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1 **Title: Peanut protein: an underutilized byproduct with great potential- a**
2 **review**

3 **Running title: Peanut protein**

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8

9 **Abstract**

10 Peanut (*Arachis hypogaea* L.) is the fourth important oilseed in the world. After oil extraction, defatted
11 peanut is a protein-rich byproduct containing around 50% of protein that can enable the production of
12 protein isolates (90% protein) and concentrates (70% protein). Peanut protein has an excellent amino
13 acids profile, a desirable volatile profile, a low level of anti-nutritional factors and a steady supply.
14 Despite these advantages, peanut protein is underutilized because of its poor functional properties caused
15 by the native globular structure and extraction conditions. Nutritional limitations are its deficiency in
16 methionine and lysine and its association with allergic reaction for genetically predisposed subjects. To
17 promote the valorization of peanut protein in foods, it is very important to ensure a better functionality
18 and a better nutritional value. This review intends to cover the properties of native peanut protein and
19 to discuss innovative strategies including physical, chemical, and biological methods to improve the
20 functionality and to mitigate allergens. These strategies have different degree of success in terms of
21 protein quality and functionality, yield, sustainability, and convenience. More investigation is required
22 to select the processing or the combination of processing to boost the application of peanut protein as a
23 valid alternative protein.

24

25 **Keywords:** peanut protein, byproduct, processing, functionality, allergenicity, nutrition

26

27 1. Introduction

28 The quest for new protein sources has been higher than ever. Although animal proteins are primary
29 sources with balanced amino acid profiles, plant-based protein demand has raised for several motives.
30 Plant proteins are being explored as alternatives in food applications to feed a growing population
31 expected to reach 10 billion by 2050. These proteins are affordable, available and have a low
32 environmental impact compared with those deriving from animal sources. Health concerns over the
33 overconsumption of meat and animal-based products are also alarming for some health-conscious
34 consumers choosing to reduce (the case of flexitarian) or to remove meat consumption from their diet
35 as vegan or vegetarian (Boukid, 2020). It is also quite important to highlight that the ongoing pandemic
36 COVID19 contributed into boosting the market of plant proteins due to consumers' awareness toward
37 the relatedness between nutrition and health. Beside these advantages, plant proteins are versatile and
38 can derive from different sources such as cereals, pulses and oilseeds. Among plants, pulses have played
39 an important role in the quest for new vegetable protein sources. Pulses such as pea (*Pisum sativum* L.),
40 chickpea (*Cicer arietinum* L.), fava bean (*Vicia faba* L.), or lupine (e.g. *Lupinus albus*, *Lupinus mutabilis*
41 Sweet, *Lupinus luteus* L.) have high protein content (20 to 40%) compared to cereals like wheat (11-
42 15%) (Boukid *et al.*, 2019). Gluten, derived from wheat, is a commodity food ingredient being used for
43 its functional properties in bread and bakery products (Boukid *et al.*, 2018). Rice, corn kidney bean, pea
44 and amaranth proteins are also of great importance in the gluten-free market (Shevkani and Singh, 2014;
45 Morreale *et al.*, 2019). Oat protein is gaining lot of interest since it has high concentration of proteins
46 (12–20%) and represents a good option for people having allergies to pulses (Boukid, 2021a). Oilseeds
47 are valuable sources of lipid and basically processed for their edible oils leaving behind important
48 amounts of protein-rich byproduct (Chardigny and Walrand, 2016). Proteins are usually recovered from
49 defatted meal or oilcake and marketed as food or feed ingredients. The most produced plant protein
50 derives from soybean (36-40% protein) (Silva *et al.*, 2018) and it dominates the market for years but its
51 reputation as a genetically modified crop is gradually shifting the interest to other sources
52 (MarketsandMarkets, 2019).

53 Peanut (*Arachis hypogae*, L.) is the fourth important oilseed in the world. Peanut is classified among the
54 legumes family (*fabaceae*) with comparable protein content to that of pulses, and it is generally included
55 among oilseeds due to its high oil content (Arya *et al.*, 2016). After oil extraction, defatted meal or
56 oilcake contains up to 50–60% proteins and usually used as fertilizer, feed, or fuel (Zhao *et al.*, 2020).
57 The nutritional value of peanut protein (PP) is high and resembles animal proteins. These proteins have
58 low level of anti-nutritional factors, and excellent amino acid profile that can be easily digested. They
59 have also desirable aroma and taste, and a white color to be used as a potential protein substitute (Ji *et*
60 *al.*, 2017; Phongthai *et al.*, 2020). It is of great significance to extract proteins from defatted peanut and
61 to improve its application value in the food and beverage industries (Arya *et al.*, 2016; Hu *et al.*, 2019).

62 Thus, the development of PP as a commodity is necessary to valorize an important byproduct and
63 provide a high-value ingredient for various food product formulations (Jain *et al.*, 2015). Nevertheless,
64 there are no available PP isolates (PPI) or concentrates (PPC) due to their poor functional properties
65 such as solubility, emulsifying, foaming, and gel properties that limit its applications (Ji *et al.*, 2019).
66 To promote the utilization of PP in foods, it is very important to ensure a better functionality and better
67 nutritional value. Several strategies were applied including physical (e.g., heating, freezing, microwave,
68 ultrasonic, and high-pressure), biological (hydrolysis or crosslinking enzymes) and chemical
69 (phosphorylation and glycation) methods to improve the quality of these proteins (Ji *et al.*, 2017, 2019;
70 Ma *et al.*, 2017). In light of these considerations, this review aims to provide an updated overview about
71 the extraction methods and the native characteristics of PP. Furthermore, modifications strategies for
72 improved PP were discussed with focus on mitigating protein allergenicity and enhancing functional
73 properties.

74 **2. Extraction**

75 Before the extraction, peanut seeds go through a pretreatment phase that includes steps such as cleaning,
76 dehulling and in some cases roasting. Roasting have the aim to destroy the antinutritional factors, and
77 to reduce or eliminate the spoilage and pathogenic microorganisms of raw peanuts (Yu *et al.*, 2020).
78 Defatting or oil removal can be done using different methods mainly hydraulic pressing, screw pressing,
79 solvent extraction, and pre-pressing followed by solvent extraction (Tu and Wu, 2019). Milling is the
80 most traditional approach for oil removal which requires labor and time consuming. Hydraulic press
81 applies pressure on the seeds inside a cylinder until oil release (Sena-Moreno *et al.*, 2016). Screw press
82 is the most efficient mechanic method, in which seeds are crushed in a rotating press barrel. Peanut flour
83 can be defatted by repeated extraction with n-hexane until the fat content is lower than 1 %. The use of
84 solvents can reduce extraction yield and induce undesirable changes in protein structure (Ji *et al.*, 2019).

85 PPI (about 70% protein) are produced from defatted peanut flour or also press cake by removing the
86 remaining oil and water-soluble, and non-protein components. Protein concentration can be carried out
87 by isoelectric precipitation, hexane or/ and aqueous alcohol precipitation, or ultrafiltration.
88 Ultrafiltration membrane (30 kDa) was found also efficient to obtain PPI (72% of protein) with a good
89 functionality such as emulsion stability index (Jain *et al.*, 2015). This can be attributed to the
90 preservation of the native protein structure due the mildly conditions of ultrafiltration and the absence
91 of heating or chemical addition.

92 PPI (about 90-95% protein) are produced by alkali solution and isoelectric precipitation. To remove
93 water-insoluble impurities, peanut flour can be subjected to an aqueous washing phase. Then peanut
94 flour is suspended in alkaline solution (pH 8.0–8.5). After centrifugation, the pellet is discharged, and

95 the supernatant is precipitated under acid conditions (pH 4.5). After centrifugation, the recovered
96 protein was neutralized (pH 7.0). Commonly, the produced PPI is dried using a spray dryer (Ochoa-
97 Rivas *et al.*, 2017). Alkali-solution is a simple and practicable method ensuring a high protein yield and
98 it is the most commonly used for plant protein extraction. Nevertheless, there are several drawbacks
99 including high consumption of solvents and water, high production cost, and waste generation. The use
100 of recent technologies such as ultrasound-assisted extraction improved extraction efficiency, reduced
101 the processing time properties of proteins compared with those obtained using the regular alkali soluble
102 and acid precipitation methods, but further optimization is required prior to upscaling (Sun *et al.*, 2020).
103 Ultrasound-assisted extraction also improved the emulsifying activity index and emulsifying stability
104 index, increased the hydrophobic amino acids and reduced molecular weight fractions compared with
105 alkaline extraction. Ultrasound treatment changed the structure of protein by increasing of surface-to-
106 volume ratio, and thus more protein participated in forming the interfacial layer and increased the
107 emulsifying efficiency (Amiri *et al.*, 2018).

108

109 **3. Native properties of peanut protein**

110 **3.1. Nutrition**

111 PP contains all the 20 amino acids being the richest source of arginine (up to 12.5% of total proteins)
112 (Arya *et al.*, 2016), which is related to several health benefits such prevention of cardiovascular disease,
113 weight management and satiety (Smeets *et al.*, 2021). Like almost plant proteins, PP is deficient in
114 methionine, lysine, threonine and tryptophan. The true protein digestibility of peanuts is comparable
115 with that of animal protein (94 and 97%, respectively) and better than canola protein (84%) (FAO,
116 2017). The protein digestibility-corrected amino acid score (PDCAAS) of PP has been estimated to be
117 about 0.70, which is higher than wheat (0.46) and maize (0.46). However, it is lower than soy protein
118 (0.91) and pea protein (0.82) due the limiting amino acids (Ochoa-Rivas *et al.*, 2017).

119 Enzymes (e.g., flavourzyme and catalase) were also applied on PPI as a post-treatment enabling the
120 generation of hydrolysates rich in bioactive peptides (Phongthai *et al.*, 2020). It was reported that these
121 peptides have DPPH radical scavenging, metal chelating activity and angiotensin I-converting enzyme
122 inhibitory effects, which indicates they may be beneficial for blood pressure regulation (Yu *et al.*, 2021).
123 Antioxidants peptides were also identified in peanut hydrolysates such as Thr-Pro-Ala (286kDa),
124 Ile/Leu-Pro-Ser (315kDa) and Ser-Pro (202kDa) (N *et al.*, 2014).

125 Peanut allergy is considered to be one of the most severe food allergies with a prevalence around 2%
126 (Li *et al.*, 2020a). Currently, strict avoidance is the only treatment and rescue medication upon accidental

127 exposure to peanuts since peanut is a common food ingredient (Zhang *et al.*, 2021a). PP is mainly made
128 by storage proteins, albumins and globulins. Globulins (7S and 11S) comprise of the majority of the
129 total protein (~ 75%) (Ji *et al.*, 2017), and it is subdivided into vicilins (7S globulins) and legumins (11S
130 globulins) (Mueller *et al.*, 2014). Thirteen proteins have been identified as allergens in peanuts (Zhang
131 *et al.*, 2021a). Ara h 1, 2, 3, and 6 are considered the major allergens and are often associated with severe
132 symptoms, while Ara h 5, 7, 8, 9, 10, 11, and 12/13 are considered minor allergens since they do not
133 cause life-threatening allergic reactions (Kim *et al.*, 2019). Ara h 8 has been shown to have cross-
134 reactivity with from birch pollen (Bet v 1) (Palladino and Breiteneder, 2018). Ara h 5, also called peanut
135 profilin, is associated with pollen allergy (profilins from grass and birch pollen, Phl p 12 and Bet v 2,
136 respectively) (Zhang *et al.*, 2021a). Therefore, efficient mitigation strategies are of great interest to
137 develop hypoallergenic PP.

138 **3.2. Functionality**

139 Solubility of proteins are among the most important functional properties. Naturally, due to the rigid
140 globular structures, native PP has limited solubility (Rasheed *et al.*, 2020). Compared to soy protein, PP
141 has high molecular weight and lacks ionizable groups resulting in poor solubility (Ji *et al.*, 2019). As a
142 function of pH, the maximum solubility (around 80-85%) was at pH=2-3 due to the ampholytic nature
143 of PP (Wu, 2009). At pH=4-5, the values of solubility are the minimum (up to 20%) similarly to faba
144 bean, cowpea, and chickpea proteins due to the formation of hydrophobic aggregation (Shevkani *et al.*,
145 2015a; Vogelsang-O'Dwyer *et al.*, 2020). At pH (6-10), solubility gradually increased until reaching
146 maximum values (80-90%) under basic conditions (pH> 10). As a function of extraction method, PPI
147 showed higher solubility over all pH ranges compared to PPI, which can be attributed to the partial
148 protein denaturation during extraction similarly to faba bean, pea, chickpea and soy protein isolates
149 (Boye *et al.*, 2010; Boukid, 2021b; Boukid *et al.*, 2021). For instance, at neutral pH, PPC had a solubility
150 of 80 %, compared to chickpea, faba bean, pea, kidney bean and soy protein concentrates (Shevkani *et*
151 *al.*, 2015b; Martinez *et al.*, 2016; Vogelsang-O'Dwyer *et al.*, 2020). PPI has higher water holding
152 capacity and oil binding capacity compared to those of PPC due to the high degree of degradation
153 resulting unfolding of the polypeptide chain thereby a higher capacity to water entrapment (Jain *et al.*,
154 2015). Water holding capacity of peanut was found higher than faba bean but lower than soy protein
155 isolates (Table 1). Similarly, the oil holding capacity of peanut was found higher than faba and to soy
156 protein isolates (Tontul *et al.*, 2018).

157 Foaming capacities of PPI were found higher than chickpea; within the same range with cowpea, pigeon
158 pea and lower than kidney bean, faba bean, pea, and amaranth proteins (Shevkani *et al.*, 2014; Martinez
159 *et al.*, 2016; Mohanan *et al.*, 2020). Emulsifying capacity of PPI was compared to faba bean, pea and
160 soy protein concentrates (Table 1). Furthermore, foaming and emulsifying capacities of the PPC were
161 higher than those of PPI as PCC have higher number of polypeptide chains allowing more fluid to be

162 incorporated (Yu, 2007; Jain *et al.*, 2015). Overall, several factors can influence the functionality of PP
163 including protein concentration, protein structure, viscosity, and pH (Shevkani *et al.*, 2014). Considering
164 the method of extraction, the functional properties of PP change significantly since processing impact
165 protein structure. Further studies on the impact of processing on PP will provide insightful information
166 to modulate protein functionality and thus tailor their properties to fit specific food applications.

167 **Table 1

168 **4. Modification strategies for peanut protein with improved properties**

169 **4.1. Mitigation strategies to reduce adverse reaction to peanut protein**

170 Boiling of PP resulted in Ara h 1 degradation and the aggregation of fragments resulting in increased
171 surface hydrophobic index and a decreased content of α -helixes in rAra h 1. This suggests that the
172 epitope lost its native structure due to the heat treatment, which reduced the allergenic nature of rAra h
173 1 (Tian *et al.*, 2018). Boiling and frying of peanut reduced the contents of Ara h 2, 6, and 7 as well as
174 Ara h 8 and Ara h 9 up to 50-70% (Dhital *et al.*, 2014). This can be due to the dissolution of allergens
175 in boiling water or oil. Roasting ($> 130^{\circ}\text{C}$ for 20 min) reduced the sensitivity of Ara h1, while over
176 140°C enhanced IgE-binding capacity of Ara h 1 and Ara h 2 (Zhang *et al.*, 2018). This can be due to
177 higher level of trypsin inhibition activity in roasted peanut and the formation of new complex molecules
178 having allergenic potential (Shah *et al.*, 2019). Cold plasma, high-pressure, gamma irradiation, and
179 pulsed-electric field are also being explored. Cold plasma reduced antigenicity by 65% for Ara h 1 and
180 66% Ara h 2 (Venkataratnam *et al.*, 2020). The allergenicity could be reduced by limited enzymatic
181 hydrolysis and high-pressure homogenization (Ma *et al.*, 2017). The combination of high pressure (500
182 and 600 MPa) and thermal treatment (at 75°C) reduced IgE binding to Ara h 2 (Long *et al.*, 2016). Ara
183 h 6 was completely degraded after being treated with gamma irradiation (Luo *et al.*, 2013). Pulsed-
184 electric field had limited effect on the structural alteration of peanut (Ara h 2, Ara h 6) and thus on the
185 allergenic potential (Zhang *et al.*, 2021b).

186 Regarding chemical modifications, Ara h 2 and Ara h 6 were reduced after a treatment with dithiothreitol
187 followed by alkylation with iodoacetamide. This treatment altered the tertiary and secondary structure
188 of protein due to the loss of the α -helix and the increase in the β -sheets (Apostolovic *et al.*, 2013). The
189 complexation of polyphenols to peanut flour was found efficient in reducing the allergenic potential of
190 PP by mitigating cell degranulation (Plundrich *et al.*, 2017). The phytic acid treatment of peanut extract
191 resulted in the formation of a complex with Ara h 1 and Ara h 2, and reduced IgE binding of the obtained
192 solution (Chung and Champagne, 2007). This can be explained by the ability of phytic acid to precipitate
193 the allergen and reduce its exposure to digestive enzymes.

194 Enzymatic hydrolysis is a safe, no added- chemical and requires low energy inputs for the reduction of
195 peanut immunogenicity. Enzymatic hydrolysis using papain, ficin and bromelain has been reported as
196 an efficient strategy to decrease IgE-binding up to 85-95% (Meng *et al.*, 2020). Likewise, the hydrolysis
197 of allergens from peanut extracts using alcalase and flavourzyme reduced 91.8% of IgE binding (R *et*
198 *al.*, 2015). Hydrolyzing raw peanuts with alcalase and papain was efficient to reduce IgE binding to
199 Ara h 1 (up 100%), Ara h 2 (up 99%), and Ara h 6 (up 88%) and Ara h 3 (up to 46%) (Mikiashvili and
200 Yu, 2018). In addition, combining physical and enzymatic treatments drastically decreased Ara h1
201 and h2 amounts (Ma *et al.*, 2017). Cross-linking enzymes (microbial polyphenol oxidase and laccase)
202 modified tertiary structure of PP, and increased production of IgG2a antibodies and reduced IL-13
203 secretion (Mihajlovic *et al.*, 2016). Microbial transglutaminase formed a compact structure and reduced
204 surface hydrophobic index and increased steric hindrance of rAra h 1 (Hu *et al.*, 2019). As a result, the
205 formed complex did not bind with antibodies, and consequently had reduced allergic reaction (Tian *et*
206 *al.*, 2020).

207 **4.2. Strategies for better functionality**

208 To improve the functional characteristics of PP and to meet the additional requirements of the food
209 industry, high-pressure microfluidization treatment enabled to increase PPI solubility. This treatment
210 induced protein disaggregation by changing the polar environment and promoting surface
211 hydrophobicity in aqueous dispersion (Gong *et al.*, 2019). Atmospheric cold plasma treatment decreased
212 the degree of protein aggregation and increased the number of protein surface hydrophilic groups,
213 thereby unfolding the protein secondary structure. This would enhance the polarity on the protein surface
214 and generate more protein–water binding sites, thereby improving the water solubility, emulsion
215 stability, and water holding capacity (Ji *et al.*, 2017, 2019). Multiple freeze-thaw cycles increased the
216 carbonyl content and particle size of PPI resulting in improving the emulsifying properties. The best
217 emulsifying ability was obtained after 3 cycles where the obtained emulsion had small particle size and
218 uniform distribution (Feng *et al.*, 2020). Thermosonication followed by proteolysis unfolded proteins
219 and reduced particle size resulting in a remarkable increase in protein solubility for the hydrolysates
220 (Zhang *et al.*, 2019). Nanotechnology resulted in increasing the surface hydrophobicity of PPI compared
221 to untreated PPI. The formed nanoparticle improved emulsion ability and stability (Ning *et al.*, 2020)

222 As for chemical modifications, pH-shifting (pH 2, pH 4, pH 10, and pH 12) enabled the modulation of
223 PPI properties (Li *et al.*, 2020b). At pH 10, water holding capacity was improved due to the decreased
224 particle size, increased solubility, free sulfhydryl group content and surface hydrophobicity, while at
225 pH12 gel ability was lost due to protein aggregation. Phosphorylation using sodium trimetaphosphate
226 improved emulsifying activity of PPI (Sánchez-Reséndiz *et al.*, 2018).

227 Partial hydrolysis of protein improved peanut functional properties (solubility foaming capacity, foam
228 stability and emulsifying ability) due to the exposure of hydrophobic groups, liberation of ionizable
229 groups and the formation of hydrolyzed proteins with better ability to bind oil and water (Pan *et al.*,
230 2017; Chen *et al.*, 2018). However, protein hydrolysates may lose functional properties, depending on
231 the type of enzyme and degree of hydrolysis (Chen *et al.*, 2018). Enzymatic hydrolysis using papain
232 combined with high-pressure homogenization reduced particle size and improved solubility but to less
233 extent emulsifying, fat-binding and foaming abilities (Ma *et al.*, 2017). Extrusion pretreatment and
234 papain-induced proteolysis increased the degree of hydrolysis and protein solubility leading to improved
235 emulsifying ability of the hydrolysates (Chen *et al.*, 2018).

236 **5. Conclusion**

237 Plant-based proteins are having a momentum in the food and beverage industries, which gives room to
238 emerging sources such as PP. The use of peanut as a source of proteins have several advantages mainly
239 valorizing an important byproduct of peanut oil/butter industry, and thus reducing the environmental
240 impact of this industry. PP can be a promising food ingredient due to their desirable color and flavor,
241 and good composition of essential and non-essential amino acids. Nevertheless, PP has some cons
242 specially their deficiency in methionine and threonine levels, which can be overcome by making blends
243 of proteins from different sources. PP is also among the list of foods allergens but several approaches
244 are being applied to mitigate epitopes triggering allergenic reactions. Functionally, the low solubility
245 among other properties is limiting the use PP as a food ingredient. Nevertheless, chemical modification
246 was found efficient in improving protein functionality but it includes the uses of chemical and generates
247 high wastes; physical modification requires high energy consumption and in some cases increases
248 allergen reaction; while biological modification can provide chemical-free ingredients but more
249 investigation is required to control the outcome of the proteolytic process. Developing hurdle approaches
250 such as enzymes and ultrasound can be promising taking into account safety, functionality, price and
251 sustainability.

252 **Acknowledgements**

253 This work was supported by CERCA Programme (Generalitat de Catalunya).

254 **Author contributions**

255 **Fatma Boukid:** Conceptualization (lead); Methodology (lead); Writing-original draft (lead); Writing-
256 review & editing (lead).

257 **Conflict of interest**

258 None.

259 **Compliance with ethics requirements**

260 This article does not contain any studies with human or animal subjects.

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