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- 1 Banker plants and landscape composition influence colonisation precocity of tomato
- 2 greenhouses by mirid predators
- 3 Agnès Ardanuy^{1,2*}, Martí Figueras³, Montserrat Matas⁴, Judit Arnó³, Nuria Agustí³, Òscar
- 4 Alomar³, Ramon Albajes¹, Rosa Gabarra³
- ¹Universitat de Lleida, Agrotecnio Center, Rovira Roure 191, 25198 Lleida, Spain; ²School of
- 6 Earth and Environmental Sciences, The University of Manchester, Oxford Road, M13 9PT,
- 7 Manchester, United Kingdom; ³IRTA, Ctra Cabrils km 2, 08348 Cabrils (Barcelona), Spain;
- 8 ⁴ADV del Baix Maresme, Ctra NII km 639, Mercat de la Flor, 08340 Vilassar de Mar
- 9 (Barcelona), Spain.
- *Corresponding author: agnes.ardanuy@gmail.com

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Key Message

- Effective biological control occurs when predators colonise crops and pests are low.
- We tested which greenhouse elements encourage the colonisation of tomato by mirid
 predators.
- Herbaceous habitats promoted early colonisation of tomato by *Macrolophus pygmaeus*.
 - Calendula banker plants favoured early colonisation by M. pygmaeus.
 - Banker plants are a key low-cost tool to foster biological control in tomato.

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Abstract (150-200 words)

31 Conservation biological control involves manipulation of the environment to enhance the 32 effectiveness of natural enemies in controlling crop pests. In this study we combined historical 33 data, sticky trap sampling of tomato greenhouses and beat-sampling of adjacent vegetation to identify which greenhouse characteristics, habitat management practices and landscape features 34 favour an early colonisation of tomato greenhouses by the key mirid predator Macrolophus 35 36 pygmaeus and its establishment in NE Spain. Results show that landscape composition and the 37 use of Calendula officinalis banker plants inside the greenhouse are key factors. In general, greater amounts of herbaceous semi-natural cover at the landscape scale promoted M. pygmaeus 38 39 colonisation; while the use of C. officinalis banker plants encouraged M. pygmaeus colonisation independently of the landscape context. We identified host plants adjacent to tomato 40 41 greenhouses that sustain M. pygmaeus populations, however, they did not have a major effect on 42 M. pygmaeus colonisation compared to larger landscape and banker plant effects. Early 43 colonisation of greenhouses by this predator species also translated into lower accumulated incidence of pests at the end of the sampling period. This study demonstrates the importance of 44 45 active habitat management practices in promoting the early arrival of M. pygmaeus in 46 greenhouses with delayed spontaneous colonisation.

Keywords

Tomato, colonisation, banker plants, landscape, *Macrolophus*, biological control

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Introduction

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Conservation biological control (CBC) involves manipulation of the environment to enhance the 53 54 survival, fecundity, longevity, and behaviour of natural enemies to increase their effectiveness 55 in controlling crop pests. The key to an effective CBC is an early colonisation and establishment of natural enemies in a crop, when pest populations are still at low densities (Wiedenmann and 56 Smith 1997; Symondson et al. 2002). This can be particularly complex in the case of ephemeral 57 58 habitats, like annual crops, as it requires natural enemies to successively disperse between crops and alternative habitats following a seasonal cycle (Wissinger 1997; Tscharntke et al. 2007; 59 60 Schellhorn et al. 2014). Thus, for CBC to become a reliable pest management strategy it is necessary to understand which factors are involved in promoting natural enemy colonisation 61 and early establishment in crops. Early establishment is particularly relevant for spring 62 greenhouse crops as they grow in a climate that is favourable for the fast development of pest 63 64 populations (Albajes and Alomar, 1999). Alternative host plants within agricultural landscapes 65 provide key resources to natural enemies (such as alternative prey and host, nectar and pollen) 66 and can directly influence native natural enemy population dynamics (Landis et al. 2000; Norris 67 and Kogan 2005; Gurr et al. 2017). Promoting host plants at either the field, the farm 68 (Letourneau et al. 2011) or the landscape level (Bianchi et al. 2006) may therefore foster natural enemy spill over to crops by minimizing the distance between crops and alternative hosts, while 69 70 maximizing the overlap of host plant resources in time (Wissinger 1997; Schellhorn et al. 2015). 71 Biological control is a key strategy of pest controlling protected tomato crops in the 72 Mediterranean region (Perdikis et al. 2011; Arnó et al. 2018). Pest management strategies on 73 tomato greenhouses have relied for many years on inoculative releases of commercial natural 74 enemies (Messelink et al. 2014). However, the early findings of spontaneous colonisation of 75 unsprayed greenhouses by native natural enemies (see review by Arno et al 2018) led to an 76 increased interest on CBC as a promising tool to increase the sustainability and profitability of 77 protected tomato crops. Among the native natural enemies on tomato crops in the Mediterranean region, polyphagous predatory mirid bugs (Heteroptera: Miridae) of the genera Macrolophus, 78 79 Dicyphus, and Nesidiocoris have proved very successful in controlling whiteflies and other key 80 tomato pests (Lykouressis et al., 2001; Alomar et al., 2002; Gabarra et al., 2004; Ingegno et al., 2009; Lenteren et al., 2020). Polyphagy has been considered an advantage because it encourages 81 82 the early establishment of the predators in the crop when the target pest is still at low densities, 83 and it allows predators to sustain their populations in the crop once biological control of the target prey has been achieved (Albajes and Alomar 1999; Symondson et al. 2002; Castañé et al. 84 85 2016).

87 Currently, the predator *Macrolophus pygmaeus* (Rambur) is the focus mirid species in tomato crops in NE Spain and France given the prevalence of their naturally occurring populations 88 89 relative to other mirid bugs in the region, their persistence in low prey density patches 90 (Montserrat et al. 2004), and more importantly because this species does not produce damages 91 in open tomato greenhouses (Castañé et al. 2011; Arnó et al. 2018). Nevertheless some 92 controversy still exists about its use in other geographical regions (Sanchez et al. 2018). An 93 appropriate management of *M. pygmaeus* populations in tomato greenhouses may save many 94 inoculative releases by sustaining predator populations through the different tomato crop cycles 95 around the year. Several studies have related the presence of adjacent host plants to adequate 96 colonisation of tomato fields by predatory mirids (Alomar et al. 2002; Gabarra et al. 2004; 97 Ingegno et al. 2009). This, however, might not be the only factor, as good levels of colonisation 98 of tomato crops by mirid bugs have also been observed for crops with no adjacent host plants 99 present (<75 m) (Alomar et al. 2002). A likely explanation for these observations is that mirid 100 predators colonise tomato crops both from local host plant sources (field scale), and from more 101 distant sources (farm and landscape scale). 102 The common marigold, Calendula officinalis L. (Asteraceae), is one of the main host plants of 103 M. pygmaeus and it has been proposed as a banker plant to preserve populations of M. 104 pygmaeus between crop cycles and/or to provide an on-farm refuge for spontaneous populations 105 (Alomar et al. 2006; Messelink et al. 2014; Balzan 2017). Planting C. officinalis strips in crop 106 edges as a banker plant has become a CBC strategy in northern Spain and the south of France 107 (Lambion, 2014; Arnó et al., 2018; Agustí et al., 2020), and this practice has been related to 108 lower tomato leaf damage by Lepidoptera (Balzan 2017). However controversy remains given 109 the potential of C. officinalis plants to also sustain the mirid Nesidiocoris tenuis (Reuter), which 110 is known to inflict damage to the crop when present at high densities (Castañé et al. 2011). 111 Apart from marigold, other non-crop plants have been identified as overwintering refuges for 112 these predatory mirid bugs (e.g. Lykouressis et al., 2001; Alomar et al., 2002; Ingegno et al., 113 2009), together with tomato, eggplant and potato crops. To date, only one study to our knowledge has studied the effects of landscape composition and 114 115 configuration on mirid predator populations in protected tomato crops (Aviron et al. 2016). Key 116 findings showed greater levels of *M. pygmaeus* colonisation in greenhouses embedded in 117 landscapes with larger fallow area, while colonisation was reduced in greenhouses associated 118 with larger orchard area at the landscape scale. Yet, these landscape effects on mirid 119 colonisation were smaller than management practices associated to organic and conventional 120 agriculture. Further research is needed to understand the relative importance of habitat 121 management practices, e.g. presence of banker plants and other crops and non-crop host plants

(Thomine et al. 2020), and the landscape context on CBC by mirid predators in protected tomato crops.

This study aims to identify which greenhouse characteristics, habitat management practices, and landscape features favour an early colonisation of protected tomato crops by mirid predators and their establishment. We hypothesized that (i) mirid predator colonisation precocity is stable through time for each of the studied greenhouses e.g. showing consistently early vs late colonisation; (ii) larger proportion of semi-natural non-crop cover at the landscape scale enhance mirid colonisation by promoting spillover to tomato crops early in spring, and (iii) greenhouse habitat management practices, using *Calendula* banker plants inside the greenhouse and/or by maintaining diverse host plants at the farm scale, favour early colonisation of tomato greenhouses by mirid bugs.

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Methods

Study site and greenhouse selection

The study was carried out in the Maresme county, NE Spain, where protected tomato is a key horticultural crop. The region is located NE of Barcelona following the coastline and between the Litoral mountain range and the sea. Land-use in the area is traditionally agricultural with increased urban pressure over the last 40 years. Natural and semi-natural habitats in the region are comprised of woodland, shrub, herbaceous and ephemeral stream habitats. Twelve greenhouses from a Grower's association (Associació de Defensa Vegetal del Baix Maresme, ADV) and supervised by a plant protection advisor were used for the study. Tomato seedlings were transplanted ranging from mid February to early April. Greenhouses were characterized by their structure (wood/metallic), openings (lateral/zenital), date of transplant (February/March/ April), crop diversity (tomato/mixed), number of tomato varieties (one/more), altitude (sea level/>50m), and size (Table 1); together with the use of Calendula banker plants inside the greenhouse (5 greenhouses with banker plants and 7 without, Table 2). In 4 out of the 5 greenhouses with banker plants, Calendula plants were established before the previous tomato spring season (> 1 year before the start of the experiment) while in one of the greenhouses a banker plant margin was established in February just before the tomato transplant. Pest management strategies in the selected greenhouses were based on the recommendations of the plant protection advisor following the IPM rules developed for tomato crops in the area (Arnó et al. 2018).

Historical data collection

To confirm previous observations that some greenhouses consistently showed earlier colonisation and pest control than others over the years (Castañé et al. 2004), plant health monitoring data were extracted from the plant protection advisor reports for the same season of our sampling (see next section) and the four previous seasons. For greenhouse H9 data on previous seasons were not available, since that greenhouse was not managed by the ADV at that time. Data extracted were: transplant date; date of the first observation of *M. pygmaeus* adults on tomato plants (colonisation precocity); date of the first observation of *M. pygmaeus* nymphs; and pest/disease control treatments and their dates. All historical data colonisation dates were expressed as Julian days. Julian day is a date expresses as the number of days that have passed since the 1st of January of each year. Meteorological data were retrieved from RuralCat (https://ruralcat.gencat.cat/agrometeo, Generalitat de Catalunya). Accumulated rainfall was obtained for the winter-spring period (December-April) and degree-days above 10 °C (DD₁₀) were calculated for the spring period (January-April). The decision to use 10°C for the calculation of DD was based on the lower thermal thresholds for *M. pygmaeus* eggs and nymphal development (Martínez-García et al. 2017).

Arthropod sampling in tomato greenhouses

Tomato greenhouses were sampled every other week with yellow sticky traps from mid-March to mid-June, with the intention to detect the arrival of mirid bugs in each greenhouse and estimate the amount of prey present. Greenhouses were sampled five times during this period, with the exception of two greenhouses that were only sampled four times because of delayed transplanting. Yellow sticky traps (31x21 cm, Entomopraxis, Barcelona, Spain) were attached to wooden sticks and placed between tomato plants along the crop rows. Trap height was adjusted to be at the same level as the top of the plant canopy at the early stages of tomato growth, and at 1.20 m in full grown plants. Nine yellow sticky traps were used per greenhouse and sampling date, and were evenly distributed in the greenhouse to cover its surface. Sticky traps were recovered one week later. Traps were then covered by plastic film and placed in a cold climatic chamber (4 °C) until processed. Mirid predator species and key pest groups (whiteflies, aphids, leafmining diptera and thrips) were later identified to taxonomic units and counted. Colonisation precocity was determined as the number of sampling event in which the first mirid was captured, e.g if a mirid predator was first detected on the third sampling event the colonisation precocity of the greenhouse was assigned to 3.

Predatory mirid surveys in adjoining vegetation and Calendula banker plants

The abundance and composition of mirid species on plants adjacent to the greenhouse and on banker plants inside the greenhouse was determined three times for early planting greenhouses and two times for late planting greenhouses during the sampling period. The vegetation was sampled every 10 m around the outer perimeter of the tomato greenhouses whenever plants were present. For each sampling point, approximately a 0.5 m² vegetation area was beaten three times with a bat and insects were collected on a white plastic tray (DIN-A4 size). Adult and nymph mirid bugs were collected by means of a mouth aspirator and were placed in tubes in a cooling box to avoid predation. Plants present in each sampling point were identified to genus level in most cases. Whenever a sampling point had more than one plant species, the proportion of each plant species in the mixture was visually estimated. If Macrolophus spp. were collected in points with plant mixtures, all plant species present in the mixture were individually resampled to be able to relate a particular *Macrolophus* species to the host plant (see next section). Calendula banker plants inside the greenhouses were sampled at the same sampling dates and in the same manner than the adjoining vegetation. Back in the laboratory, nymphs were placed in boxes containing a green bean pod, Ephestia kuehniella Zeller (Lepidoptera: Pyralidae) eggs and a water supply to complete their development to adults and allow further identification to species level. Adult individuals were morphologically classified as either *Macrolophus spp.*, Nesidiocoris tenuis, Dicyphus bolivari Lindberg or Dicyphus errans (Wolff). As the number of sampling events differed between greenhouses, only the last two samplings of adjacent vegetation were considered (early and late April). Variables related to habitat management (listed in Table 2) were used in further analysis. Among those, the abundance of *Macrolophus* spp., the number of sampling points, and the identified host plant species may indirectly represent a farmer's habitat management practices outside the greenhouse, e.g. herbicide treatments, conservation of weedy margins and active encouragement of plant diversity at the farm scale.

Macrolophus spp. molecular identification

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- Two sympatric *Macrolophus* species present in the area, *M. pygmaeus* and *M. melanotoma*,
- 215 (Costa) have a great morphological similarity, and have often been confused (Perdikis et al.
- 216 2003; Martinez-Cascales et al. 2006; Castañé et al. 2013). Yet only *M. pygmaeus* is known to
- establish on tomato crop; for this reason, we used molecular markers to distinguish the two
- 218 Macrolophus species and to find out to what extent both species coexisted on the sampled non-
- 219 crop host plants. Subsamples of adult and nymphs of Macrolophus spp. specimens collected in
- each host plant were identified using conventional PCR using the specific primers Mp1F (5'-
- 221 GTAACATAGATAAAATCCCATTTC-3') Mp4R.2 (5'-
- 222 CCTAATAATTGTGGTTCTCACAA-3') for M. pygmaeus, and Mm1F (5'-

- 223 CTTCTTGATGCCTTTTATTGTGGC-3') Mm3R (5'-
- 224 TTATCATACCTATGTAGTCCTTGATT-3') for *M. melanotoma*. These primers were
- previously described in Castañé et al. (2013), with the exception of Mp4R.2, which is a
- 226 modification of Mp4R described there. Individual insects were DNA extracted using the
- 227 SpeedTools Tissue DNA Extraction Kit (BioTools; Madrid, Spain) following the
- 228 manufacturer's protocol. PCR reactions were conducted as described in Castañé et al. (2013).
- Target DNA (*M. pygmaeus* and *M. melanotoma*) and water were always included as positive
- and negative controls, respectively. Resulting PCR products were separated by electrophoresis
- in 3.5% agarose gels, stained with ethidium bromide and visualised under UV light. The number
- of individuals analysed per host plant species varied from 9 to 15 (Table 3).

Landscape cover characterization

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- Land use composition surrounding each greenhouse was quantified using a circular buffer area
- at two scales (250 and 150 m) with ArcGIS 9.3 (ESRI, 2005). The radius was chosen according
- 236 to the results reported by previous biological control studies on M. pygmaeus (Alomar et al.
- 237 2002; Aviron et al. 2016). The land use in these landscapes was mapped based on geo-
- 238 referenced aerial photographs and available data on cover types SIGPAC (Institut Cartogràfic i
- Geològic de Catalunya, http://www.icc.cat/vissir3/). During field inspections, landscape patches
- 240 were classified as crop (including the three cover classes: protected horticulture, open field
- agriculture, and olive/vineyard trees) or non-crop (including urban and four semi-natural habitat
- cover classes: herbaceous, shrub, woodland, and riparian). Land use was verified in the terrain,
- and corrections were made during the digitalization process. The proportions of all cover classes
- in each landscape buffer were calculated for all the greenhouses used in the study.

Statistical analysis

- 246 Historical data was used to establish the importance of yearly variation and greenhouse identity
- 247 on predator colonisation precocity, and the relationship of this precocity with their
- establishment in tomato greenhouses. First, a linear model was fitted with log transformed M.
- 249 pygmaeus precocity (expressed as Julian day) as dependent variable; and the year, greenhouse,
- 250 transplant date and their interaction as independent variables. Then, meteorological variables
- 251 were used to explore whether they could help explain the yearly variation in colonisation by
- 252 replacing factor year in the previous model by the variables spring accumulated degree days
- 253 (DD₁₀) and accumulated winter-spring rainfall. Pair-wise comparisons were evaluated with
- Tukey's post hoc test with Bonferroni correction. Second, a linear model was fitted with log
- 255 transformed M. pygmaeus nymph detection date as dependent variable and adult M. pygmaeus

precocity as explanatory variable. In order to do that, data points where nymphs were detected before adults were excluded from analysis.

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To evaluate whether yellow sticky traps could be reliable in detecting *M. pygmaeus* colonisation of tomato greenhouses for the season when the study was conducted, a linear model was built with the first detection of *M. pygmaeus* adults by yellow traps as dependent variable (approximate Julian day), and the first detection of *M. pygmaeus* adults by plant sampling as an independent variable. As sticky traps were placed twice a month and left in the field for a week, approximate Julian days were calculated.

To test whether mirid colonisation precocity and abundance were associated with greenhouse characteristics (Table 1), habitat management (Table 2) or landscape composition, models were fitted with mirid captures on yellow sticky traps as the dependent variable. Analyses of mirid precocity and abundance were focused on M. pygmaeus, given that D. bolivari adults were only detected in two of the 12 greenhouses at the end of the sampling period. Linear models were used for M. pygmaeus colonisation precocity, and generalized linear models following a negative binomial distribution and log link were used for abundance to account for overdispersion of the data. First, in order to establish the importance of a given predictor variable for explaining colonisation or population abundance, separate models were fitted for each landscape composition variable at two spatial scales (150m and 250m from the greenhouse, landscape cover class variables are described in previous section and listed in the Supplementary material Fig S1 and Fig S.2), and for each greenhouse habitat management variable (Calendula banker plant presence and other numerical variables obtained by adjoining vegetation sampling and listed in Supplementary material Fig S3) and greenhouse characteristics (all variables in Table 1). In the case of mirid abundance models, the variable mirid colonisation precocity was also used as an explanatory variable. As multicollinearity can influence the interpretation of models results Spearman coefficients between each set of numerical variables were calculated to establish the relationship between them (Supplementary material Fig. S1, Fig. S2 and Fig. S3). Numerical adjoining vegetation variables were highly correlated (>0.7, Supplementary material Fig. S3) so only the total *M. pygmaeus* abundance in adjoining vegetation (variable Mp, Fig S3) was selected together with the categorical variable Calendula banker plants (Cal, presence/absence) as habitat management variables in further analyses. Then, mirid precocity and abundance were modelled using the best landscape predictors in interaction with each of the greenhouse characteristics and habitat management variables. This approach was followed to avoid a priori selection of the explanatory variables included in the analysis. All explanatory variables were standardized (mean = 0, SD = 1) which allowed the comparison of effect sizes between predictors.

Finally, in order to evaluate the effects of M. pygmaeus time of colonisation on pest abundance, a composite pest abundance index per greenhouse was used. This global pest abundance index was built by standardizing (0-1) the accumulated abundances of each pest taxon (whiteflies, thrips, aphids and leafminers) and adding their values (all pests had the same weight in the index). The composite pest abundance index, ranging between 0 and 4, is thus an aggregate measurement of pest pressure. This index was calculated for two times during the season: t_c time of detection of the first M. pygmaeus and t_f time of final sampling. A linear model was then fitted to evaluate whether colonisation precocity by M. pygamaeus and pest abundance index at time of colonisation (t_c) explained pest abundance index at the end of the sampling period (t_f).

Pest abundance index at t_f was log-transformed to achieve model assumptions.

All models were evaluated according to their performance based on Akaike's Information Criterion for small samples sizes (AICc) following Burnham & Anderson (2002). Briefly, the best model with the lowest AICc value was identified together with any competing model with $\Delta AICc < 2$ that was considered as receiving equal support from the data (including the null model). For all models R^2_{adj} (linear model) or D^2_{adj} (glm negative binomial model, Guisan & Zimmermann, 2000), predictor estimates, and their interval of confidence (95%) were calculated. Assumptions of linearity and homogeneity of variances on residuals from the best models were checked graphically. No spatial autocorrelation was detected for the residuals of the best models (Moran's I statistic, Ape package; Paradis & Schliep, 2019). All statistical analyses were performed using R version 3.5.2 (R Core Team, 2018) and figures were produced using the package ggplot2 (Wickham 2009).

Results

Historical mirid colonisation and establishment

The main mirid predator species colonizing spring tomato protected crops in the study area was *M. pygmaeus*. In only two occasions in a five-year period, *M. pygmaeus* adults were not detected in tomato crops, and in both cases the farmer had sprayed insecticides (spinosad and flubendiamide, as stated by the plant protection advisor report). In general, only *Bacillus thuringensis* Berliner (Bt) based insecticides were applied before colonisation by *M. pygmaeus* when infestation levels of tomato plants by the leafminer *Tuta absoluta* (Meyrick) were high.

Linear models showed that greenhouse identity ($F_{10,41}$ =4.66, p<0.0001) was the main effect accounting for early colonisation of tomato crops by M. pygmaeus together with DD_{10} ($F_{1,39}$ =4.94, p=0.02) and accumulated spring rainfall ($F_{1,39}$ =3.74, p=0.03), with a R^2_{adj} =0.51 and

- lowest AICc (-14.96). Models including tomato transplant date had greater AICc values and were less parsimonious. The combination of the meteorological predictors explained better the seasonal variation in predatory mirid colonisation than the factor year. Colonisation precocity by *M. pygmaeus* differed between greenhouses (Fig. 1A), with greenhouses H3 and H10 representing the early (mid March) and late (mid May) colonisation extremes respectively. In general, years with warm and wet winter and spring seasons showed earlier colonisation of protected tomato crops by *M. pygmaeus* than colder and drier years.
- 331 The detection of the first nymphs of M. pygmaeus in greenhouses was correlated with the detection of the first adults (R^2_{adj} = 0.57, Fig. 1B), indicating that the establishment of this predator in a tomato greenhouse depended on the time of arrival of the first colonisers. On average the first nymphs were detected four weeks after the first adult detection.

Predatory mirid surveys in adjoining field margins and Calendula banker plants

A total of 435 *Macrolophus* spp. and 60 *Dicyphus* spp. specimens were collected from the vegetation adjoining the tomato greenhouses (Table 2), while 740 *Macrolophus* spp. and 43 *Dicyphus* spp. were collected from *Calendula* banker plants located inside the tomato greenhouses. Some individuals of *N. tenuis* were collected on *Calendula* banker plants in the two greenhouses where this species had been released years earlier as part of a crop protection strategy (H12 historical data, and H10 personal communication from the farmer), and one nymph was captured in H1 (Table 2). Few individuals were also collected on *C. officinalis* plants adjoining two of the greenhouses with no known history of *N. tenuis* releases (Table 2).

Almost all *Macrolophus* spp. specimens sampled from beat sheet sampling were identified as *M. pygmaeus* by molecular analysis (Table 3). In eight of the plant species surveyed, both adults and nymphs of *M. pygmaeus* were detected, which indicates the ability of this predator to reproduce and feed on those plants and can therefore be considered confirmed host plants. Confirmed host plants for *M. pygmaeus* comprise both ornamental plants (and therefore intentionally or unintentionally part of the farm habitat management practices) and weeds. The number of host plant taxa around to each greenhouse ranged from 0 to 9 (Table 2, mirid host plant species breakdown for each greenhouse can be found in Supplementary material Table S1). Overall *C. officinalis* plants harboured the largest *M. pygmaeus* populations in comparison with the other seven host plants. Aromatic ornamentals *Lavandula* spp. and *Salvia* spp., and other ornamentals like *Dimorphotheca ecklonis* (DC.), sustained considerable populations of *M. pygmaeus* (Table 3). Weeds like *Erodium* spp., *C. arvensis*, *P. officinalis* and *Borago* spp. were also identified as hosts for *M. pygmaeus*. The cryptic *M. melanotoma* was only detected in two host plants in very low numbers: *D. viscosa* (2 adults) and *Lavandula* spp. (1 nymph) (Table 3).

Landscape and greenhouse management effects on mirid colonisation and pest abundance

Data recorded with the two sampling techniques for one year, visual sampling and yellow sticky traps, associated with each other with an acceptable reliability ($R^2 = 0.46$, Fig. 1C). Given that sampling took place twice a month for both sampling methods and sticky traps were left in the greenhouse for 1 week, a mismatch of ± 15 days between methods was to be expected.

Colonisation precocity by *M. pygmaeus* was dependent on both the amount of herbaceous seminatural cover at the 250m buffer around greenhouses (PS₂₅₀) and the presence of *Calendula* banker plants (Cal) inside the greenhouses, as shown by the interaction of the two predictors (Table 4). In general greater amounts of herbaceous semi-natural cover favoured early colonisation of tomato greenhouses by *M. pygmaeus*; while the use of *Calendula* banker plants inside the greenhouses attenuated the negative effects of the low proportion of surrounding favourable habitats by promoting early colonisation of the tomato crop (Fig. 2). Landscape variables at 150m performed worse than those at 250m and hence were not included in the best or competing models. Likewise no other greenhouse characteristics or habitat management variables, apart from *Calendula* banker plant presence (Cal), were included in the best or competing models.

Accumulated abundance of *M. pygmaeus* adults at the end of the sampling period was explained by its colonisation precocity (Table 4): greenhouses with earlier colonisation had a greater number of predator adults than those with late colonisation. Pest abundance index at the end of the sampling period was explained jointly by *M. pygmaeus* colonisation precocity and the covariable pest abundance index at the time of *M.pygmaeus* colonisation t_c (Table 4), indicating a negative relationship between pest index and *M. pygmaeus* precocity.

Discussion

This study aimed at understanding how greenhouse characteristics, habitat management practices and landscape features favour early colonisation of protected tomato crops by mirid predators and their establishment. Historical data collected by the plant protection adviser showed that the identity of a greenhouse was central in determining time of colonisation by the most common predator, *M. pygmaeus*. This effect of greenhouse identity on colonisation precocity was further explored as a combination of 1) greenhouse characteristics, 2) habitat management, and 3) landscape composition. Results show that the key factors determining precocity in the colonisation of tomato crops by *M. pygmaeus* are landscape composition and habitat management by use of *Calendula* banker plants inside greenhouses. In general, greater

amounts of herbaceous semi-natural cover at the landscape scale promoted *M. pygmaeus* colonisation of tomato greenhouses; however, the use of *Calendula* banker plants encouraged *M. pygmaeus* colonisation independently of the abundance of herbaceous semi-natural cover. Early colonisation of greenhouses by mirid predators also translated into lower accumulated incidence of pests, and therefore potentially into a lower likelihood of pesticide application later in the season (Li et al. 2020). These findings encourage the use of banker plants as a key element for CBC in tomato protected crops in NE Spain.

Historical data confirmed that the main predator observed in the 5-year records was *M. pygmaeus* as it is usual in the area (e.g Castañé *et al.*, 2004). While meteorological factors - accumulated degree days above 10 °C (DD₁₀) and winter rainfall - partly explained yearly patterns in colonisation, it was the identity of the greenhouse that explained most of the variation in colonisation precocity by *M. pygmaeus*. Data sustained the idea that colonisation precocity of a particular greenhouse was consistent across years. Overall models showed no significant effects of greenhouse characteristics on *M. pygmaeus* colonisation precocity, whereas Aviron *et al.* (2016) detected strong effects of greenhouse crop management. This suggests that the greenhouses in the current study had relatively homogeneous crop practices resulting from the recommendations of a unique plant protection advisor, using practically no pesticides before mirid colonisation, while the former work compared greenhouses following a broad spectrum of practices grouped into organic vs. conventional. In addition, while the present study focused mainly on the first detection of predators in the crop as a proxy for colonisation, Aviron *et al.* (2016) focused on accumulated abundance which would potentially reflect in turn the cumulative crop protection practices in each greenhouse.

Colonisation precocity by *M. pygmaeus* in a greenhouse was best explained by the interactive effect of herbaceous semi-natural cover at the landscape scale and the use of *Calendula* as banker plant inside the greenhouses. Herbaceous cover enhanced the early arrival of *M. pygmaeus* adults to tomato crops in greenhouses without banker plants. These results are consistent with the study in SE France where greater abundances of this mirid species were observed in those greenhouses associated with greater fallow area (Aviron et al. 2016). The habitat cover type categorized as herbaceous semi-natural in this work comprises the continuum between semi-natural vegetation to non-cultivated former agricultural fields where ruderal vegetation predominates. When *Calendula* banker plants were used, the colonisation precocity of *M. pygmaeus* was independent of the abundance of herbaceous semi-natural cover at the landscape scale. This was a result of banker plants encouraging early colonisation in those greenhouses associated to small amounts of herbaceous semi-natural habitat. These findings agree with previous work in open agriculture and flower strip planting that indicate that

landscapes which have experienced greater damage from agricultural intensification have more to gain from habitat management practices (Thies and Tscharntke 1999; Haenke et al. 2009;

428 Jonsson et al. 2015).

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Banker plants can potentially be as effective as inoculative releases in delivering *M. pygmaeus* to tomato crops. Predatory *M. pygmaeus* have been reported to move from *Calendula* banker plants to tomato in a continuous flux of individuals 1.6 adults/m² every 3 days at an average planting of 4 tomatoes/m² (Agustí *et al.*, 2020). This represents densities consistent with the ranges of recommended commercial releases (0.25-0.5 adults/m² every 1-2 weeks for a total of 2-4 releases) (Moerkens et al. 2017). Promising results about the colonisation of tomato greenhouses by *M. pygmaeus* were also obtained using *Ballota hirsuta* Benth (Lamiaceae) as banker plants in SE Spain (Sanchez et al. 2020). In this study, the highest colonisation of tomato greenhouses by *M. pygmaeus* and pest control was obtained when *B. hirsuta* plants were deployed for 30 days as opposed to 1 day, as greater amount of founder *M. pygmaeus* individuals in tomato contributed to greater predator growth rates (Sanchez et al. 2020). Collectively these results indicate that the permanent establishment of banker plants in tomato greenhouses actively promotes early colonisation of the crop by *M. pygmaeus*, and this effect will be more notorious in greenhouses with limited spontaneous colonisation e.g. embedded in landscapes with small amount of herbaceous semi-natural habitat.

Another important aspect to be considered when studying the establishment of mirid bugs in a greenhouse is the distribution of colonisation sources within a greenhouse (e.g openings and banker plants) and how do predators distribute in the crop (Alomar et al. 2002; Gabarra et al. 2004). In a field study, Alomar et al., (2002) showed that adult M. pygmaeus were more abundant in outer tomato rows, particularly in fields with close predator host plant sources; while later in the season predator nymphs were distributed following adult predator or prey spatial pattern. The dispersal of commercial M. pygmaeus adults within a tomato greenhouse has been estimated to be <3 m from the release plant (Moerkens et al. 2017). Therefore, M. pygmaeus seem to exhibit a limited dispersal within the crop despite their ability to colonise tomato from semi-natural habitats present at < 300 m (this study; Aviron et al., 2016), which can be related to their ability to persist in low prey patches (Montserrat et al. 2004). In order to overcome this limitation and ensure a quick and even distribution of this predator in a greenhouse, banker plants should be ideally placed evenly spaced between tomato rows at 5m intervals (Moerkens et al., 2017; Agustí et al., 2020). However some caution is needed in the use of C, officinalis as a banker plant, since they are also hosts of N. tenuis, which has been shown to produce damage to tomato plants in some circumstances (Sanchez 2008; Calvo et al. 2009; Arnó et al. 2010). Given that the relative abundance of N. tenuis over M. pygmaeus has

increased in tomato crops in the area of study in the last years (RG, JA, J. Riudavets unpublished data), banker plants should be closely monitored during the season, and plant protection decisions on *Calendula* banker plant management will need to be taken on a pergreenhouse basis.

Apart from *C. officinalis*, other host plant species identified in this work and previous research (Alomar et al. 2002; Ingegno et al. 2009, 2011; Lambion 2014) can be favoured at the farm scale to sustain mirid populations through their life cycle. Although results reported in this work show that adjacent vegetation does not seem to be a major factor on *M. pygmaeus* colonisation in tomato greenhouses compared to stronger landscape and banker plant effects, host plant species richness can potentially provide insurance habitats to natural enemies at the farm scale (Tscharntke et al. 2007). For example these plants could constitute a refuge for mirid predators in agricultural intensive landscapes where crops with a high pressure of chemical sprayings, like orchards, are common (Aviron et al. 2016; Ricci et al. 2019; Clemente-Orta et al. 2020); and could act as a host "stepping stone" after the spring crop and contribute predators to adjacent open tomato crops later in the season.

Our findings provide insight into the local and landscape factors driving mirid colonisation in tomato greenhouses in NE Spain. This study demonstrates the importance of herbaceous seminatural habitats and the use of *Calendula* banker plants in promoting *M. pygmaeus* colonisation. Most importantly, results show that farmers with greenhouses surrounded by lower proportion of herbaceous semi-natural habitats can promote early *M. pygmaeus* colonisation of tomato through active habitat management by using banker plants. The use of banker plants in a farm is in control of the producer, as opposed to landscape management, and therefore it has the potential to become a key CBC strategy for protected tomato crops. Yet, both habitat management in farms by means of host plants and the conservation of semi-natural habitats beyond those individual farms might be needed to ensure that natural enemies persist over time (Schellhorn et al. 2015; Tooker et al. 2020), and for CBC to become a reliable crop protection strategy. Further research is needed to test how host plant conservation strategies can be used at the farm scale to facilitate timely colonisation of tomato and other crops by natural enemies through the farm's crop cycles.

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640	Ethics approval
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642	Availability of data and material
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645	Not applicable
646	Authors' contributions
647	AA, RG, RA, JA and OA conceived and designed research. NA optimized molecular
648	identification of Macrolophus species. AA and MF conducted experiments. MM contributed
649	with historical data collected as plant protection adviser and facilitated AA and MF access to
650	commercial greenhouses. RG supervised MF's Master Thesis. AA extracted historical data
651	analysed data. AA wrote the first draft of this paper and edited it based on significant comments
652	from RA, JA, OA, NA and RG. All authors read, improved and approved the manuscript.

Table 1. Characteristics of the 12 greenhouses sampled to study mirid colonization in tomato crops.

ID	Structure	Openings	Transplant	Crop diversity	Tomato varieties	Altitude	Area (m²)
H1	metal	zenital	8/2	tomato	one	>50m	4198
H2	wood	zenital/lateral	15/2	mixed	one	sea level	3846
Н3	metal	zenital/lateral	25/2	mixed	more	>50m	2491
H4	wood	lateral	3/3	mixed	more	sea level	1729
H5	wood	lateral	1/3	tomato	more	sea level	1323
Н6	metal	zenital	3/4	mixed	more	>50m	3266
H7	metal	zenital/lateral	8/3	mixed	more	>50m	3079
Н8	metal	zenital	6/3	mixed	one	sea level	1570
Н9	metal	zenital	1/3	tomato	more	sea level	5972
H10	wood	lateral	11/2	tomato	one	sea level	1608
H11	wood	lateral	15/3	tomato	one	>50m	1985
H12	metal	zenital	23/2	tomato	one	sea level	2387

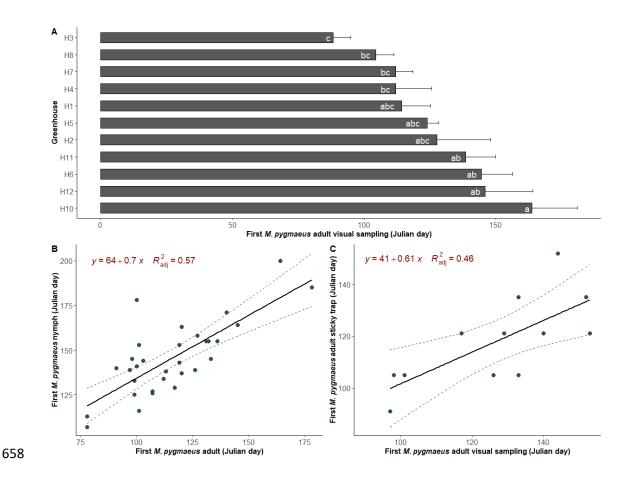


Fig 1. A) Day of the first *M. pygmaeus* adult detected (+SE) on tomato greenhouses in 5 consecutive seasons. Different letters indicate significant differences between groups (Tukey post-hoc test with Bonferroni correction, P<0.05). B) Relationship between the day of the first *M. pygmaeus* nymph detection with the day of the first adult detection by visual sampling. Data in which nymphs were recorded before or on the same day as adults have been removed. C) Relationship between the first detection of *M. pygmaeus* in the greenhouse when two different sampling methodologies were used in one sampling season: visual sampling and yellow sticky trap sampling. In all cases the first detection of *M. pygmaeus* is expressed as Julian days. The dashed lines represent 95% confidence limits.

Table 2. Summary of the number of mirids recovered from beat sheet sampling in (a) vegetation adjoining tomato greenhouses and (b) *Calendula* banker plants inside the greenhouses (when present). Greenhouses are numbered according to the total *Macrolophus* spp. recovered from the adjoining vegetation. In (a) *Macrolophus* spp. are divided into those recovered from *C. officinalis* and from other host plants. A plant taxa is considered as host when both adults and nymphs of a mirid species were recovered (includes host plants from all mirid species). The list of host plants adjacent to each of the greenhouses can be found in Supplementary material Table S1. Also in (a), the mirid host plant richness and their abundance in the adjoinining vegetation is shown. Richness refers to the number of host plant species present in the adjacent vegetation of each greenhouse. Points with host plant presence refers to the number of sampling points with confirmed host plants around the greenhouse perimeter. Potential points refers to the potential sampling points at 10m intervals based on the greenhouse perimeter. *Data on *Dicyphus* spp. includes *D. bolivari* and *D. errans*.

						G	reenho	use ID					
a) Adjoining vegetation	H1	H2	Н3	H4	Н5	Н6	H7	Н8	Н9	H10	H11	H12	
Macrolophus spp.	Total	175	145	52	28	12	12	7	3	1	0	0	0
	C. officinalis	150	145	0	0	0	0	0	0	0	0	0	0
	Other host plants	25	0	52	28	12	12	7	3	1	0	0	0
Other mirids	Dicyphus spp.*	15	9	28	0	1	7	0	0	0	0	0	0
	N. tenuis	0	3	0	0	0	0	0	0	0	8	0	0
Host plant (HP)	Richness	8	2	9	3	2	3	5	7	2	0	0	0
• • •	Points with HP presence	9	17	23	18	4	16	6	9	1	0	0	0
	Potential points	36	31	32	25	22	30	28	23	41	22	26	28
b) Banker plants													
Macrolophus spp.	Total	157	177					324			52		25
Other mirids	Dicyphus spp.*							4			38		1

N. tenuis 1 0 0 15 6

Table 3. Total number of Macrolophus spp. and Dicyphus bolivari individuals per host plant taxa obtained from beat sheet samples in vegetation surrounding tomato greenhouses. The total number of M. pygmaeus and M. melanotoma identified by PCR from subsamples of each host plant is also shown. n = n0 number of greenhouses with the host plant present in the adjoining vegetation.

			Ма	Dicyphus bolivari					
Host plant	n	Adults	Nymphs	M. pygmaeus (PCR)	M. melanotoma (PCR)	n	Adults	Nymphs	
Borago officinalis	2	8	1	9		1		1	
Calendula arvensis	3	7	5	10					
Calendula officinalis	5	122	255	15		4	5	30	
Dimorphoteca ecklonis	1	4	18	9					
Dittrichia viscosa	1		2		2				
Erodium spp.	3	21	11	9		3	5	15	
Gallium spp.	1	1							
Lavandula spp.	2	4	29	12	1				
Leucanthemum spp.						1		1	
Malva spp.	1	1				2	5	4	
Parietaria spp.	7	37	39	10		3	1	5	
Salvia officinalis	1	8	29	10		1		2	
Sonchus spp.	2	1	1						

Table 4. Estimates, confidence intervals (CI), AICc and model fit for the best models (with lowest AICc) with (a) colonization precocity and (b) abundance of *M. pygmaeus* in tomato greenhouses, and (c) the pest index at the end of the sampling period as dependent variables. All selected variables in the models are relevant as their CI do not overlap with 0. Colonization precocity is expressed as the sampling event in which the first *M. pygmaeus* adults were detected in each greenhouse by means of yellow traps, with smaller values representing early colonization.

		Selected	Estimat	CI	CI	AIC	R^2_{adj}/D^2a
		Variables	e	(2.5%)	(97.5%)	c	dj
						32.7	
a)	Colonization precocity	Intercept	4.24	3.8	4.68	1	0.796
		PS250	-1.21	-1.66	-0.76		
		Cal	-0.87	-1.55	-0.19		
		$PS250 \times Cal$	0.90	0.18	1.61		
						72.5	
b)	Abundance	Intercept	1.42	0.84	1.98	6	0.712
		Precocity	-1.57	-2.23	-1.02		
						31.9	
c)	Pest index t _f	Intercept	-0.47	-0.85	-0.08	9	0.622
		Pest index t _c	1.12	0.55	1.7		
		Precocity	-0.65	-1.22	-0.07		

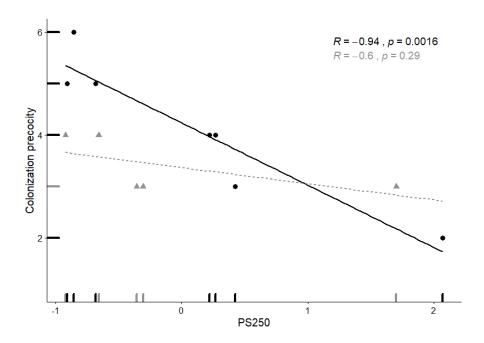


Fig. 2. Tomato greenhouse colonization precocity of the predator *M. pygmaeus* in relation to the herbaceous semi-natural cover at 250m scale (PS250). Colonization precocity is expressed as the sampling event in which the first *M. pygmaeus* adults were detected in each greenhouse by means of yellow traps, with smaller values representing early colonization. The grey dashed line represents the fit for greenhouses with *Calendula* banker plants (triangles); the black solid line represents the fit for those greenhouses without banker plants (circles). For both regressions, R represents the correlation between colonization precocity and herbaceous semi-natural vegetation at 250m scale, and p is the p-value of the correlation.

Supplementary material

Table S1. List of the host plant species of mirid predators adjoining to tomato greenhouses and their presence in the greenhouses sampled in this study. Two greenhouses (H10 and H11) are not included the identified host plants were not present in their surroundings. Greenhouses are ordered according to the number of host plants present, ranging from 10 (H3) to 1 (H12). * is only hosting *M. melanotoma* (Table 3). ^ is a potential host plant of *Macrolophus* spp. (molecular identification was not performed on those individuals).

Host plant species	Family	Type	Habit	Н3	H1	Н8	Н7	H4	Н6	H2	Н5	Н9	H12
Borago officinalis	Boraginaceae	Weed	annual	X	X		X						
Calendula arvensis	Asteraceae	Weed	annual		X		X		X				
Calendula officinalis	Asteraceae	Ornamental	perennial		X					X			
Dimorphoteca ecklonis	Asteraceae	Ornamental	perennial	X									
Dittrichia viscosa*	Asteraceae	Weed	perennial	X									
Erodium spp.	Geraniaceae	Weed	annual	X	X	X	X						
Gallium spp.	Rubiaceae	Weed	annual	X		X							
Lavandula spp.	Lamiaceae	Ornamental	perennial	X									
Malva spp.	Malvaceae	Weed	annual	X	X	X		X					
Leucanthemum spp.	Asteraceae	Ornamental	perennial			X							
Parietaria spp.	Urticaceae	Weed	perennial	X	X	X	X	X	X		X	X	
Salvia officinalis	Lamiaceae	Ornamental	perennial	X									
Sonchus spp.^	Asteraceae	Weed	annual	X	X	X		X	X	X	X	X	X

Fig S.1 Spearman correlations between landscape cover variables at the 250 m scale. IV: protected horticulture, CROP: open field agriculture, AG: riparian a, FO: forest semi-natural; PR: shrub semi-natural, PS: herbaceous semi-natural, URBAN: urban, SN: sum of all semi-natural covers (FO, PR, PS). Circle size and colour represent Spearman correlations with blue representing positive correlations and red negative correlations, all with p <0.05 are highlighted with a *.

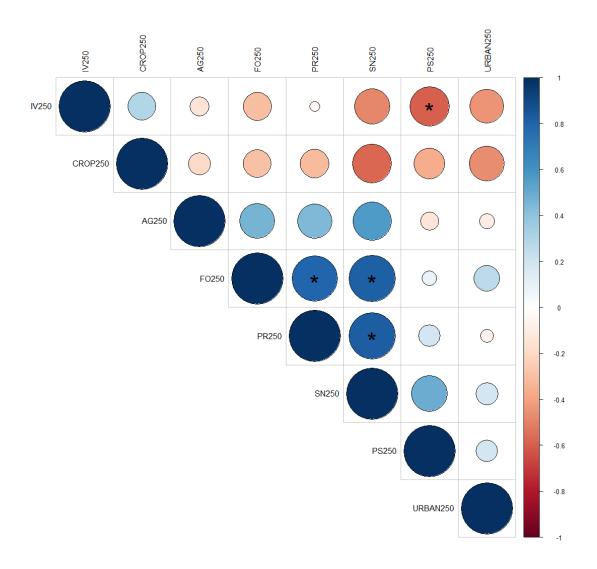
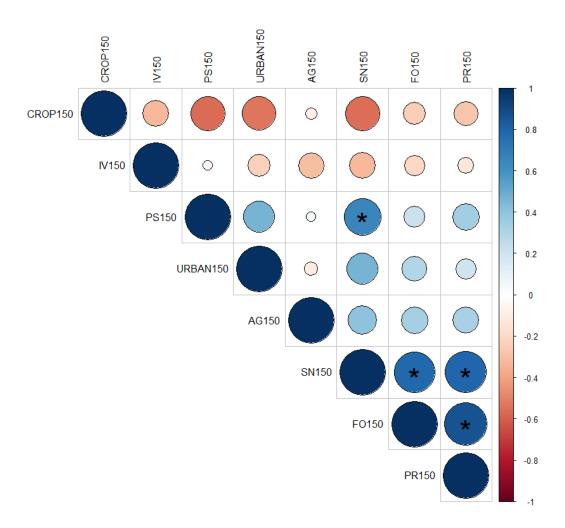


Fig S.2 Spearman correlations between landscape cover variables at the 150 m scale. IV: protected horticulture, CROP: open field agriculture, AG: riparian a, FO: forest semi-natural; PR: shrub semi-natural, PS: herbaceous semi-natural, URBAN: urban, SN: sum of all semi-natural covers (FO, PR, PS). Circle size and colour represent Spearman correlations with blue representing positive correlations and red negative correlations, all with p < 0.05 are highlighted with a *.



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Fig S.3. Spearman correlations between numerical habitat management variables. These variables were obtained by adjoining vegetation sampling in each greenhouse (Table 2). Mp: number of M. pygmaeus captures in adjacent vegetation, Mp_noCal: number of M. pygmaeus captures in adjacent vegetation excluding Calendula officinalis plants, Nt: number of N. tenuis captures in adjacent vegetation, Dt: number of D. tenuis captures in adjacent vegetation, Total_points: number of points of adjacent vegetation sampled in 10 m intervals surrounding the greenhouse, HostR: mirid host plant richness in adjacent vegetation, Points_HostPl: Points with host plant presence, refers to the number of sampling points with confirmed host plants around the greenhouse perimeter. Circle size and colour represent Spearman correlations with blue representing positive correlations and red negative correlations, all with p < 0.05 are highlighted with a *.

