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- 1 Impact of fruit bagging and postharvest storage conditions on quality and decay of
- 2 organic nectarines
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9 ABSTRACT

10 Abiotic factors such as light influence the physicochemical properties of fruit and may alter the 11 response of the fruit to the environment. This study aimed to investigate the effect of two 12 postharvest storage conditions on the overall quality and natural fungal disease incidence (fruit 13 decay) of organic nectarines. Experiments were conducted with four organically grown nectarine 14 cultivars (two early-mid season and two late-season) that were unbagged or bagged during 15 preharvest. After harvest, they were stored for 7-9 days in darkness or under a treatment with 16 lighting. Quality parameters (weight, diameter, firmness, soluble solids content, titratable acidity, 17 and single index of absorbance difference), ethylene production, and fruit decay (as a percentage 18 of rot incidence) were evaluated. Preharvest bagging reduced fruit decay in the late-season 19 cultivars, in which storage under darkness reduced fungal decay (up to 100%) more than storage 20 under lighting treatment (47.1% of reduction). Bagging altered the initial fruit quality, but values 21 were within official recommendations. Storage conditions reduced differences attributed to 22 bagging, especially under storage with lighting. This work highlighted the importance of 23 modulating the light, both in the field by fruit bagging and during postharvest, to reduce fruit 24 decay and improve fruit quality. This may serve as a tool for both farmers and postharvest chain 25 managers.

Keywords: Fungal diseases; late-season cultivars; lighting treatment; postharvest chain; stone
 fruit; sustainable fruit production

28 Introduction

29 Peach, nectarine, plum, cherry, and apricot (*Prunus* genus) are the most economically 30 important species of stone fruit (Mari et al. 2019). In 2019, the worldwide production of 31 peach and nectarine was 25.7 Mt and China, Spain, Italy, and Greece were the main 32 producers (FAO 2021). Stone fruit can suffer pathological diseases and physiological 33 disorders, which lead to fruit losses (Mari et al. 2019; Manganaris and Crisosto 2020). 34 Fruit decay can occur both preharvest and during the postharvest chain (Eckert and 35 Ratnayake 1983), although postharvest losses tend to be greater than orchard losses (Porat et al. 2018). The most destructive and economically important fungal disease is brown 36 37 rot, caused by *Monilinia* spp. (Mari et al. 2019; Mustafa et al. 2021), producing up to 7% 38 and over 60% of incidence at harvest and after postharvest, respectively (Villarino et al. 39 2012). Other relevant diseases are caused by pathogens such as Rhizopus spp., Mucor 40 spp., and Geotrichum candidum (Mari et al. 2019).

Currently, fungal diseases are mainly controlled with a combination of cultural practices (e.g. tree management and removal of natural inoculum sources) (Villarino et al. 2012; Bussi et al. 2015; Casals et al. 2015), biological control, and chemical fungicide programs applied in the orchard (De Oliveira Lino et al. 2016; Mari et al. 2019). Nevertheless, health concerns related to the environmental footprint and toxicological risks have led to a demand for chemical-free fresh fruit (Usall et al. 2015), encouraging more sustainable systems and organic agriculture.

48 Fruit bagging is an environmentally friendly strategy for plant protection in 49 organic production that is extensively used in several fruit crops [e.g. apple (*Malus* spp.), 50 pears (Pyrus spp.), mango (Mangifera spp.)] (Sharma et al. 2014). It is also a required 51 agricultural practice in the 'Calanda peach' origin appellation from Teruel, Aragón 52 (Spain) (Faci et al. 2014). This mechanical technique consists of introducing the fruit into 53 a bag during the stone hardening phase until harvest when it is removed. Bags can be 54 made of many materials (e.g. paraffin, plastic, paper) and can be of different colours (e.g. 55 white, yellow, brown) (Ali et al. 2021). Bagging reduces physical injuries, fruit decay 56 [e.g. brown rot (Monilinia spp)], and cracking and russeting incidence in peaches (Prunus

persica) (Keske et al. 2014; Sharma et al. 2014; Campbell et al. 2021), as well as
improving visual quality (e.g., colour development) and altering fruit quality (Zhou et al.
2019; Ali et al. 2021) by affecting the solar radiation that fruit receives on the tree.
However, the results of this strategy are contradictory among investigations, probably
due to external factors (i.e. type of bag and storage conditions) or the fruit's intrinsic
properties (Sharma et al. 2014).

63 After harvest, the conditions in which stone fruit is stored are crucial to avoid 64 disease and physiological disorders (Manganaris and Crisosto 2020). Temperature and 65 relative humidity have been extensively studied. Still, the effect of white artificial lighting 66 along the postharvest chain (i.e. packinghouses, markets, and consumers' homes) on fruit 67 quality and disease incidence (fruit decay) has not been studied. Artificial lighting can 68 alter many physicochemical fruit properties and improve fruit quality in peaches. For 69 example, blue light increases total sugar content in peaches (Gong et al. 2015), and UV-70 B radiation reduces firmness, but it does not affect the soluble solids content and titratable 71 acidity (Santin et al. 2019). UV-B radiation also affects plant defence signalling (Ballaré 72 2014) and the peach phenolic response to Monilinia fructicola (Santin et al. 2018). 73 Recently, Balsells-Llauradó et al. (2021) studied the effect of postharvest storage under a 74 photoperiod of unbagged and bagged fruit in response to artificial inoculations of 75 *Monilinia* spp. These authors found that the light received by nectarines during preharvest 76 modified the intrinsic fruit properties, influencing the response to Monilinia spp. once 77 stored under postharvest treatments with lighting. Still, the effect of photoperiod and fruit 78 bagging on fruit quality after postharvest storage remains unknown.

Fruit quality includes all aspects related to physical, mechanical, sensory, nutritive, and appearance properties, and properties related to food safety (Crisosto and Costa 2008). The purposes of this study were i) to evaluate the effect of bagging on fruit quality and ethylene production of four nectarine cultivars at harvest, ii) to assess the effect of fruit bagging on natural fungal disease incidence (fruit decay) under two postharvest storage treatments (darkness and lighting treatment), iii) to decipher the effect of these postharvest storage treatments on fruit quality.

86 Materials and methods

87 Plant material and fruit bagging

88 Four yellow-fleshed cultivars of nectarines (P. persica var. nucipersica (Borkh.) 89 Schneider) were used for the studies. Two early-mid season cultivars (Fantasia and 90 Venus) and two late season cultivars (Albared and Nectatinto) were obtained from organic orchards located in Lleida (Catalonia, Spain), which followed the European and 91 92 national standards of organic agriculture (Generaliat de Catalunya 2022). The incidence 93 of fruit decay and the quality measurements were assessed on unbagged fruit and fruit 94 that was bagged in the orchard (bagged fruit). Commercial single layer white paper bags (16.5 x 21.5 cm, 32 g m⁻²) (Gràfiques Salaet, Gandesa, Catalonia), impregnated with 95 96 paraffin wax, were used to bag fruit before harvest (185, 172, 185, and 197 Julian days 97 (Julian days = January 1st was considered as day 1) for Fantasia, Venus, Albared, and 98 Nectatinto, respectively) using a staple to fasten the bag to the branch. The harvest date 99 was at commercial fruit maturity based on the grower's recommendations. Fruit was 100 harvested at 218, 221, 250, and 260 Julian days for Fantasia, Venus, Albared, and 101 Nectatinto, respectively. Bagged and unbagged fruit from the same sun-side of trees to 102 avoid fruit position effects (Minas et al. 2018) were randomly harvested. Fruit was 103 homogenized based on the single index of absorbance difference (IAD) using a portable 104 DA-Meter (TR-Turoni, Forli, Italy). A lux meter was used to assess the incident solar 105 radiation inside the bags. Bags were removed upon arrival at the laboratory before

106 conducting the assays and postharvest storage.

107 Storage conditions and evaluation of postharvest decay losses

108 Fruit was stored as described by Balsells-Llauradó et al. (2021). Briefly, the fruit was 109 stored at high humidity (20 °C, 90 \pm 3 % RH) for 24 h at darkness, and then placed in a 110 postharvest chamber under two controlled shelf-life conditions, both at 22 ± 2 °C and 50-111 90 % RH. The darkness treatment consisted of complete darkness, and the treatment with 112 light consisted of a photoperiod of 12h/12h (light/darkness) with four incandescent white 113 TL-D 36 W/827 fluorescent lights (temperature = 2700 K, 3350 lm, 350 - 740 nm, 630 114 nm max.; Philips, Madrid, Spain). Experiments were conducted with 4 replicates of 5 115 fruits each in each bagging condition and postharvest storage combination for each 116 cultivar. Fruit was examined daily to detect decayed tissue. The evaluation was recorded 117 for 9 days in early-mid season cultivars. Due to the early and high perishability in late-118 season cultivars, evaluations were conducted for up to 7 days. The incidence of fruit decay 119 was calculated as the percentage of fruit with natural fungal disease symptoms. 120 Identification of fungal agents was carried out following the EPPO standard PM 7/18 (3) 121 (Bulletin OEPP/EPPO 2020) and Mari et al. (2019).

An economic evaluation between bagged and unbagged fruit was conducted considering the production of an organic orchard of one hectare in Ebro Valley area. Orchard characteristics and average prices (cost of paper bags and labour input for bagging and bag removal) are listed in Table 1. Once the results of fruit decay after postharvest storage were obtained, the cost-effectiveness of fruit bagging was also calculated in Table 2.

128 *Quality characteristics and ethylene measurements*

129 Quality characteristics were measured according to Baró-Montel et al. (2019), i.e. weight,

130 cheek diameter (CD), flesh firmness (FF), soluble solids content (SSC), titratable acidity 131 (TA), and the I_{AD}. These measurements were performed on the harvest day (initial fruit 132 quality) and at the end of the postharvest storage period. Experiments were conducted with 4 replicates of 5 fruits each in each bagging condition and postharvest storage 133 134 combination for each cultivar. After postharvest storage, changes in quality were 135 calculated in relation to initial fruit quality (as percentages) for each cultivar, bagging 136 condition, and postharvest storage, following the formula [(initial quality - final quality) 137 / initial quality x 100]. Ethylene measurements of fruit at harvest were determined as 138 described by Giné-Bordonaba et al. (2017). Fruit was placed in 3.8 L sealed flasks for 2 139 h. After ethylene measurements, the fruit was returned to their respective postharvest 140 storage condition. Ethylene was measured using 4 replicates of 3 fruit each, for each 141 bagging condition and postharvest storage combination per cultivar.

142 Statistical analysis

143 JMP® software version 14.2.0 (SAS Institute Inc., Cary, NC, USA) was used to 144 analyse the data statistically. All data were checked for the assumptions of parametric statistics and were transformed when needed. Ethylene production data (nL kg⁻¹ h^{-1}) were 145 146 subjected to Log transformation. Analysis of variance (ANOVA) was applied to the data, 147 and when the analysis was statistically significant, Tukey's HSD test ($p \le 0.05$) was used 148 to compare the incidence of fruit decay at each time point for each cultivar. To compare 149 the two different means of bagging conditions or the two postharvest storage conditions, 150 Student's T-test ($p \le 0.05$) was used. Pearson correlation analyses were conducted 151 between fruit decay and fruit quality at the end of the storage for cv Albared using the 152 software SigmaPlot (v. 13.0). Correlations were significant at $p \le 0.05$.

153

154 **Results**

155 Preharvest fruit bagging slightly impaired fruit quality and ethylene at harvest

156 To assess the effect of fruit bagging on quality, and the decay of nectarines after 157 postharvest storage, the quality at harvest (initial quality) was initially evaluated. Paper 158 bags allowed up to 76% of the light intensity to pass on the south-south-east side of the 159 trees. This led to significant differences in all quality parameters based on the different 160 bagging conditions of the cultivars (Table 3). In cv. Venus, bagged fruit had significantly 161 smaller weight and CD than unbagged fruit (15.5% and 6.5% lower, respectively), 162 whereas only Albared bagged fruit was significantly larger (25.7% and 8.3% higher 163 weight and CD, respectively) than unbagged fruit. Bagging the fruit also significantly (p 164 ≤ 0.05) impaired the I_{AD} (e.g., reduced maturity) in the early-mid season cv Venus (Table 3). In contrast, in the late season cultivars the IAD was significantly smaller (more mature 165 166 fruit) in the bagged fruit than in the unbagged fruit. Fruit bagging also altered FF in both 167 early-mid season cultivars, although not in the same direction (Table 3). Ethylene levels 168 differed significantly in Fantasia only, i.e., unbagged fruit produced 6.9-fold higher 169 ethylene levels than bagged fruit.

170 Fruit bagging reduced fruit decay during postharvest and was cost-effective

In unbagged fruit, the incidence of disease was higher in the late cultivars (up to 75 85%) than in the early-mid season ones (up to 30 - 35%) (Figure 1). The onset of disease
was observed earlier (one day after storage) in the late cultivars than in the early-mid ones
(4 - 6 days after storage). The fungal pathogens detected were mainly *Monilinia* spp.
(especially *M. fructicola*), and *Rhizopus* spp.

Overall, fruit bagging reduced and even prevented the appearance of decay during
postharvest, in some cases to 0 (Figure 1). In unbagged cv Fantasia, fruit decay was found

to be 5% under darkness and 10% under lighting storage, which was slightly higher than bagged fruit (0 and 5%, respectively) at the end of storage, although not statistically significant (Figure 1A). Unbagged nectarines of cv Venus had more disease under both darkness and lighting (35 and 20%, respectively) than bagged fruit (0% in both postharvest conditions) after 9 days of storage (Figure 1B). This represented a 100% reduction in both postharvest conditions.

184 In both late season cultivars, fruit decay was observed from the first day of 185 storage, and gradually increased along storage time (Figure 1C, D). Fruit decay was 186 observed in cv Nectatinto in all conditions on all days of storage. In unbagged fruit of 187 Nectatinto, decay was already 15 and 10% under darkness and lighting after 1 day of 188 storage, respectively, and increased steadily thereafter. At day 7, decay of fruit ranged 189 from 35 to 65%, although the differences among bagging and storage conditions were not 190 significant on any individual day. In cv Albared, the disease incidence in unbagged fruit 191 was prominent at day one of storage in both treatments (35 and 50%, respectively. Figure 192 1D). Decay increased with increasing storage time and reached 75 and 85%, respectively, 193 at 7 days. Interestingly, no decay was observed in the bagged fruit stored under darkness. 194 Contrarily, the bagged fruit stored in the lighting treatment developed disease symptoms 195 on day 4 of storage. Decay in unbagged fruit was significantly higher ($p \le 0.05$) than 196 bagged fruit under both postharvest treatments (100 and 47.1% of reduction, 197 respectively).

The economic evaluation indicated that fruit bagging was cost-effective for the four nectarine cultivars tested in this study. For example, the organic nectarine orchard produced approximately 22.5 t ha⁻¹ of fruit and achieved up to 27 k \in ha⁻¹ (Table 1). In this study, the unbagged early-mid season cultivars showed 25% postharvest losses (after 6 days of storage under darkness at 20 °C) representing a loss of 6750 \in ha⁻¹ (Table 2). Similarly, the late season cultivars displayed postharvest losses of up to 60% (after 3 days of storage under darkness at 20 °C), representing a loss up to 16,200 \in ha⁻¹. Considering the cost of the paper bags and the labour input for bagging and bag removal (Table 1), fruit bagging was still worthwhile compared to production without bags, being much more cost-effective when considering the late cultivars (+ 10,575 \in) compared with the early-mid season cultivars (+ 1,125 \in).

209 Postharvest storage minimised fruit quality differences between bagging conditions

The effect of bagging was also evaluated in terms of fruit quality changes suffered after postharvest storage. Under darkness, most of the quality parameters were similar in bagged and unbagged fruit, although there were small but significant differences in two cultivars (Table 4). Unbagged Fantasia fruit was significantly larger (i.e. weight and CD) and had lower I_{AD} and FF compared to bagged fruit. However, in Albared, unbagged fruit had significantly higher I_{AD}, FF, and SSC/TA ratio but lower SSC and TA than bagged fruit.

Under storage with lighting storage, the quality of bagged and unbagged fruit was more uniform in all cultivars (Table 5). Bagged fruit of Fantasia had significantly higher firmness, SSC, and TA values than unbagged fruit (9 vs 6.6 N, 12.7 vs 11.9 °Brix and 7.5 vs 5.8 g L⁻¹, respectively). However, no effect attributable to bagging was observed in Venus under lighting, and only a significant difference in weight or CD was observed in the late cultivars (Table 5).

To putatively relate fruit quality with fruit decay, a correlation analysis was conducted. The results showed that, for example, in Albared, TA was negatively correlated with fruit decay ($R^2 = -0.97$, p = 0.026) whereas the correlation between SSC/TA ratio and decay was positive ($R^2 = 0.93$, p = 0.0067; Figure 1, Table 4, Table 5).

227 Changes in fruit quality under darkness vs lighting in relation to harvest day

228 To evaluate which postharvest storage condition triggered a greater change in fruit 229 quality, the percentage of reduction or increase was calculated relative to the initial 230 quality for each postharvest condition, bagging condition, and quality parameter. There 231 were differences between storage conditions within each bagging condition in some 232 cultivars (Figure 2). Size parameters (weight and CD) altered the least in comparison to 233 initial quality; reductions were below 19% for all cultivars except Nectatinto (16 - 33%)234 and Albared bagged and stored under darkness (30%). In contrast, reductions in FF and I_{AD} were the highest (40 to 92% reduction). 235

236 In bagged fruit, the reductions of weight and CD under darkness were 237 significantly greater than under lighting conditions, in both Fantasia and Albared (Figure 238 2A). The reduction in I_{AD} under darkness was significantly lower than under lighting in 239 Nectatinto, and in Fantasia the reduction in FF under darkness was significantly lower 240 than under lighting (Figure 2B). In addition, the SSC/TA ratio increased in Fantasia and 241 Nectatinto cultivars under both storage conditions. However, the changes in SSC/TA ratio 242 in darkness vs lighting were significant in Venus and Albared, but in opposite directions 243 (Figure 2C).

Changes observed in unbagged fruit were like those observed for bagged fruits. The reduction of weight and CD under darkness was significantly higher than under lighting in both Fantasia and Nectatinto nectarines (Figure 2D). Although there were no differences in the I_{AD}, the reduction of FF was significantly smaller in Fantasia under darkness than under lighting (Figure 2E). The increase of SSC/TA ratio under lighting was higher than under darkness in all unbagged cultivars, although the difference was significant only in Fantasia (Figure 2F).

251 Discussion

252 Effects of preharvest fruit conditions on fruit quality at harvest

253 Fruit undergoes physiological changes throughout its development, and external factors 254 are crucial in determining fruit quality. Bagging fruit during its development influences 255 the quantity (intensity) of solar irradiation that it receives in the field but also the light 256 quality (wavelength of electromagnetic spectrum, i.e. colour) that irradiates the fruit. In 257 the work reported here, the reduction of light intensity was around 24%, suggesting that 258 bagging could have impaired some fruit quality parameters. This could explain the 259 differences between bagged and unbagged fruit in FF and SSC in Fantasia and Venus, 260 and the relatively small effect in Nectatinto and Albared. In a study conducted with UV-261 B radiation, it was shown that UV-B radiation reduced the activity of cell wall-modifying 262 enzymes (e.g. pectin methylesterase and polygalacturonase), leading to loss of firmness, 263 but without affecting the SSC and the titratable acidity (Santin et al. 2019). This suggests 264 that a better understanding of the mechanisms underlying the effects of white light quality 265 on FF and sugar content could help to ensure desired quality.

266 In organic peaches, fruit bagging was also found to alter the I_{AD} (Campbell et al. 267 2021), and I_{AD} values were correlated with chlorophyll content (Spadoni et al. 2016). 268 In the presented work, the I_{AD} in 3 out of 4 of the bagged cultivars was strongly 269 differentiated on the harvest day. This could also be explained by the cultivar-dependent 270 effect that influence the fruit quality (e.g. FF, SSC, and TA) (Iglesias and Echeverría 271 2009). The bagging process as well as shortening the bagging duration, can delay 272 chlorophyll degradation and improve the anthocyanin content in peach peel, respectively 273 (Zhou et al. 2019). Overall, changes related to FF, chemical content, and pigmentation 274 were related to the presence or absence of bags in the field. This suggests that the producer 275 should consider the type of bag and the specific cultivar response before bagging the fruit.

276 Altered fruit quality and fruit decay incidence

277 Host susceptibility to pathogens can depend on the ongoing physicochemical and 278 physiological changes during fruit development and ripening, as well as the fruit 279 characteristics intrinsic to the cultivar (Baró-Montel et al. 2020). In the work reported 280 here, TA and SSC/TA were negatively and positively correlated, respectively, with fruit 281 decay. Sugars are the major soluble solids in fruit juice and have been implicated in biotic 282 (Kou et al. 2018) and abiotic stress responses (Wang et al. 2013). Among sugars, sucrose 283 was the major soluble sugar, ranging from 55.74% to 72.96% of the total sugar content 284 (Reig et al. 2013). The development of brown rot (*Monilinia* spp.), the main disease of 285 stone fruit, has been positively associated with sucrose (Baró-Montel et al. 2020) and 286 SSC (Gradziel 1994) as nutrients for fungal growth. Hence, fruit quality of different 287 cultivars stored under different conditions either favoured or restricted the onset of fruit 288 decay.

289 Ethylene is also involved in the responses to abiotic (Müller and Munné-Bosch 290 2015) and biotic stresses, either acting against necrotrophic pathogens (Glazebrook 2005) 291 or being conductive to disease susceptibility (Van Der Ent and Pieterse 2012). Here, 292 ethylene production was reduced by fruit bagging in Fantasia (6.9-fold difference 293 between unbagged and bagged fruit), but increased slightly in the other cultivars tested 294 (Table 3). Ethylene is required for fruit softening (Hayama et al. 2006), and, as expected, 295 the highest ethylene production was accompanied by a reduced FF. The high ethylene 296 production in unbagged Fantasia fruit may have increased susceptibility to fruit decay, as 297 well as the high ethylene production in late-season cultivars, which presented an 298 increased fruit decay incidence. In nectarines artificially inoculated with *Monilinia* spp., 299 fruit bagging altered ethylene production during postharvest, but all fruit was susceptible 300 to Monilinia spp. under both treatments (Balsells-Llauradó et al. 2021). In the study

301 reported here, ethylene production in late cultivars may have favoured ripening-302 associated events, such as loss of FF, which made the fruit more susceptible to decay.

303 The reduction of fruit decay by fruit bagging was cultivar- and postharvest storage304 dependent, but was cost-effective

305 Infections occurring along the postharvest chain can remain quiescent or cause latent 306 infections until favourable factors trigger disease development (Luo et al. 2005; Garcia-307 Benitez et al. 2020). Incubation in humidity with photoperiod lighting favours naturally 308 occurring diseases in peaches (Villarino et al. 2012). Here, the incubation period of the 309 observed decay suggested that the early-mid season cultivars probably had relatively 310 more quiescent conidia that developed later in time. In contrast, the late cultivars probably 311 had relatively more field-occurring infections that remained briefly latent and were 312 visible early in storage (Figure 1A, B). For peaches, bagging is common in late cultivars, 313 which are exposed to more favourable climatic conditions for pests and diseases than 314 early cultivars, to protect the fruit against insects such as the Mediterranean fly (Ceratitis 315 capitata) (Faci et al. 2014) and other fungal diseases such as brown rot caused by 316 Monilinia spp. (Mari et al. 2019). However, in orchards with high brown rot disease 317 pressure, neither biological nor chemical treatment were completely effective (Casals et 318 al. 2021). Thus, the low efficacy of bagging could be attributed to a high inoculum 319 pressure in the field, especially in Nectatinto.

In this study, exposure to darkness also reduced fruit decay, in a cultivar- and bagging-dependent manner (Figure 1B, D). Roeber et al. (2021) found that impaired solar radiation affected both abiotic and biotic stress-triggered responses. UV-B radiation can also regulate plant metabolisms such as gene expression of terpene synthases and the content of terpenoids and phytoalexins in peaches (Liu et al. 2017; Santin et al. 2021). In particular, the expression of terpenoids and phenylpropanoids has been implicated in both susceptibility and resistance of nectarines to *Monilinia laxa* (Balsells-Llauradó et al. 2020). Hence, in this study, the distinct level of solar radiation caused by fruit bagging may have induced changes in fruit that differentially affected their ability to face pathogens under different postharvest storage conditions. Deciphering the role of secondary metabolites (e.g. phenolics, terpenoids, and phenylpropanoids) in response to both pathogens and lighting conditions could improve our understanding of the disease development.

333 Scarce information exists related to the economic viability of fruit bagging (Blasi 334 et al. 2017), which does not specify whether or not the losses occurring during the 335 postharvest chain are considered. The economical evaluation conducted in this study to 336 test the differences between an orchard with or without bagged fruit, suggested that if 337 fruit bagging is applied in similar orchards to the ones reported in this study (i.e. Ebro 338 Valley area), bagging would be cost-effective, especially in late-season cultivars.

339 Fruit quality parameters were within official and recommended ranges

340 All cultivars were harvested at commercial maturity date according to grower's 341 recommendation, and all quality characteristics on harvest day and after either storage 342 condition were within international recommendations (OECD 2010; European 343 Commission 2019). Fruit size on harvest day and after the different storage conditions 344 (Tables 3, 4, 5) was within specifications and accepted tolerances (European Commission 345 2019). There are no official recommendations for IAD, but all cultivars were within the 346 limits for nectarines at harvest date (0.3-1.5), as described by Reig et al. (2012). Values 347 of I_{AD} after postharvest storage were also within commercial maturity limits (0 - 1.5) 348 described by Spadoni et al. (2016). Published studies report that FF values should range 349 from 40 to 65 N after harvest, depending on the intended use, and decrease during 350 postharvest storage (Reig et al. 2017). This range was slightly below our results except for cv Albared (Table 3), but we also found that FF decreased during storage. The recommended FF at consumption ranges from 3 to 13 N (Crisosto 2002; Bonany et al. 2014), which was in line with the results of this study (except for cv Nectatinto), after both storage conditions.

355 SSC should be ≥ 8 °Brix (OECD 2010; European Commission 2019), although 356 some studies suggest at least 10 °Brix for consumer acceptance (e.g., Crisosto and 357 Crisosto 2005). Initial TA values ranged from 3.3 to 10 (which included the range of 358 sweet and nonsweet nectarine cultivars; Colaric et al. 2005; Reig et al. 2012), except for 359 Nectatinto, which had TA < 3.3, placing this cultivar in the sub-acid category (Iglesias 360 and Echeverría 2009; Reig et al. 2012). However, eating quality is better described by the 361 sugar-to-acid ratio (SSC:TA) rather than TA or SSC alone (Crisosto et al. 2006; Iglesias 362 and Echeverría 2009; Bonany et al. 2014). After storage, Nectatinto remained in the sub-363 acid cultivar under both postharvest storage conditions due to its high SSC/TA ratio (> 364 2). After storage at both postharvest conditions, also Albared nectarines became sub-acid, 365 especially after storage with lighting for both bagged and unbagged fruit.

366 Fruit quality was better maintained under storage with lighting

367 After postharvest storage under darkness, the bagging condition had more pronounced 368 effects on the quality of fruits stored under darkness than those stored under lighting 369 conditions in comparison to the initial quality (Tables 4 and 5). The bag effect was 370 conspicuous mainly in Fantasia and Albared under darkness and lighting. In addition to 371 changes attributable to cultivar (Iglesias and Echeverría 2009), a recurrent photoperiod 372 can reduce the response to subsequent stresses (Roeber et al. 2021). This suggested that 373 the effect of sunlight on fruit may have subsided after storage under darkness or lighting 374 in Fantasia and Albared.

375 The percentage change relative to initial quality suggested which postharvest 376 storage condition had a greater effect on fruit quality for each bagging condition. A 377 moderate weight loss was observed after both treatment storages, with some exceptions 378 (Figure 2). Loss of 5 to 8% of the fruit's water content may cause visual shrivelling in 379 peaches and nectarines, although the degree of shrivelling is cultivar-dependent (Crisosto 380 et al. 2020). In cv. Nectatinto, which presented the highest weight loss in almost all 381 conditions, shrivelling was barely appreciable (data not shown). Interestingly, storage 382 with lighting induced a lower weight reduction than darkness in half of the cultivars, 383 suggesting that lighting may have maintained fruit integrity. However, further research 384 integrating all factors that affect water loss in fruit is needed (Lufu et al. 2020).

385 The IAD and FF values differed little between storage conditions within each 386 bagging condition. The IAD values decreased greatly in all cultivars (53 and 92%), 387 probably because of the ongoing ripening during shelf life (Manganaris et al. 2017), 388 causing a decrease in the chlorophyll content and an increase in other pigments, such 389 anthocyanins (Bassi and Monet 2008; Ramina et al. 2008). No studies have reported the 390 effect of white lighting on these quality parameters, but a combination of white, blue, and 391 green light irradiation was reported to increase the anthocyanin content and phenylalanine 392 ammonia lyase activity in sweet cherries (Kokalj et al. 2019). In the work reported here, 393 FF was also reduced sharply (75 to 92%) in all cultivars. Flesh firmness is regulated by a 394 variety of cell wall modifications, including depolymerization and modifications of 395 polymers (Brummell et al. 2004). Beyond the white light spectrum, blue light treatment 396 reduces firmness in peaches during storage (Gong et al. 2015). Hence, investigation of 397 the effects of white light on factors related to ripening (e.g., I_{AD} and FF) is needed.

398 Depending on the bagging condition and cultivar, storage conditions with lighting
399 increased the SSC/TA ratio in some cases (Figure 2), suggesting that light irradiation can

400 favour the conversion of starch to sugars, and hence, decrease the acidity. Although there 401 are no previous reports of the effect of white artificial lighting on fruit quality, a treatment 402 with artificial blue light enhances total sugar content in peaches during storage (Gong et 403 al. 2015). Hence, the results reported here suggested that lighting during the postharvest 404 chain influenced the fruit quality, although dependent on the preharvest conditions 405 (bagging or not).

406 Conclusions

407 The results demonstrated that fruit bagging reduced the incidence of fruit decay during 408 postharvest storage, especially in fruits from orchards with high inoculum pressure (late-409 season cultivars). Fruit bagging was cost-effective for both late and early-mid season 410 cultivars. Postharvest storage under lighting increased fruit losses, and hence, storage 411 under darkness was preferable. Fruit quality on harvest day and after storage were within 412 international recommendations, irrespective of bagging conditions. Therefore, both 413 preharvest and postharvest management (bagged fruit and postharvest storage like the 414 described darkness condition) should be considered by growers and distributors for 415 sustainable fruit production and to ensure desirable fruit quality for the marketplace.

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419 **Disclosure statement**

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425 **References**

- 426 Ali MM, Anwar R, Yousef AF, Li B, Luvisi A, De Bellis L, Aprile A, Chen F. 2021.
- 427 Influence of bagging on the development and quality of fruits. Plants. 10(2):1–17.
- 428 Ballaré CL. 2014. Light regulation of plant defense. Annu Rev Plant Biol. 65:335–363.

429 Balsells-Llauradó M, Silva CJ, Usall J, Vall-llaura N, Serrano-Prieto S, Teixidó N,

430 Mesquida-Pesci SD, de Cal A, Blanco-Ulate B, Torres R. 2020. Depicting the battle

431 between nectarine and *Monilinia laxa*: the fruit developmental stage dictates the

432 effectiveness of the host defenses and the pathogen's infection strategies. Hortic Res.

- 433 7(167):1–15. http://dx.doi.org/10.1038/s41438-020-00387-w
- Balsells-Llauradó M, Torres R, Vall-llaura N, Casals C, Teixidó N, Usall J. 2021. Light
 intensity alters the behavior of *Monilinia* spp. *in vitro* and the disease development on
 stone fruit-pathogen interaction. Front Plant Sci. 12(666985):1–12.
- Baró-Montel N, Giné-Bordonaba J, Torres R, Vall-llaura N, Teixidó N, Usall J. 2020.
 Scrutinising the relationship between major physiological and compositional changes
 during 'Merrill O'Henry' peach growth with brown rot susceptibility. Food Sci Technol
 Int. 27(4):366–379.
- Baró-Montel N, Torres R, Casals C, Teixidó N, Segarra J, Usall J. 2019. Developing a
 methodology for identifying brown rot resistance in stone fruit. Eur J Plant Pathol.

- 444 Bassi D, Monet R. 2008. Botany and taxonomy. In: Layne DR, Bassi D., editors. peach
 445 Bot Prod uses. Wallingford: CAB International; p. 1–36.
 446 http://www.cabi.org/cabebooks/ebook/20083291730
- Blasi E, Pancino B, Passeri N, Franco S. 2017. Environmental and economic benefits of
 the preharvest fruit bagging technique: Trade-off evaluation in a Mediterranean area. Acta
 Hortic. 1160:313–318.
- 450 Bonany J, Carbó J, Echeverria G, Hilaire C, Cottet V, Iglesias I, Jesionkowska K,
- 451 Konopacka D, Kruczyńska D, Martinelli A, et al. 2014. Eating quality and European
- 452 consumer acceptance of different peach (Prunus persica (L.) Batsch) varieties. J Food,
- 453 Agric Environ. 12(1):67–72.
- Brummell DA, Dal Cin V, Crisosto CH, Labavitch JM. 2004. Cell wall metabolism
 during maturation, ripening and senescence of peach fruit. J Exp Bot. 55(405):2029–
 2039.
- 457 Bulletin OEPP/EPPO. 2020. PM 7/18 (3) *Monilinia fructicola*. EPPO Bull. 50(1):5–18.
- Bussi C, Plenet D, Merlin F, Guillermin A, Mercier V. 2015. Limiting brown rot
 incidence in peach with tree training and pruning. Fruits. 70(5):303–309.
- 460 Campbell D, Sarkhosh A, Brecht JK, Gillett-Kaufman JL, Liburd O, Melgar JC,
- 461 Treadwell D. 2021. Bagging organic peaches reduces physical injuries and storage decay
 462 with minimal effects on fruit quality. HortScience. 56(1):52–58.
 463 https://doi.org/10.21273/HORTSCI15391-20
- 464 Casals C, Guijarro B, De Cal A, Torres R, Usall J, Perdrix V, Hilscher U, Ladurner E,

- Smets T, Teixidó N. 2021. Field validation of biocontrol strategies to control brown rot
 on stone fruit in several European countries. Pest Manag Sci. 77(5):2502–2511.
 https://onlinelibrary.wiley.com/doi/10.1002/ps.6281
- 468 Casals C, Segarra J, De Cal A, Lamarca N, Usall J. 2015. Overwintering of *Monilinia*
- 469 spp. on mummified stone fruit. J Phytopathol. 163(3):160–167.
- 470 Colaric M, Veberic R, Stampar F, Hudina M. 2005. Evaluation of peach and nectarine
- 471 fruit quality and correlations between sensory and chemical attributes. J Sci Food Agric.
 472 85(15):2611–2616.
- 473 Crisosto CH. 2002. How do we increase peach consumption? In: Acta Hortic. Vol. 592;
 474 p. 601–605.
- 475 Crisosto CH, Costa G. 2008. Preharvest factors affecting peach quality. In: Layne DR,
 476 Bassi D, editors. Peach Bot Prod Uses. Cambridge, MA, USA: CAB International; p.
- 477 536–549.
- 478 Crisosto CH, Crisosto GM. 2005. Relationship between ripe soluble solids concentration
- 479 (RSSC) and consumer acceptance of high and low acid melting flesh peach and nectarine
- 480 (*Prunus persica* (L.) Batsch) cultivars. Postharvest Biol Technol. 38(3):239–246.
- 481 Crisosto CH, Crisosto GM, Garner D. 2006. Understanding tree fruit consumer
 482 acceptance. Acta Hortic. 682:865–870.
- 483 Crisosto CH, Echeverría G, Manganaris GA. 2020. Peach and Nectarine. In: Crisosto CH,
- 484 Crisosto GM, editors. Man postharvest Handl Mediterr tree fruits nuts. Wallingford: CAB
- 485 International; p. 53–87.
- 486 Eckert JW, Ratnayake M. 1983. Post-Harvest Physiology and Crop Preservation.

- 487 Lieberman M, editor. Boston, MA: Springer, Boston, MA.
 488 http://link.springer.com/10.1007/978-1-4757-0094-7
- 489 European Commission. 2019. COMMISSION DELEGATED REGULATION (EU)490 2019/428.
- 491 Faci JM, Medina ET, Martínez-Cob A, Alonso JM. 2014. Fruit yield and quality response
- 492 of a late season peach orchard to different irrigation regimes in a semi-arid environment.
- 493 Agric Water Manag. 143:102–112.
- 494 FAO. 2021. Database of Food and Agriculture Organization of the United Nations.
- 495 [accessed 2021 Apr 29]. http://www.fao.org/faostat/en/#data/QC/visualize
- 496 Garcia-Benitez C, Casals C, Usall J, Sánchez-Ramos I, Melgarejo P, De Cal A. 2020.
- 497 Impact of postharvest handling on preharvest latent infections caused by *Monilinia* spp.
- 498 in nectarines. J Fungi. 6(4):1–14.
- 499 Generaliat de Catalunya. 2022. Organic agri-food production. [accessed 2022 May 17].
- 500 http://pae.gencat.cat/ca/normativa/index.html
- 501 Giné-Bordonaba J, Echeverria G, Ubach D, Aguiló-Aguayo I, López ML, Larrigaudière
- 502 C. 2017. Biochemical and physiological changes during fruit development and ripening
- 503 of two sweet cherry varieties with different levels of cracking tolerance. Plant Physiol
- 504 Biochem. 111:216–225.
- 505 Glazebrook J. 2005. Contrasting mechanisms of defense against biotrophic and 506 necrotrophic pathogens. Annu Rev Phytopathol. 43(1):205–227.
- 507 Gong D, Cao S, Sheng T, Shao J, Song C, Wo F, Chen W, Yang Z. 2015. Effect of blue
- 508 light on ethylene biosynthesis, signalling and fruit ripening in postharvest peaches. Sci

- 509 Hortic (Amsterdam). 197:657–664.
- 510 Gradziel TM. 1994. Changes in susceptibility to brown rot with ripening in three
- 511 clingstone peach genotypes. J Amer Soc Hort Sci. 119(1):101–105.
- 512 Hayama H, Shimada T, Fujii H, Ito A, Kashimura Y. 2006. Ethylene-regulation of fruit
- 513 softening and softening-related genes in peach. J Exp Bot. 57(15):4071-4077.
- 514 https://academic.oup.com/jxb/article-abstract/57/15/4071/548558
- 515 Iglesias I, Echeverría G. 2009. Differential effect of cultivar and harvest date on nectarine
- 516 colour, quality and consumer acceptance. Sci Hortic (Amsterdam). 120(1):41–50.
- 517 Keske C, Treutter D, Neumüller M. 2014. Effect of bagging on brown rot incidence in
- 518 European Plum. Ecofruit 16th Int Conf Org Grow Proceedings, 17-19 Febr 2014,
 519 Hohenheim, Ger.(47):228–231.
- 520 Kokalj D, Zlatić E, Cigić B, Vidrih R. 2019. Postharvest light-emitting diode irradiation
- 521 of sweet cherries (*Prunus avium* L.) promotes accumulation of anthocyanins. Postharvest
- 522 Biol Technol. 148(October 2018):192–199.
- 523 Kou J, Wei Y, He X, Xu J, Xu F, Shao X. 2018. Infection of post-harvest peaches by
- 524 Monilinia fructicola accelerates sucrose decomposition and stimulates the Embden-
- 525 Meyerhof–Parnas pathway. Hortic Res. 5(46). http://dx.doi.org/10.1038/s41438-018-
- 526 0046-x
- 527 Liu H, Cao X, Liu X, Xin R, Wang J, Gao J, Wu B, Gao L, Xu C, Zhang B, et al. 2017.
- 528 UV-B irradiation differentially regulates terpene synthases and terpene content of peach.
- 529 Plant Cell Environ. 40(10):2261–2275.
- 530 Lufu R, Ambaw A, Opara UL. 2020. Water loss of fresh fruit: Influencing pre-harvest,

- 531 harvest and postharvest factors. Sci Hortic. 272:109519.
- 532 Luo Y, Michailides TJ, Morgan DP, Krueger WH, Buchner RP. 2005. Inoculum
- 533 dynamics, fruit infection, and development of brown rot in prune orchards in California.
- 534 Phytopathology. 95(10):1132–1136.
- 535 Manganaris GA, Crisosto CH. 2020. Stone fruits: Peaches, nectarines, plums, apricots.
- 536 In: Gil M, Beaudry R, editors. Control Modif Atmos Fresh Fresh-Cut Prod.: Elsevier Inc.;
- 537 p. 311–322. http://dx.doi.org/10.1016/B978-0-12-804599-2.00017-X
- 538 Manganaris GA, Drogoudi P, Goulas V, Tanou G, Georgiadou EC, Pantelidis GE,
- 539 Paschalidis KA, Fotopoulos V, Manganaris A. 2017. Deciphering the interplay among
- 540 genotype, maturity stage and low-temperature storage on phytochemical composition and
- 541 transcript levels of enzymatic antioxidants in Prunus persica fruit. Plant Physiol
- 542 Biochem. 119:189–199. https://doi.org/10.1016/j.plaphy.2017.08.022
- 543 Mari M, Spadaro D, Casals C, Collina M, De Cal A, Usall J. 2019. Stone Fruits. In: Palou
- 544 L, Smilanick JL, editors. Postharvest Dis Fresh Hortic Prod. New York: CRC Press; p.
 545 111–140.
- 546 Minas IS, Tanou G, Molassiotis A. 2018. Environmental and orchard bases of peach fruit
 547 quality. Sci Hortic (Amsterdam). 235(January):307–322.
 548 https://doi.org/10.1016/j.scienta.2018.01.028
- Müller M, Munné-Bosch S. 2015. Ethylene response factors: A key regulatory hub in
 hormone and stress signaling. Plant Physiol. 169(1):32–41.
- 551 Mustafa MH, Bassi D, Corre M-N, Lino LO, Signoret V, Quilot-Turion B, Cirilli M.
- 552 2021. Phenotyping brown rot susceptibility in stone fruit: A literature review with

- 553 emphasis on peach. Horticulturae. 7(5):115.
- 554 OECD. 2010. Peaches and nectarines, International Standards for Fruit and Vegetables:
- 555 OECD Publishing, Paris; [accessed 2021 May 8]. www.oecd.org/bookshop
- 556 De Oliveira Lino L, Génard M, Signoret V, Quilot-Turion B. 2016. Physical host factors
- 557 for brown rot resistance in peach fruit. Acta Hortic. 1137:105–112.
- 558 Porat R, Lichter A, Terry LA, Harker R, Buzby J. 2018. Postharvest losses of fruit and
- 559 vegetables during retail and in consumers' homes: Quantifications, causes, and means of
- 560 prevention. Postharvest Biol Technol. 139:135–149.
- 561 Ramina A, Tonutti P, Mcglasson W. 2008. Ripening, Nutrition and Postharvest
- 562 Physiology. In: Layne DR, Bassi D, editors. Peach Bot Prod Uses. Wallingford: CAB
 563 International; p. 550–574.
- Reig G, Alegre S, Cantín CM, Gatius F, Puy J, Iglesias I. 2017. Tree ripening and
 postharvest firmness loss of eleven commercial nectarine cultivars under Mediterranean
 conditions. Sci Hortic. 219:335–343. http://dx.doi.org/10.1016/j.scienta.2017.03.001
- Reig G, Alegre S, Iglesias I, Echeverría G, Gatius F. 2012. Fruit quality, colour
 development and index of absorbance difference (I_{AD}) of different nectarine cultivars at
 different harvest dates. Acta Hortic. 934:1117–1126.
- 570 Reig G, Iglesias I, Gatius F, Alegre S. 2013. Antioxidant capacity, quality, and
- 571 anthocyanin and nutrient contents of several peach cultivars [*Prunus persica* (L.) *Batsch*]
- 572 grown in Spain. J Agric Food Chem. 61(26):6344–6357.
- 573 Roeber VM, Bajaj I, Rohde M, Schmülling T, Cortleven A. 2021. Light acts as a stressor
- and influences abiotic and biotic stress responses in plants. Plant Cell Environ. 44(3):645–

- 575 664. https://onlinelibrary.wiley.com/doi/10.1111/pce.13948
- 576 Santin M, Giordani T, Cavallini A, Bernardi R, Castagna A, Hauser MT, Ranieri A. 2019.
- 577 UV-B exposure reduces the activity of several cell wall-dismantling enzymes and affects
- 578 the expression of their biosynthetic genes in peach fruit (*Prunus persica* L., cv. Fairtime,
- 579 melting phenotype). Photochem Photobiol Sci. 18(5):1280–1289.
- 580 Santin M, Neugart S, Castagna A, Barilari M, Sarrocco S, Vannacci G, Schreiner M,
- 581 Ranieri A. 2018. UV-B Pre-treatment alters phenolics response to Monilinia fructicola
- 582 infection in a structure-dependent way in peach skin. Front Plant Sci. 9(1598).
- 583 Santin M, Ranieri A, Hauser M-T, Miras-Moreno B, Rocchetti G, Lucini L, Strid Å,
- 584 Castagna A. 2021. The outer influences the inner: Postharvest UV-B irradiation
- 585 modulates peach flesh metabolome although shielded by the skin. Food Chem. 338(July
- 586 2020):127782. https://linkinghub.elsevier.com/retrieve/pii/S0308814620316447
- 587 Sharma RR, Reddy SVR, Jhalegar M. J. 2014. Pre-harvest fruit bagging: A useful
- approach for plant protection and improved post-harvest fruit quality A review. J Hortic
 Sci Biotechnol. 89(2):101–113.
- 590 Spadoni A, Cameldi I, Noferini M, Bonora E, Costa G, Mari M. 2016. An innovative use
- 591 of DA-meter for peach fruit postharvest management. Sci Hortic (Amsterdam). 201:140–
- 592 144. http://dx.doi.org/10.1016/j.scienta.2016.01.041
- 593 Usall J, Casals C, Sisquella M, Palou L, De Cal A. 2015. Alternative technologies to
- 594 control postharvest diseases of stone fruits. Stewart Postharvest Rev. 11(4):1–6.
- 595 Van Der Ent S, Pieterse CMJ. 2012. Ethylene: multi-tasker in plant-attacker interactions.
- 596 In: Michael T. McManus, editor. Annu Plant Rev. Vol. 44: Blackwell Publishing Ltd; p.

597 343–377.

Villarino M, Melgarejo P, Usall J, Segarra J, Lamarca N, De Cal A. 2012. Secondary
inoculum dynamics of *Monilinia* spp. and relationship to the incidence of postharvest
brown rot in peaches and the weather conditions during the growing season. Eur J Plant
Pathol. 133(3):585–598.

- Wang K, Shao X, Gong Y, Zhu Y, Wang H, Zhang X, Yu D, Yu F, Qiu Z, Lu H. 2013.
- 603 The metabolism of soluble carbohydrates related to chilling injury in peach fruit exposed
- 604 to cold stress. Postharvest Biol Technol. 86:53–61.
- 605 http://dx.doi.org/10.1016/j.postharvbio.2013.06.020
- 606 Zhou H, Yu Z, Ye Z. 2019. Effect of bagging duration on peach fruit peel color and key
- 607 protein changes based on iTRAQ quantitation. Sci Hortic (Amsterdam). 246(October
- 608 2018):217–226. https://doi.org/10.1016/j.scienta.2018.10.072
- 609
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- **Table 1.** Orchard characteristics for one hectare and average costs (bags, labour, removal
- 612 of bags) in Ebro Valley area, Catalonia, Spain.

Orchard data		
	Trees ha ⁻¹	625
	Fruits tree ⁻¹	180
	Average fruit weight (kg fruit ⁻¹)	0.2
	Fruit yield ha ⁻¹ (kg ha ⁻¹)	22,500
	Price kg ⁻¹ (\notin kg ⁻¹)	1.2
	Total production (€ ha ⁻¹)	27,000
Bagged orchard		
Base data	Cost of white paper bags (€ bag ⁻¹)	0.008
	Bagging rate by worker (bags hour ⁻¹)	400
	Labour time input for bagging (hours ha ⁻¹)	281.3
	Average cost for labour (€ hour ⁻¹)	10
	Costs of bag removal (€ kg ⁻¹)	0.085
Costs of bagging		
	Cost of bags (€ ha ⁻¹)	900.00
	Cost of labour for bagging (€ ha ⁻¹)	2812.50
	Cost of labour for bag removal (€ ha ⁻¹)	1912.50
	Total cost of bagging (€ ha ⁻¹)	5625.00

618 **Table 2.** Cost-effectiveness of fruit bagging between a bagged and unbagged orchard.

619 Percentages of postharvest losses were taken from Figure 1.

620

Bagged orchard Assuming 0% 21,375€ Production - cost of postharvest losses ha⁻¹ bagging Unbagged orchard **Differences between** bagged and unbagged production Mid-early cv. (assuming 25% losses) Losses 6,750 € ha⁻¹ Final production 20,250 € Bagged - unbagged 1,125€ ha⁻¹ ha⁻¹ Late cv. (assuming 60% losses) 16,200€ Losses ha⁻¹ 10,800€ Final production Bagged - unbagged 10,575€ ha⁻¹ ha⁻¹

Table 3. Fruit quality on harvest day of four nectarine cultivars. Quality parameters are listed with the measurement unit in brackets. Weight, cheek diameter (CD), single index of absorbance difference (I_{AD}), flesh firmness (FF), soluble solids content (SSC), titratable acidity (TA), SSC/TA ratio, and ethylene levels of preharvest bagged fruit (B) and unbagged fruit (UB). Values represent the mean (4 replicates, 5 fruits each) \pm the standard error of the mean. Lower case letters indicate significant differences ($p \le 0.05$) between bagging conditions within each cultivar. No letter indicates no significant differences.

Cultivar	Pre-	Weight (g)	CD (mm)	I _{AD}	FF (N)	SSC (°Brix)	TA (g L ⁻¹)	SSC/TA	Ethylene
	harvest							ratio	$(\mathbf{nL} \mathbf{kg}^{-1} \mathbf{h}^{-1})$
Fantasia	В	144.5 ± 7.1	63.4 ± 1.1	0.9 ± 0.0	75.7 ± 1.2 a	12.2 ± 0.1 a	8.1 ± 0.6	1.6 ± 0.1	$139.8 \pm 67.3 \text{ b}$
	UB	160.1 ± 7.2	65.4 ± 1.0	1.0 ± 0.0	$70.2\pm1.8~b$	$11.9\pm0.1~b$	9.7 ± 0.4	1.3 ± 0.1	961.6 ± 298.1 a
Venus	В	$178.9\pm9.8~b$	$67.3\pm1.3~b$	0.7 ± 0.0 a	$68.8\pm1.3~b$	$10.6\pm0.2~b$	$6.6\pm0.6\ b$	1.5 ± 0.1	58.2 ± 8.2
	UB	211.7 ± 10.2 a	$72.0 \pm 1.2 \text{ a}$	$0.6\pm0.0\;b$	$74.1 \pm 2.1 \text{ a}$	11.5 ± 0.2 a	$8.6 \pm 0.2 a$	1.3 ± 0.0	54.9 ± 2.8
Nectatinto	В	249.0 ± 11.1	77.8 ± 1.2	$1.1\pm0.0\ b$	73.7 ± 3.7	11.7 ± 0.3	2.9 ± 0.1	4.2 ± 0.2	208.9 ± 40.0
	UB	221.2 ± 10.7	75.4 ± 1.0	1.2 ± 0.0 a	68.9 ± 3.8	12.3 ± 0.3	3.0 ± 0.1	4.1 ± 0.1	158.5 ± 24.1
Albared	В	251.5 ± 7.1 a	$77.1 \pm 0.7 \text{ a}$	$0.7\pm0.0\ b$	59.6 ± 2.0	14.8 ± 0.3	7.9 ± 0.5	2.0 ± 0.1	237.9 ± 103.0
	UB	$200.0\pm8.3~b$	$71.1\pm1.0\ b$	$0.8\pm0.0\;a$	58.9 ± 3.1	13.7 ± 0.6	7.3 ± 0.7	1.9 ± 0.1	193.6 ± 65.4

Table 4. Fruit quality of four nectarine cultivars after darkness postharvest storage. The storage period (22 ± 2 °C and 50-90 % RH) was 9 days for Fantasia and Venus cultivars, and 7 days for Nectatinto and Albared cultivars. Weight, cheek diameter (CD), single index of absorbance difference (I_{AD}), flesh firmness (FF), soluble solids content (SSC), titratable acidity (TA), and SSC/TA ratio. Values represent the mean (4 replicates, 5 fruits each) \pm the standard error of the mean. Lower case letters indicate significant differences ($p \le 0.05$) between bagging conditions, within each cultivar. No letter indicates no significant differences.

Cultivar	Pre-	Weight (g)	CD (mm)	IAD	FF (N)	SSC (°Brix)	TA (g L ⁻¹)	SSC/TA
	harvest							ratio
Fantasia	В	$116.9\pm6.4~b$	$57.3\pm1.0~b$	$0.3 \pm 0.0 a$	13.9 ± 1.7 a	12.7 ± 0.2	7.1 ± 0.6	1.8 ± 0.2
	UB	$134.7 \pm 5.5 \text{ a}$	60.6 ± 0.9 a	$0.2\pm0.0\ b$	$7.4\pm0.4\;b$	12.6 ± 0.2	8.1 ± 0.3	1.6 ± 0.1
Venus	В	160.1 ± 7.9	62.3 ± 1.2	0.2 ± 0.0	$7.7 \pm 0.6 a$	12.8 ± 0.4	7.3 ± 0.2	1.8 ± 0.1
	UB	177.6 ± 5.7	66.7 ± 0.9	0.2 ± 0.0	$6.1\pm0.3~b$	12.5 ± 0.5	7.8 ± 0.5	1.6 ± 0.1
Nectatinto	В	168.0 ± 8.0	66.7 ± 1.2	0.7 ± 0.1	15.5 ± 1.1	14.2 ± 0.5	3.3 ± 0.1	4.5 ± 0.3
	UB	161.2 ± 8.6	66.7 ± 1.2	0.5 ± 0.1	17.2 ± 1.4	13.0 ± 0.5	3.1 ± 0.2	4.2 ± 0.2
Albared	В	175.0 ± 6.2	65.3 ± 1.0	$0.1 \pm 0.0 \text{ b}$	$6.0 \pm 0.4 \text{ b}$	15.7 ± 0.3 a	7.0 ± 0.4 a	2.2 ± 0.1 b
	UB	174.0 ± 5.9	66.2 ± 0.8	0.1 ± 0.0 a	$8.4\pm0.8\;a$	$14.7\pm0.4\ b$	$4.5\pm0.3\;b$	3.4 ± 0.4 a

Table 5. Fruit quality of four nectarine cultivars after postharvest storage under light. The storage period 22 ± 2 °C and 50-90 % RH) was 9 days for Fantasia and Venus cultivars, and 7 days for Nectatinto and Albared cultivars. Weight, cheek diameter (CD), single index of absorbance difference (I_{AD}), flesh firmness (FF), soluble solids content (SSC), titratable acidity (TA), and SSC/TA ratio. Values represent the mean (4 replicates, 5 fruits each) \pm the standard error of the mean. Lower case letters indicate significant differences ($p \le 0.05$) between bagging conditions, within each cultivar. No letter indicates no significant differences.

Cultivar	Pre- harvest	Weight (g)	CD (mm)	IAD	FF (N)	SSC (°Brix)	TA (g L ⁻¹)	SSC/TA ratio
Fantasia	В	137.4 ± 7.9	62.4 ± 0.9	0.3 ± 0.1	$9.0 \pm 0.7 \text{ a}$	12.7 ± 0.2 a	7.5 ± 0.2 a	1.7 ± 0.0
	UB	152.9 ± 4.8	63.8 ± 0.9	0.2 ± 0.0	$6.6\pm0.4\ b$	$11.9\pm0.2~b$	$5.8\pm0.2\;b$	2.0 ± 0.1
Venus	В	168.4 ± 6.1	64.8 ± 1.0	0.2 ± 0.0	7.6 ± 0.6	11.8 ± 0.2	8.2 ± 0.3	1.5 ± 0.1
	UB	181.8 ± 7.3	67.1 ± 0.9	0.2 ± 0.0	7.1 ± 0.5	12.8 ± 0.2	7.5 ± 0.3	1.7 ± 0.1
Nectatinto	В	166.6 ± 10.6	66.8 ± 1.4 b	0.5 ± 0.1	15.9 ± 1.5	12.5 ± 0.5	3.2 ± 0.3	4.2 ± 0.6
	UB	185.5 ± 6.0	70.1 ± 0.9 a	0.6 ± 0.1	15.0 ± 1.1	13.5 ± 0.4	3.0 ± 0.2	4.8 ± 0.3
Albared	В	227.1 ± 12.3 a	72.3 ± 1.3	0.1 ± 0.0	7.8 ± 0.7	15.6 ± 0.5	4.9 ± 0.3	3.2 ± 0.2
	UB	$183.1 \pm 7.2 \text{ b}$	69.0 ± 1.0	0.1 ± 0.1	7.1 ± 1.1	14.7 ± 0.7	4.1 ± 0.2	3.3 ± 0.0



Figure 1. Fruit decay of nectarines after darkness and light postharvest conditions. Incidence of fruit decay (%) during storage in unbagged (UB) and preharvest bagged (B) fruit stored under darkness or light, of Fantasia (A), Venus (B), Nectatinto (C), and Albared (D) cultivars. Bars represent the mean of fruits with disease symptoms (n = 4 replicates, 5 fruits per replicate), and error bars represent the standard error of the means. Different lower-case letters indicate significant differences ($p \le 0.05$) of fruit decay incidence among postharvest storage × bagging conditions at each time point. No letters indicate no significant differences. Asterisks indicate significant differences ($p \le 0.05$) at each time point between bagged and unbagged fruit for each postharvest condition.



Figure 2. Changes in quality characteristics relative to initial fruit quality of nectarine cultivars after postharvest storage. Percentage change calculated for bagged (A-C) and unbagged (D-F) fruit, stored under darkness or light. Weight and cheek diameter (CD) (A, D); I_{AD} and flesh firmness (FF) (B, E); SSC/ TA ratio (C, F). Bars represent the mean (n = 4 replicates, 5 fruits each) and error bars represent the standard error of the means. Asterisks indicate significant difference $(p \le 0.05)$ between postharvest storage conditions, within each cultivar and bagging condition.