonads from Fangar Bay after Storm Gloria. A) Section of the digestive gland at low magnification (20x) showing normal

digestive tubules (DT). B-C-) Higher magnification (40x) micrograph of the gonads, with acini full of regressing previtellogenic (B), and vitellogenic oocytes (C). In both cases, very few viable oocytes (OO) can be still observed attached to the developing follicle. Abundant hemocytes (*) coupled with brown cells (BC) can be observed. A-C). H&E staining (for interpretation of the references to color in this figure legend, the reader is referred to the web version of this article).

Fig. 6. Variability in physicochemical variables across the study period (January 2018 to March 2020) in the 3 zones of Alfacs Bay, and in Fangar Bay. A-B) Temperature (°C); C-D) Salinity; E-F) dissolved oxygen; and G-H) pH. Error bars are SE.

Fig. 7. Association between cumulative mortality rates in each zone and circle and salinity values in July-August 2018 and 2019. The significant R^2 value and the line equation are indicated.

Fig. 8. Changes in FQ variables in Fangar Bay before and after Storm Gloria. A) Temperature (°C); B) salinity; C) dissolved oxygen (%); D) pH; E) Secchi disc visibility (m); F) Chlorophyll (μg·L⁻¹). The dotted lines indicate the dates of the mortality assessment on the 11th February 2020 (individuals were found in an advanced stage of decomposition) and the 3rd of March 2020. Error bars are SE.

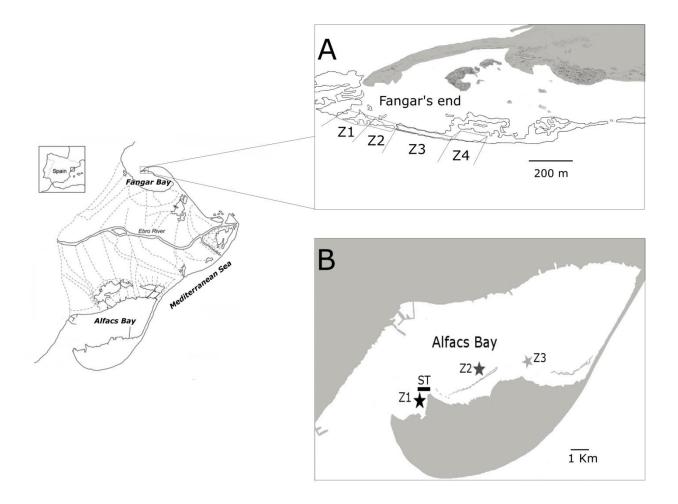
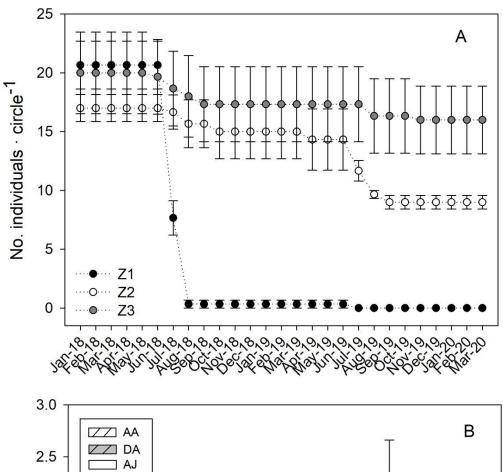


Fig. 1



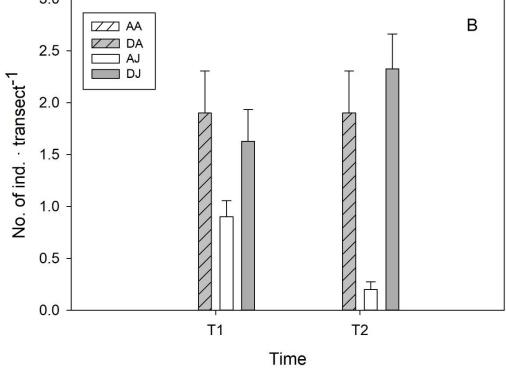


Fig. 2

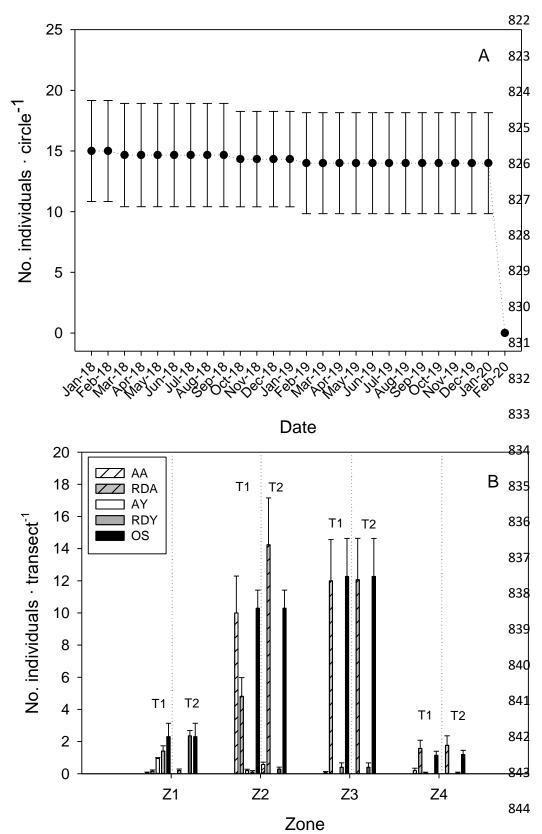


Fig. 3

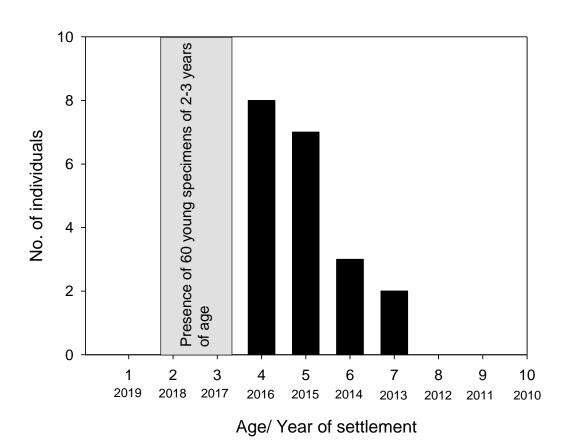


Fig. 4

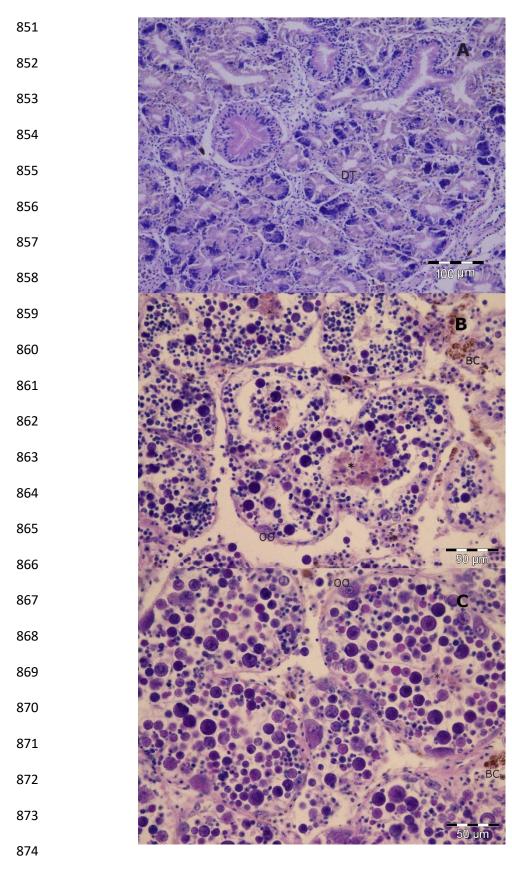


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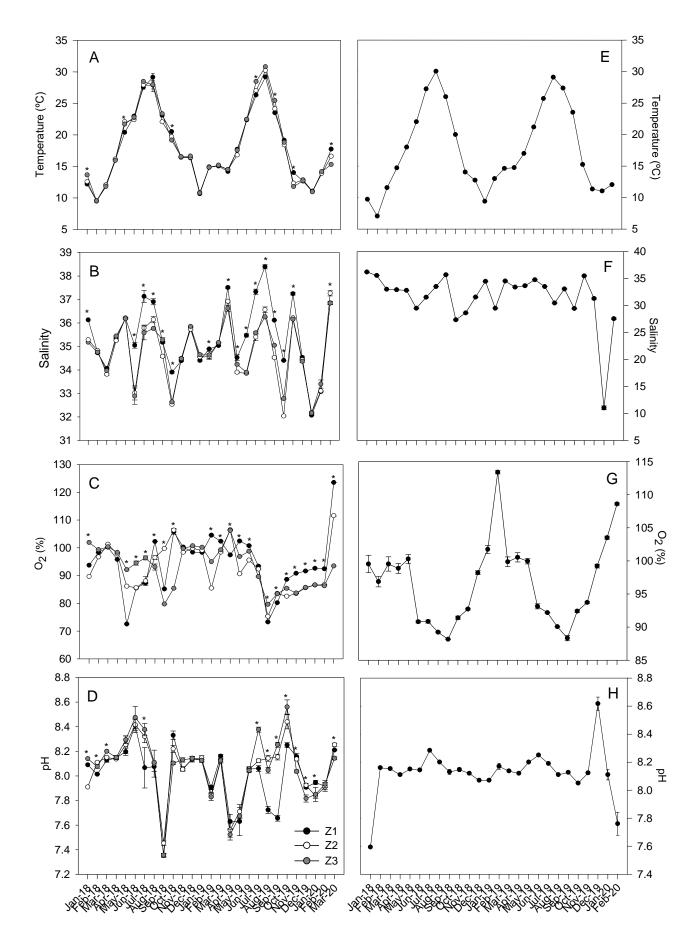


Fig. 6

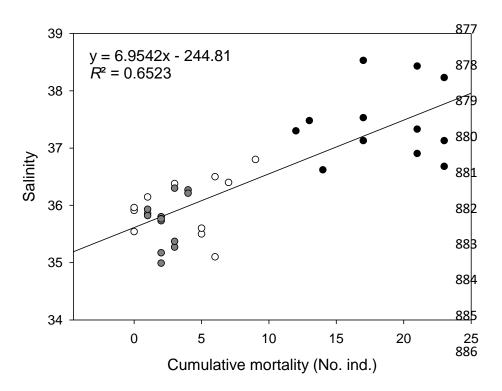


Fig. 7

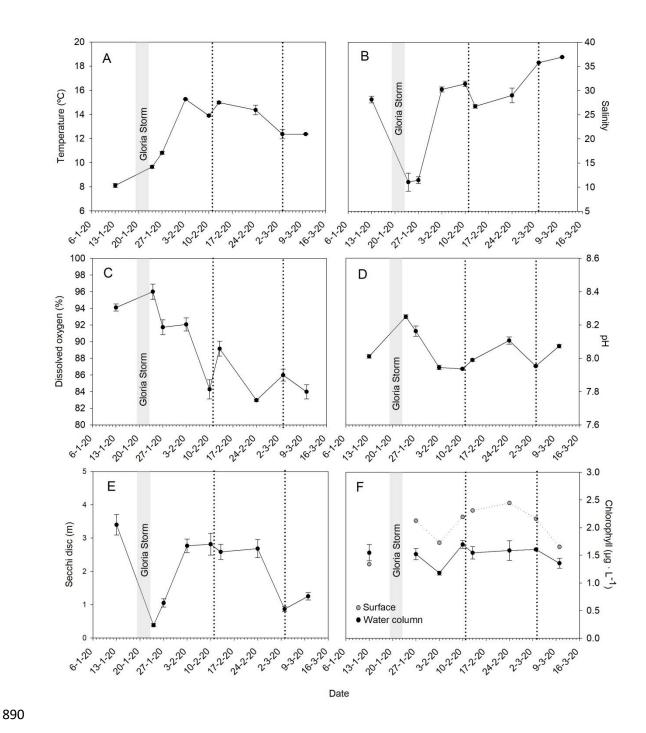


Fig. 8

Table 1. Mortality patterns in Alfacs Bay. A) One-way RM-ANOVA for monthly cumulative mortality of pen shells during the study period (January 2018 to March 2020) showing differences among sampling times (within subjects) and zones (among subjects). Since the Sphericity assumption was not meet, the Greenhouse-Geisser correction was used. B) Two-way RM-ANOVA testing for differences in the abundance of living and dead individuals of *Pinna nobilis* (juvenile and adults) in the impacted area of the bay (Zone 1 area closer to the mouth of the bay). D= Dead, A= Alive; Significant results are indicated in **bold**.

- 14							
Alfacs Bay							
A) One-way-RM-ANOVA	Type III SS	df	MS	F	p		
Time (Ti)	11.19	1.20	9.31	164.03	0.000		
Ti x Z	13.38	2.40	5.57	98.07	0.000		
Error (Ti)	0.409	7.207	0.057				
Zone (Z)	37.74	2	18.87	68.26	0.000		
Error (Z)	1.65	6	0.276				
LSD (Ti)	J18-Jun18> Jul 18> Aug18-Jun19> Jul19> Aug19>Sep19-Mar20						
LSD (Z)	Z1> Z2> Z3						
B) 2-way RM-ANOVA	Type III SS	df	MS	F	р		
Time (Ti)	0.00	1	0.00	0.00	1.000		
Ti x A	0.00	1	0.00	0.00	1.000		
Ti x IC	9.80	1	9.80	50.28	0.000		
Ti x A x IC	9.80	1	9.80	50.28	0.000		
Error (Ti)	30.40	156	0.195				
Age (A)	7.81	1	7.812	1.43	0.233		
Ind. condition (IC)	221.11	1	221.11	40.53	0.000		
AxC	4.51	1	4.51	0.82	0.364		
Error	850.95	156	5.45				
LSD (IC)	D> A						

Table 2. Mortality patterns in Fangar Bay. A) RM-ANOVA for monthly cumulative mortality of pen shells during the study period from January 2018 to March 2020 (within subjects' effects). Since the Sphericity assumption was not meet, the Greenhouse-Geisser correction was used. B) RM-MANOVA testing for differences in the abundance of individuals of different conditions (recently dead and living juvenile and adults, and the presence of old shells) in the four zones of Fangar Bay at two times after Storm Gloria. LA= Living Adults, RDA= Recently Dead Adults, LY= Living Young specimens, RDY= Dead young specimens, OS= Old Shells. Statistically significant results are indicated in **Bold**.

							911
Fangar	Вау						912
A) RM-ANOVA		Type III SS		df	MS	F	<i>p</i> 913
Time (Ti)		3.853		1.112	3.40	66 93.138	09 00 7
Error (Ti)		0.083		2.223	0.03	37	915
Error		1.8	83	2	0.94		916
LSD (Ti)		Feb 2020> All months		T-	917		
B) RM-MANOVA		Type III SS	df		k's λ	F	9 <u>18</u> p
Time (Ti)		. , pc 55	3	0.3		36.987	09990
Ti x Z	,		8	0.19		18.728	0.000
Zone	LA	836.30	3	278	3.77	18.89	09200
	RDA	3711.42	3	123	37.14	15.01	0.000
	LY	5.69	3	1.8	9	53.86	09000
	RDY	87.12	3	29.	04	16.95	0.000
	OS	3500.88	3	116	6.96	21.27	09990
Error	LA	1079.13	73	14.	78		022
	RDA	6015.55	73	82.	40		923
	LY	2.57	73	0.0	_		924
	RDY	125.02	73	1.7			524
	OS	4005.00	73	54.	86		025

Table 3. Two-way ANOVA (Alfacs) and one-way ANOVA (Fangar), testing for differences in the physicochemical variables of the bays. A) Temperature (°C), b) salinity, c) dissolved oxygen (%), d) pH. In SNK of Alfacs Bay, significant differences among study zones are indicated. For significant temporal variability (SNK) see Fig. 5. Statistically significant results are indicated in **Bold.**

									033
ANOVA	Alfacs			Fangar				934	
a) Temp	df	MS	F	р	e) Temp	df	MS	F	<i>p</i> 935
Date= D	25	315.78	3454.0	0.000	Date= D	25	139.60	8181	0 ₉ ცც0
Zone= Z	2	0.27	0.0	0.992	Error	52	0.02		937
ZxD	51	0.91	9.9	0.000					938
Error	162	0.09							939
b) Salinity	df	MS	F	р	f) Salinity	df	MS	F	<i>p</i> 940
Date= D	25	15.6	495	0.000	Date= D	25	71.87	1547	0<u>9</u>0<u>0</u>0
Zone= Z	2	10.4	6	0.004	Error	52	0.05		942
ZxD	51	0.7	21	0.000					943
Error	162	0.1							944
SNK (Z)	Z1> Z	Z2= Z3							945
c) DO	df	MS	F	р	g) DO	df	MS	F	<i>p</i> 946
Date= D	2	94	1.18	0.308	Date= D	25	118.4	72.9	09000
Zone= Z	25	505.69	95.89	0.000	Error	52	1.6		948
ZxD	51	102.42	19.42	0.000					949
Error	162	5.27							950
d) pH	df	MS	F	р	h) pH	df	MS	F	<i>p</i> 951
Date= D	25	0.452	55.43	0.000	Date= D	25	0.088	1.62	09 52 1
Zone= Z	2	0.06	0.9	0.406	Error	52	0.054		953
ZxD	51	0.027	3.41	0.000					954
Error	162	0.008							955
									956