



# Article Identification of Vitis vinifera L. Local Cultivars Recovered in Andalusia (Spain) by Using Microsatellite Markers

Ana Jiménez-Cantizano <sup>1,\*</sup>, Anna Puig-Pujol <sup>2</sup> and Rosa Arroyo-García <sup>3</sup>

- <sup>1</sup> Department of Chemical Engineering and Food Technology, Vegetal Production Area, University of Cadiz, Agrifood Campus of International Excellence (CeiA3), IVAGRO, P.O. Box 40, 11510 Puerto Real, Spain
- <sup>2</sup> IRTA-INCAVI Institute of Agrifood Research and Technology-Catalan Institute of Vine and Wine, Department of Enological Research, Plaça Àgora 2, 08720 Vilafranca del Penedès, Spain
- <sup>3</sup> CSIC-INIA (CBGP) Centro de Biotecnología y Genómica de Plantas, UPM-INIA, Parque Científico y Tecnológico de la UPM Campus de Montegancedo, 28223 Madrid, Spain
- \* Correspondence: ana.jimenezcantizano@uca.es; Tel.: +34-9-5601-6470

**Abstract:** In Andalusia (Spain), there are different wine regions that have a great recognized tradition. In these regions, the cultivation of the vine is ancient and there are still vineyards planted with local varieties of *Vitis vinifera* L. that have not yet been identified. The aim of this research study was to identify 49 accessions of grapevine collected in the districts of four provinces in Andalusia (Spain). All samples were genotyped with 20 microsatellite markers in order to ascertain the identity and analyze the genetic diversity of the collected material. In total, 30 different genotypes were obtained, 22 of them which were identified with named, known varieties by comparison to the Spanish or European microsatellite databases, and eight which are referred to as new genotypes. All loci were polymorphic, and a total of 159 alleles were detected, ranging from 4 to 12 alleles per locus, with an average allele number of 7.95. The overall observed heterozygosity was 0.763 and was slightly higher than expected (0.715), while the gene diversity per locus varied between 0.167 (VVIN73) and 0.967 (VVMD5). A dendrogram representing the genetic similarities among cultivars was depicted using the UPGMA method to investigate their relationships. The eight new genotypes identified in this research work could represent ancient local varieties in danger of extinction. These new cultivars may be used to determine original wines.

Keywords: grapevine; genetic characterization; synonymies

## 1. Introduction

The Andalusia (Spain) region, in the south of the Iberian Peninsula, is one of the most ancient and important wine regions in Spain [1]. Archaeological, paleobotanical, and historical sources confirm that grapevines were spread and cultivated for a long time in this area. The presence of the species *Vitis vinifera* L. has been verified by pollen analysis performed in different Phoenician sites of Andalusia located in the provinces of Cádiz, Málaga and Almería [2]. In addition, numerous archaeological remains have been found at these sites, which may be associated with the existence of a wine industry [3–5]. The first evidence of planting techniques characteristic of protohistoric viticulture in the west has been documented in an archaeological site located in Huelva (Andalusia) dating back to the 1st millennium BC [6].

There are many citations that reference the diversity of grapevine (*Vitis vinifera* L.) varieties grown in Andalusia. Roxas Clemente [7], in his paper *Essay of common grapevine varieties that are growing in Andalusia*, includes 119 varieties grouped in two sections and 15 tribes. In 1831, James Busby, considered the "father of Australian viticulture" introduced 678 varieties in Australia [8]. These varieties originated in France and Spain. According to Morilla Critz [8], at least half of these varieties were from Andalusia.



Citation: Jiménez-Cantizano, A.; Puig-Pujol, A.; Arroyo-García, R. Identification of *Vitis vinifera* L. Local Cultivars Recovered in Andalusia (Spain) by Using Microsatellite Markers. *Horticulturae* **2023**, *9*, 316. https://doi.org/10.3390/ horticulturae9030316

Academic Editor: Paolo Sabbatini

Received: 1 February 2023 Revised: 22 February 2023 Accepted: 23 February 2023 Published: 28 February 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Nevertheless, the genetic diversity of the Andalusia grapevine (*Vitis vinifera* L.) has been declining due to the phylloxera (*Daktulosphaira vitifoliae*) attack of the late 19th century [9], when severe regulations were approved, and the grapevine varieties authorized for wine production were restricted and the vineyard was restructured, frequently stimulated by subsidies. In Spain, previous to this vineyard restructuring, which begin in the 1970s, all vines were grafted in the field with mass-selected *Vitis vinifera* material from older vine-yards, which often included different varieties [10]. With the aim of preserving grapevine phytogenetic resources, numerous studies on the surveying, localization, characterization, and maintaining of cultivars in germplasm banks are being carried out worldwide [11–20]. In Andalusia, a germplasm bank was established in 1940, and it was replanted between 1984 and 1987, and the number of accessions substantially increased [21]. Actually, this collection preserves 1417 accessions according to the *Vitis* International Variety Catalogue (*VIVC*, www.vivc.de accessed on 26 December 2022) [22].

The recovery of autochthonous or local varieties allows a genetic, ecological and agronomic enrichment capable of dealing with various diseases, improving the adaptation to edaphoclimatic conditions [23] or facilitating the adaptation in the face of future market changes [24]. For this reason, the accurate identification of local cultivars and their conservation could prevent their disappearance and preserve them for future needs. Traditionally, the identification of grape varieties has been based on the morphological features of vegetative and reproductive structures [25], but phenotypic traits are not sufficiently reliable for the classification of closely related varieties due to genotype–environment interactions [26]. Therefore, molecular characterization is the favoured technique for varietal identification. At the present time, there are different molecular markers available to carry out a molecular identification of a grape variety. However, microsatellites or Simple Sequence Repeats (SSRs) markers are the most used for this purpose [27,28]. In this sense, microsatellite markers have been widely used to identify and genotype grapevine cultivars collected in old vineyards of the Iberian Peninsula [29–31]. In addition, SSRs have been used for studies of genetic diversity and genetic relationships [32].

The main objective of this research work is focused on the molecular identification of a total of 49 vine accessions collected in old Andalusian vineyards. The genotyping of these accessions could help to detect new local cultivars growing in Andalusia aiming to provide a solid basis to develop a regional germplasm collection to protect local biodiversity.

## 2. Materials and Methods

## 2.1. Plant Material

After prospecting more than 200 vineyards throughout the provinces of Almería, Cádiz, and Huelva y Málaga of the Andalusia region (Spain), those plants that were not visually identified as common varieties cultivated in Andalusia were sampled and placed in the germplasm bank at the Rancho de la Merced. This grapevine collection is located in Jerez de la Frontera (Cádiz, Spain) (36°41′10″ N; 6°08′10″ W; alt. 20 m). The list of the 49 accessions used in this study are shown in the Supplemental Table S1. Each accession was identified with a code of three letters and a number. The initials correspond to the name of the municipality where it was collected.

Two internationally known cultivars ('Cabernet Sauvignon' and 'Syrah') were also included to compare the genetic profiles obtained with the different published databases.

### 2.2. Molecular Analysis

DNA was extracted from young leaves collected from each accession and stored at -80 °C, using a DNeasy Plant Mini Kit (Qiagen, Hilden, Germany). A genotypic characterization was performed for 20 nuclear microsatellite loci located in the 19 linkage groups of grapevine genome VMC1b11, VMC4F3-1 (*Vitis* Microsatellite Consortium); VVMD5, VVMD7, VVMD21, VVMD24, VVMD25, VVMD27, VVMD28, VVMD32 [33,34]; VVS2 [35]; VVIB01, VVIH54, VVIN16, VVIN73, VVIP31, VVIP60, VVIQ52, VVIV37, and VVIV67 [36]. Two multiplex PCR tests were set up to amplify the 20 microsatellite loci in a 20 µL reaction mix according to Vargas et al. [37]. PCR reactions were carried out in the 44 Applied Biosystems 9700 thermocycler.

Amplified products were separated by capillary electrophoresis using an automated sequencer (ABI Prism 3130, Applied Biosystems, Foster City, CA, USA). Fluorescently labelled fragments were detected and sized using GeneMapper v. 3.7 software (Applied Biosystems), and fragment lengths were determined with the help of internal size standards (GeneScan-500 LIZTM, Applied Biosystems, Foster City, CA, USA).

The identification of redundant genotypes was determined by comparing microsatellite genotypes with data contained in the Spanish microsatellite grapevine databases Rancho de la Merced [38–40] and the *Vitis* Germplasm Bank (BGV) at the Finca El Encín (IMIDRA, Alcalá de Henares, Spain) [41–43] and other European databases [22,44]. Genotype comparisons were carried out using the Microsatellite toolkit v. 9.0 software package [45].

### 2.3. Data Analysis

# 2.3.1. Genetic Diversity Analyses

For the calculation of the number of alleles (Na), expected (He) and observed (Ho) heterozygosity, frequency of null alleles (r) and probability of identity (PI), the GENALEX software [46] was used. The polymorphism information content (PIC) of each microsatellite loci was determined using an online tool [47].

## 2.3.2. Genetic Relationships among Cultivars

Genetic distances between grapevine genotypes were calculated as [-ln (proportion shared alleles)] using Microsat [48]. The obtained data was used for the construction of a dendrogram using the programs EXE from the PHYLIP package software [49] and MEGA version 7 [50].

## 3. Results and Discussion

## 3.1. Microsatellite Analysis and Genetic Diversity

The molecular analysis performed with the 49 studied accessions resulted in 30 non-redundant genotypes (Table 1). These genotypes were used for the calculation of genetic parameters (Table 2) in order avoid overestimation. A total of 159 alleles, ranging from 12 in VVMD7 and four in VVIN73, were detected, with an average of eight alleles per locus, similar to the mean Na attained by Fernández-González et al. [51]. The most frequent allele was VVIN73-264, which showed a frequency up to 90%, and 27 alleles were unique.

The expected heterozygosity (He, gene diversity) ranged from 0.185 at locus VVIN73 to 0.866 at locus VVIP31, with a mean value of 0.715. The observed heterozygosity (Ho) varied between 0.167 at locus VVIN73 and 0.967 at locus VVMD5. For 16 loci, Ho was higher than He, and the probability of null alleles was always negative, except for VMC4F31, VVMD21, VVMD25, VVMD28, VVIN73 and VVIP60. Samples in which only one single allele per locus was detected were considered as homozygous genotypes instead of heterozygous with a null allele. The VVIN73 and VVIP31 markers displayed the minimum (0.1769) and maximum (0.8522) *PIC* values, respectively. The 20 microsatellite loci showed a mean *PIC* value of 0.67241.

The 20 microsatellite loci used reflected a high discrimination power and a low probability that two randomly chosen individuals had identical genotypes using the 20 loci (PI.  $1.74 \times 10^{-19}$ ). This indicates the probability that two of the 30 varieties analyzed randomly were chosen to share the same genotype using the set of these 20 microsatellite loci.

Cultivars <sup>a</sup>	Accession Code	VVIB01	U VM	C1b11	VMC4	F31 V	/MD5	VVN	1D7	VVM	ID21	VVM	1D24	VVN	4D25	VVN	1D27	VVM	D28	VVM	D32	VVIE	154	VVIN	116	VVIN	173	vvi	P31	VVII	<b>°</b> 60	VVI	Q52	vv	S2	VVI	V37	vvi	IV67
AFUS ALI	Man-4	291 2	95 184	184	168	190 22	4 228	236	246	255	255	209	209	246	252	186	186	260	260	256	270	166	178	151	151	264	264	174	184	318	318	83	85	131	133	151	163	358	362
AHMEUR BOU AHMEUR	Lau-5, Lau-6	291 2	95 184	184	190	190 22	8 236	236	246	249	255	209	209	252	264	184	194	250	256	250	254	166	168	151	153	264	264	184	192	322	322	83	89	133	146	161	161	358	366
ALICANTE HENRI BOUSCHET	Lau-8	291 2	95 182	188	174	206 22	4 236	236	240	243	249	209	211	240	240	182	194	246	262	248	270	164	168	151	159	256	264	176	184	322	322	83	89	131	144	161	171	358	364
ATTIKA SEEDLESS	Pla-2	291 2	95 166	184	184	206 23	5 238	250	252	249	249	213	217	246	252	186	194	246	246	254	270	164	166	151	157	256	264	186	188	320	326	85	85	133	151	163	181	372	379
BEBA	Chu-2	291 2	95 184	188	188	188 23	4 238	240	246	249	255	209	211	252	252	182	190	246	260	254	270	164	166	151	153	256	264	190	192	318	322	83	85	133	142	161	163	366	372
BOBAL	Ron-1	291 2	95 184	188	174	184 22	5 232	236	240	243	243	209	211	240	264	182	190	236	262	248	270	166	168	151	153	264	264	176	186	326	326	83	85	144	146	159	163	358	362
CARIGNAN NOIR	Lau-17	291 2	95 174	184	180	188 22	4 226	236	236	249	253	209	215	240	252	182	186	250	260	248	250	166	166	151	153	264	264	176	176	318	326	83	85	142	144	163	171	362	375
CAYETANA BLANCA	Alb-1, Chu-3, Lau-12, Lau-13, Lau-10, Man-3	291 3	07 168	188	188	204 23	2 234	240	246	249	255	209	211	240	252	182	182	236	250	250	254	166	168	151	153	264	264	176	180	322	322	85	89	135	144	163	177	366	375
COJONATA	Ron-2	291 2	91 184	188	174	188 22	8 234	236	240	243	255	209	209	240	264	182	194	250	250	250	270	166	168	153	153	264	264	176	184	322	322	83	85	144	146	159	163	358	366
CORNICHÓN BLANC	Ins-2, Pla-1, Rot-2	291 2	91 166	184	168	204 23	4 244	244	246	249	265	209	213	238	246	180	182	246	260	256	260	160	166	149	153	264	264	182	196	318	318	89	95	144	149	173	181	348	358
DONA MARÍA	Ins-4	291 2	91 166	184	190	206 22	4 226	236	248	255	255	209	213	246	252	186	194	236	270	256	262	166	178	149	151	264	264	174	188	318	322	83	85	133	149	163	175	362	375
IMPERIAL NAPOLEÓN	Ins-3	291 2	95 166	188	188	190 23	2 236	246	248	249	255	209	211	238	252	184	194	246	250	254	270	166	166	153	153	264	264	184	186	322	322	85	89	131	133	153	161	362	366
JACQUEZ	Alb-2	289 2	91 178	184	184	184 22	5 240	236	238	237	249	209	217	254	256	180	190	232	238	250	250	166	166	149	151	262	270	176	182	316	318	85	85	137	142	159	171	334	364
JAÉN TINTO	Com-2, Lau-2, Lau-7	291 2	91 184	188	174	188 23	2 238	236	240	249	249	209	209	240	240	182	190	246	250	254	256	166	168	151	153	264	264	180	192	318	322	85	89	131	144	167	177	362	366
LISTÁN PRIETO	Ron-4	291 2	91 184	184	168	174 22	5 238	236	246	243	249	209	209	238	240	186	190	236	246	254	256	166	168	151	151	256	264	176	192	318	322	85	89	131	133	163	167	362	364
MANTEUDO	Chu-1, Man-1, Rot-1	291 3	07 184	184	168	176 22	) 224	236	246	243	265	209	211	252	252	182	182	246	246	270	270	166	168	151	151	264	264	176	176	322	322	85	89	142	142	163	167	366	375
MOLINERA	Ins-1	291 2	91 186	188	168	188 23	2 236	240	246	249	249	211	211	240	252	182	194	236	260	250	270	166	168	151	153	264	264	186	190	318	322	85	89	135	144	161	163	372	375
PEDRO XIMENES	Chu-5	291 3	07 168	188	168	174 23	4 238	236	236	243	249	209	213	240	246	182	186	260	266	248	270	166	166	149	153	264	264	176	188	322	322	89	89	131	144	163	177	364	366
ROAL	Chu-4, Man-2	291 3	07 168	184	204	206 23	4 236	246	250	255	255	209	209	240	254	182	184	246	246	250	254	168	168	151	153	264	264	176	180	318	322	85	89	135	142	163	167	358	366
ROME TINTO	Com-4	291 2	95 184	188	188	190 23	5 238	236	236	243	249	209	209	240	252	182	194	238	260	254	270	166	166	153	153	264	264	176	190	318	326	83	89	135	144	163	171	366	375
TINTO VELASCO	Lau-1, Lau-14, Lau-15, Lau-18, Lau-19	291 2	91 172	184	188	206 22	8 236	232	250	255	255	209	215	238	238	180	186	250	262	250	250	164	166	151	151	264	264	184	190	322	322	89	89	131	131	159	159	358	375
XARELLO	Lau-9	291 2	95 184	184	168	180 23	4 238	236	240	249	253	209	209	238	252	182	190	238	260	248	254	166	168	151	153	264	264	190	196	322	326	85	89	131	142	161	163	360	372
Unknown 1 *	Can-1	291 2	93 174	188	190	190 22	5 228	240	260	249	255	209	209	240	254	184	194	238	262	250	254	144	166	151	153	264	264	180	180	318	326	85	89	131	144	153	153	358	368
Unknown 2 *	Com-3	291 2	91 166	188	190	206 22	8 236	238	246	249	255	209	213	246	254	182	194	246	260	270	270	166	166	151	153	264	264	176	190	318	322	83	83	131	135	153	163	375	375
Unknown 3 *	Lau-3, Lau-4, Lau-16	291 3	07 168	188	174	176 22	5 234	236	246	243	255	209	217	240	240	182	186	238	250	254	270	166	166	151	151	264	264	180	192	318	322	83	89	131	131	153	171	364	366
Unknown 4 *	Lau-11	289 2	95 172	184	180	188 22	4 226	230	254	255	255	209	213	238	252	186	190	250	262	250	254	150	164	151	153	264	264	174	190	316	316	85	89	131	131	159	161	364	375
Unknown 5 *	Man-5	291 2	91 166	188	204	204 22	5 234	244	246	249	265	209	221	244	248	180	186	260	260	248	270	166	176	151	153	264	264	184	192	320	328	85	87	142	151	153	161	358	358
Unknown 6 *	Ron-3	291 3	07 184	188	168	174 23	2 234	236	240	249	255	209	209	240	240	182	182	236	250	238	270	166	168	151	153	264	264	186	188	318	326	85	89	131	144	161	177	364	372
Unknown 7 *	Ron-5	291 3	07 184	188	174	206 23	4 234	236	246	255	255	209	209	240	254	182	186	246	250	238	254	168	168	151	153	264	264	188	188	316	318	85	89	131	142	167	177	358	364
Unknown 8 *	Ron-6	291 3	07 184	184	174	188 23	5 238	232	240	249	249	209	211	252	252	182	186	260	262	250	270	166	166	151	153	264	264	176	186	318	322	87	89	131	142	153	163	366	375
CABERNET SAUVIGNON **		291 2	91 184	184	174	178 22	8 238	236	236	249	257	209	217	238	246	176	190	236	238	238	238	166	182	153	153	264	268	188	188	306	314	83	89	137	151	163	163	364	372
SYRAH **		291 2	.95 166	188	174	206 22	4 228	236	236	247	265	209	215	240	240	190	192	220	230	238	270	164	166	151	153	264	264	182	190	318	318	89	89	131	131	163	165	362	382

Table 1. Thirty genotypes obtained for the 49 analyzed accessions at 20 microsatellite loci. Allele sizes are given in base pairs.

<sup>a</sup> Prime names are according to VIVC, except those cultivars marked with an asterisk (\*). Reference cultivars marked with two asterisks (\*\*).

Locus	Na	He	Но	r	PIC	PI
VVIB01	5	0.546	0.700	-0.100	0.5024	0.250
VMC1b11	9	0.709	0.800	-0.053	0.6703	0.123
VMC4F31	9	0.863	0.833	0.016	0.8481	0.034
VVMD5	10	0.861	0.967	-0.057	0.8453	0.035
VVMD7	12	0.793	0.900	-0.060	0.5866	0.069
VVMD21	6	0.691	0.633	0.034	0.6368	0.150
VVMD24	6	0.541	0.600	-0.038	0.5097	0.242
VVMD25	9	0.786	0.733	0.029	0.7852	0.075
VVMD27	6	0.768	0.867	-0.056	0.7676	0.086
VVMD28	10	0.831	0.800	0.017	0.8089	0.050
VVMD32	8	0.790	0.867	-0.043	0.7593	0.075
VVIH54	8	0.607	0.667	-0.037	0.5600	0.201
VVIN16	5	0.585	0.733	-0.094	0.5009	0.256
VVIN73	4	0.185	0.167	0.015	0.1769	0.672
VVIP31	10	0.866	0.867	-0.001	0.8522	0.031
VVIP60	6	0.688	0.600	0.052	0.6369	0.149
VVIQ52	5	0.677	0.833	-0.093	0.6143	0.167
VVS2	9	0.817	0.867	-0.028	0.7938	0.057
VVIV37	11	0.846	0.900	-0.030	0.8132	0.040
VVIV67	11	0.847	0.933	-0.047	0.7798	0.042
TOTAL	159					$1.74 imes10^{-19}$
MEAN	7.95	0.715	0.763	-0.029	0.67241	0.140

Table 2. Characterization of 20 microsatellite markers in the 30 genotypes.

Number of alleles (*Na*), expected heterozygosity (*He*), observed heterozygosity (*Ho*), Frequency of null alleles (r), Polymorphism information content (*PIC*) and Probability of identity (*PI*).

The values obtained from the statistical characterization of the 20 microsatellite loci used in this research study (Table 2) are similar to those obtained in other studies on the genetic characterization of local grapevine cultivars using microsatellite markers [51–53]. Nevertheless, the percentage of new accessions recovered (16.3%) is higher than that obtained by Balda et al. [10] for 45 accessions recovered in Rioja (Spain) (4.4%), Fort et al. for 223 accessions in recovered in Lanzarote (Canary Islands, Spain) (3.6%) [20], and Augusto et al. for 310 accessions recovered in northeast Portugal [32]. This suggests that the grapevine richness of the Andalusian region has not been prospected with the same degree of intensity.

#### 3.2. Cultivar Analysis

Most of the analyzed accessions were identified with known grapevine cultivars. The varietal names were assigned based on the comparison with Spanish [38–43] and European [22,44] microsatellite databases and using the genetic profile of reference varieties for adapting the allele sizes. Allele sizes of genotypes obtained for the twenty SSRs loci analyzed are shown in Tables 1 and 2, and the prime names of the identified cultivars according to VIVC [22], indicating the code of sampled accession for each cultivar (Table 3). Thirty-nine accessions corresponded to 22 known varieties and the ten accessions remaining (Can-1, Comp-3, Lau-3, Lau-4, Lau-16, Lau-11, Man-5, Ron-3, Ron-5 and Ron-6) to the eight unidentified cultivars. These cultivars showed genotypes that did not match any of the published cultivars in the Spanish and European microsatellite databases consulted in this research. Half of the identified accessions are of Spanish origin according to the VIVC database [22] (Table 3), and the country of origin of the rest was France (four accessions), Portugal (three accessions), the United States (one accession), Italy (one accession), Greece (one accession), Algeria (one accession) and Lebanon (one accession). The accessions coded as Lau-3, Lau-4 and Lau-16 showed the same genotype, and they were collected in the same location (Laujar de Andarax, Almería, Spain).

The identified accessions include table and wine grapevine varieties. The table grape varieties, identified by 'Molinera' (Ins-1), 'Imperial Napoleon' (Ins-3) and 'Attika seedless' (Pla-2), have been collected in different regions of the province of Almería (Spain). In this province the cultivation of table grapes was predominant until the 1960s [54]. Furthermore,

one hybrid interspecific ('Jacquez') was identified (Table 2). This hybrid was used for the reconstitution of European vineyards [55]. It is currently prohibited from use in Europe.

**Table 3.** Grapevine material studied with SSR identification, utilization and country of origin of the variety are according to *VIVC* [21].

Accession Code	Cultivars <sup>a</sup>	Utilization <sup>b</sup>	Country of Origin of the Variety <sup>c</sup>						
Alb-1, Chu-3, Lau-12, Lau-13, Lau-10, Man-3	CAYETANA BLANCA	W-T	Spain						
Alb-2	JACQUEZ	W	United States						
Can-1	Unknown 1 *								
Chu-1, Man-1, Rot-1	MANTEUDO	W	Portugal						
Chu-2	BEBA	W-T	Spain						
Chu-4, Man-2	ROAL	W-T	Portugal						
Chu-5	PEDRO XIMENES	W	Spain						
Com-2, Lau-2, Lau-7	JAEN NEGRO	W-T	Spain						
Com-3	Unknown 2 *								
Com-4	ROME TINTO								
Ins-1	MOLINERA	W-T	Spain						
Ins-2, Pla-1, Rot-2	CORNICHON BLANC	W-T	Italy						
Ins-3	IMPERIAL NAPOLEON	Т	Spain						
Ins-4	DONA MARIA	W-T	Portugal						
Lau-1, Lau-14, Lau-15, Lau-18, Lau-19	TINTO VELASCO	W-T	Spain						
Lau-3, Lau-4, Lau-16	Unknown 3 *								
Lau-5, Lau-6	AHMEUR BOU AHMEUR	W-T	Algeria						
Lau-8	ALICANTE HENRI BOUSCHET	W	France						
Lau-9	XARELLO	W	Spain						
Lau-11	Unknown 4 *								
Lau-17	CARIGNAN NOIR	W	France						
Man-4	AFUS ALI	W-T	Lebanon						
Man-5	Unknown 5 *								
Pla-2	ATTIKA SEEDLESS	Т	Greece						
Ron-1	BOBAL	W	Spain						
Ron-2	COJONATA	W	Spain						
Ron-3	Unknown 6 *								
Ron-4	LISTAN PRIETO	W-T	Spain						
Ron-5	Unknown 7 *								
Ron-6	Unknown 8 *								
	CABERNET SAUVIGNON **	W	France						
	SYRAH **	W	France						

<sup>a</sup> Prime names are according to VIVC, except those cultivars marked with an asterisk (\*). Reference cultivars marked with two asterisks (\*\*). <sup>b</sup> Utilization is according to VIVC: W (Wine grape) and T (Table grape). <sup>c</sup> Country of origin of the variety are according to VIVC.

One genotype (Com-4) was identified as 'Rome Tinto', after comparing it with the microsatellite database from Rancho de la Merced [38,39,56]. This variety was only conserved in the Rancho de la Merced Germplasm bank according to the VIVC database and presents a different genotype to the 'Rome' cultivar published by Ibáñez et al. [41].

Four of the varieties identified, 'Beba', 'Jaén negro', 'Pedro Ximenez' and 'Rome Tinto', were already mentioned by Rojas Clemente [6] as being present in the Andalusian region. This shows the antiquity of the cultivation of these varieties in this region. 'Jaén negro' and 'Rome Tinto' are two red grapevine cultivars that have already been identified in old vineyards in the province of Málaga [57].

Currently, most of the cultivars identified in this work have disappeared from the Andalusia vineyards, and the unique cultivar that is growing in the commercial vineyards is 'Pedro Ximenez'. All of this vegetal material recovered from old vineyards could be interesting for the wine industry in Andalusia or regions with similar agroclimatic conditions. However, many of these identified cultivars are not included in the official register of Spanish grapevine varieties for the community of Andalusia, which would make their cultivation difficult. Recently, 'Beba' has been included as an authorized variety in the regulation of wines of the Protected Designation of Origin "Jerez-Xérès-Sherry" (Spain) [58], as there is some interest in increasing the diversity of wines [59].

In addition, of the identified varieties, the eight new genotypes should be studied and evaluated in order to make their oenological potential and adaptation climate change known among the wine sector. Furthermore, these cultivars could be important genetic resources for future breeding programs.

### 3.3. Genetic Relationships among Cultivars

Based on the results of the analysis of the microsatellites, the distance matrix was used to carry out a grouping using UPGMA. To characterize the genetic structure of different genotypes obtained and two references varieties ('Cabernet Sauvignon' and 'Syrah'), a dendrogram based on the proportion of shared alleles was constructed. Figure 1 shows the resulting dendrogram of the 30 non-redundant genotypes found in this study.



Figure 1. Genetic relationships among the thirty genotypes obtained and two reference cultivars.

SSR analysis allowed for the evaluation of the genetic relationships among European cultivars and unknown accessions recollected in different regions of Andalusia. The dendrogram in Figure 1 shows the existence of two defined groups. Group I includes only one cultivar identified with 'Jacquez', which is a hybrid interspecific of the cross between *Vitis aestivalis* × *Vitis vinifera* [22]. All of the rest of the identified and unknown cultivars are included in group II and are cultivars of the *Vitis vinifera* species. The formation of these two groups may be related to the pedigree of the cultivars. In group II, there is no clear separation of different subgroups in relation to regions of origin as found in other published research papers on Sicilian varieties [60]. Varieties with different countries of origin are grouped in this cluster II (Table 3).

Two varieties, 'Cabernet Sauvignon' and 'Atikka seedless' are markedly distant from the rest of the cultivars in Group II, probably because of a different origin and use. 'Atikka seedless' is considered a seedless variety of Greek origin according to VIVC [22].

Phylogenetic distances of the subgroup where variety "Unknown 6" is included indicates that it could be a wine and table grape, since it is grouped with other grapes that are used as wine and table grapes, such as 'Cayetana Blanca', 'Molinera' and 'Jaén Tinto' (Table 3). The same behavior could be said for the variety "Unknown 7" and the 'Roal', "Unknown 4" and 'Tinto Velasco' or "Unknown 5" and 'Cornichon Blanc'.

## 4. Conclusions

Forty-nine accessions collected in Andalusia have been described by molecular methods. A total of 83.7% of these accessions analyzed have been identified by comparison to Spanish or European microsatellite databases with known cultivars. However, eight genotypes have not yet been identified and could represent old local cultivars in danger of extinction. All of these genotypes have been preserved within the Rancho de la Merced germplasm bank (Andalusia, Spain).

This study indicates an important biodiversity within the old vineyards from the Andalusia region that provides interesting information for the wine industry and that points out the wide genetic diversity of grapevines which are still unexploited. Our efforts should lead to the protection and study of local grape natural richness.

**Supplementary Materials:** The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/horticulturae9030316/s1, Table S1: List of the 49 accessions collected and their place of origin in the Andalusia region (Spain).

Author Contributions: Conceptualization, A.J.-C., A.P.-P. and R.A.-G.; methodology, A.J.-C., A.P.-P. and R.A.-G.; software, A.J.-C. and R.A.-G.; validation, A.J.-C. and R.A.-G.; formal analysis, A.J.-C.; investigation, A.J.-C., A.P.-P. and R.A.-G.; resources, A.J.-C.; data curation, A.J.-C.; writing—original draft preparation, A.J.-C., A.P.-P. and R.A.-G.; writing—review and editing, A.J.-C., A.P.-P. and R.A.-G.; visualization, A.J.-C., A.P.-P. and R.A.-G.; supervision, A.J.-C., A.P.-P. and R.A.-G.; have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Ministerio de Ciencia y Tecnología (INIA-Spain), grants number RF2004-00014-00-00, VIN00-036-C6-5X, RF2006-00011-00-00 and RF2007-00017-00-00.

**Acknowledgments:** The authors would like to thank the IFAPA for facilitating the material conserved in the Rancho de la Merced germplasm bank for the study.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- Pozo-Bayón, M.Á.; Moreno-Arribas, M.V. Sherry wines. In Advances in Food and Nutrition Research; Jackson, R.S., Ed.; Academic Press: Cambridge, MA, USA, 2011; Volume 63, pp. 17–40. [CrossRef]
- Prados Martínez, F. La producción vinícola en el mundo fenicio-púnico. Apuntes sobre cultivo de la vid y consumo del vino a través de las fuentes arqueológicas y literarias. *Gerión* 2011, 29, 9–35. [CrossRef]
- 3. González Rodríguez, R. Excavaciones de Urgencia en el Cerro Naranja (Jerez de la Frontera, Cádiz). In *Anuario Arqueológico de Andalucía, III;* Junta de Andalucía: Sevilla, Spain, 1985; pp. 90–96.

- Torres, J.R. Las Ánforas Fenicio-Púnicas del Mediterráneo Central y Occidental. Ph.D. Thesis, University of Barcelona, Barcelona, Spain, 1991.
- Aubet Semmler, M.E.; Buxó Capdevila, R. Los recursos y la economía colonial. En Aubet Semmler, M.E.; et al. ii Cerro del Villar I. El Asentamiento fenicio en la desembocadura del río Guadalhorce y su interacción con el hinterland. *Arqueol. Monogr.* 1999, 5, 334–339.
- Vera, J.C.; Echevarría, A. Sistemas agrícolas del I milenio a.C. en el yacimiento de La Orden-Seminario de Huelva: Viticultura protohistórica a partir del análisis arqueológico de las huellas de cultivo. In Proceedings of the Patrimonio Cultura de la Vid y el Vino, Madrid, Spain, 2013; pp. 95–106.
- De Rojas Clemente y Rubio, S. Ensayo Sobre las Variedades de vid Común que Vegetan en Andalucía; Imprenta Villalpando: Madrid, Spain, 1807; p. 324.
- Morilla Critz, J. La viticulture de Andalucía en 1831 vista por James Busby, padre de la viticultura Australiana. *Estud. Reg.* 1997, 49, 261–298.
- 9. Fernández de Bobadilla, G. *Viníferas Jerezanas y de Andalucía Occidental;* Instituto Nacional de Investigaciones Agronómicas: Madrid, Spain, 1956.
- 10. Balda, P.; Ibáñez, J.; Sancha, J.C.; Martínez de Toda, F. Characterization and identification of minority red grape varieties recovered in Rioja, Spain. *Am. J. Enol. Vitic.* **2014**, *65*, 148–152. [CrossRef]
- 11. Fatahi, R.; Ebadi, A.; Bassil, N.; Mehlenbacher, S.A.; Zamani, Z. Characterization of Iranian grapevine cultivars using microsatellite markers. *Vitis* **2003**, *42*, 185–192.
- 12. Martínez, L.E.; Cavagnaro, P.F.; Masuelli, R.W.; Zuñiga., M. SSR-based assessment of genetic diversity in South American *Vitis* vinifera varieties. *Plant Sci.* 2006, 170, 1036–1044. [CrossRef]
- 13. De Mattia, F.; Imazio, S.; Grassi, F.; Lovicu, G.; Tardaguila, J.; Failla, O.; Maitt, C.; Scienza, A.; Labra, M. Genetic characterization of Sardinia grapevine cultivars by SSR markers analysis. *J. Int. Sci. Vigne et du Vin.* **2007**, *41*, 175–184. [CrossRef]
- 14. Lacombe, T.; Boursiquot, J.M.; Laucou, V.; Dechesne, F.; Varès, D.; This, P. Relationships and genetic diversity within the accessions related to Malvasia held in the Domaine de Vassal grape germplasm repository. *Am. J. Enol. Vitic.* 2007, *58*, 124–131. [CrossRef]
- Dzhambazova, T.; Tsvetkov, I.; Atanassov, I.; Rusanos, K.; Martínez Zapater, J.M.; Atanassov, A.; Hvarleva, T. Genetic diversity in native Bulgarian grapevine germplasm (*Vitis vinifera* L.) based on nuclear and chloroplast microsatellite polymorphisms. *Vitis* 2009, 48, 115–121.
- 16. De Olivera, G.L.; De Souza, A.P.; De Oliver, F.A.; Zucchi, M.I.; De Souza, L.M.; Moura, M.F. Genetic structure and molecular diversity of Brazilian grapevine germplasm: Management and use in breeding programs. *PLoS ONE* **2020**, *15*, e0240665. [CrossRef]
- 17. Upadhayay, A.; Aher, L.B.; Shinde, M.P.; Mundankar, K.Y.; Datre, A.; Karibasappa, G.S. Microsatellite analysis to rationalize grape germplasm in India and development of a molecular database. *Plant Genet. Resour.* **2013**, *11*, 225–233. [CrossRef]
- Popescu, C.F.; Maul, E.; Dejeu, L.C.; Dinu, D.; Gheorge, R.N.; Laucou, V.; Lacombe, T.; Migliaro, D.; Crespan, M. Identification and characterization of Romanian grapevine genetic resources. *Vitis* 2017, 56, 173–180.
- 19. Marsal, G.; Méndez, J.J.; Mateo, J.M.; Ferrer, S.; Canals, J.M.; Zamora, F.; Fort, F. Molecular characterization of *Vitis vinifera* L. local cultivars from volcanic areas (Canary Islands and Madeira) using SSR markers. *OENO One* **2019**, *53*, 667–680. [CrossRef]
- Fort, F.; Marsal, G.; Mateo-Sanz, J.M.; Pena, V.; Canals, J.M.; Zamora, F. Molecular characterisation of the current cultivars of *Vitis vinifera* L. in Lanzarote (Canary Islands, Spain) reveals nine individuals which correspond to eight new varieties and two new sports. *OENO One* 2022, 56, 281–295. [CrossRef]
- García de Luján, A.; Lara Benítez, M. La colección de vides del Rancho de la Merced; Junta de Andalucía, Consejería de Agricultura y Pesca: Sevilla, Spain, 1997; p. 157.
- 22. Vitis International Variety Catalogue. Available online: www.vivc.de (accessed on 26 December 2022).
- Pastor, I.; Rovira, J.M. La Investigación arqueológica en los procesos de recuperación de variedades de uva autóctonas tradicionales. In Proceedings of the Patrimonio Cultura de la Vid y el Vino, Madrid, Spain, 2013; pp. 71–79.
- 24. Sancho-Galán, P.; Amores-Arrocha, A.; Palacios, V.; Jiménez-Cantizano, A. Identification and characterization of white grape varieties autochthonous of a warm climate region (Andalusia, Spain). *Agronomy* **2020**, *10*, 205. [CrossRef]
- Boursiquot, J.M.; This, P. Les nouvelles techniques utilisées en ampélographie: Informatique et marquage. J. Int. Sci. Vigne Vin 1996, 610, 13–23.
- Emanuelli, F.; Lorenzi, S.; Grzeskowiak, L.; Catalano, V.; Stefanini, M.; Troggio, M.; Myles, S.; Martínez-Zapater, J.M.; Zyprian, E.; Moreira, F.M.; et al. Genetic diversity and population structure assessed by SSR and SNP markers in large germplasm collection of grape. *BMC Plant Biol.* 2013, *13*, 39. [CrossRef]
- Sefc, K.M.; Lefort, F.; Grando, M.S.; Scott, K.D.; Steinkellner, H.; Thomas, M.R. Microsatellite markers for grapevine: A state of the art. In *Molecular Biology and Biotechnology of Grapevine*; Roubelakis-Angelakis, K.A., Ed.; Kluwer Academic: Dordrecht, The Netherlands, 2001; pp. 433–463.
- This, P.; Jung, A.; Boccacci, P.; Borrego, J.; Botta, R.; Costantini, L.; Crespan, M.; Dangl, G.S.; Eisenheld, C.; Ferreira-Monteiro, F.; et al. Development of a standard set of microsatellite reference alleles for identification of grape cultivars. *Theor. Appl. Genet.* 2004, *109*, 1448–1458. [CrossRef]
- González-Andrés, F.; Martín, J.P.; Yuste, J.; Rubio, J.A.; Arranz, C.; Ortiz, J.M. Identification and molecular biodiversity of autochthonous grapevine cultivars in the "Comarca del Bierzo", León, Spain. Vitis 2007, 46, 71–76.

- Jiménez-Cantizano, A.; Lara, M.; Ocete, M.E.; Ocete, R. Short communication: Characterization of the relic Almuñécar grapevine cultivar. Span. J. Agric. Res. 2012, 10, 454–460. [CrossRef]
- 31. Mena, A.; Martínez, M.; Fernández-González, M. Recovery, identification and relationships by microsatellite analysis of ancient grapevine cultivars from Castilla-La Mancha: The largest wine growing region in the world. *Genet. Resour. Crop Evol.* **2014**, *61*, 625–637. [CrossRef]
- Augusto, D.; Ibáñez, J.; Pinto-Sintra, A.L.; Falco, V.; Leal, F.; Martínez-Zapater, J.M.; Oliveira, A.A.; Castro, I. Grapevine Diversity and Genetic Relationships in Northeast Portugal Old Vineyards. *Plants* 2021, 10, 2755. [CrossRef] [PubMed]
- 33. Bowers, J.E.; Dangl, G.S.; Vignani, R.; Meredith, C.P. Isolation and characterization of new polymorphic simple sequence repeat loci in grape (*Vitis vinifera* L.). *Genome* **1996**, *39*, 628–633. [CrossRef] [PubMed]
- 34. Bowers, J.E.; Dangl, G.S.; Meredith, C.P. Development and characterization of additional microsatellite markers for grape. *Am. J. Enol. Vitic.* **1999**, *50*, 243–246. [CrossRef]
- 35. Thomas, M.R.; Scott, N.S. Microsatellite repeats in grapevine reveal DNA polymorphisms when analyzed as sequence-tagged sites (STSs). *Theor. Appl. Genet.* **1993**, *86*, 985–990. [CrossRef] [PubMed]
- Merdinoglu, D.; Butterlin, G.; Bevilacqua, L.; Chiquet, V.; Adam-Blondon, A.F.; Decroocq, S. Development and characterization of a large set of microsatellite markers in grapevine (*Vitis vinifera* L.) suitable for multiplex PCR. *Mol. Breed.* 2005, 15, 349–366. [CrossRef]
- Vargas, A.M.; Velez, M.D.; De Andrés, M.T.; Laucou, V.; Lacombe, T.; Boursiquot, J.M.; Borrego, J.; Ibáñez, J. Corinto bianco: A seedless mutant of Pedro Ximenes. Am. J. Enol. Vitic. 2007, 58, 540–543. [CrossRef]
- Jiménez-Cantizano, A. Caracterización Molecular del Banco de Germoplasma de vid del Rancho de la Merced. Ph.D. Thesis, University of Cádiz, Cádiz, Spain, 2014. Available online: http://hdl.handle.net/10498/17919 (accessed on 12 November 2022).
- Jiménez-Cantizano, A.; García De Luján, A.; Arroyo-García, R. Molecular characterization of table grape varieties preserved in the Rancho de la Merced Grapevine Germplasm Bank (Spain). Vitis 2018, 57, 93–101.
- Cretazzo, E.; Moreno Sanz, P.; Lorenzi, S.; Benítez, M.L.; Velasco, L.; Emanuelli, F. Genetic Characterization by SSR Markers of a Comprehensive Wine Grape Collection Conserved at Rancho de la Merced (Andalusia, Spain). *Plants* 2022, 11, 1088. [CrossRef] [PubMed]
- 41. Ibáñez, J.; Vargas, A.M.; Palancar, M.; Borrego, J.; De Andrés, M.T. Genetic Relationships among Table-Grape Varieties. *Am. J. Enol. Vitic.* **2009**, *60*, 35–42. [CrossRef]
- Vargas, A.M.; De Andrés, M.T.; Borrego, J.; Ibáñez, J. Pedigrees of fifty tables-grape cultivars. Am. J. Enol. Vitic. 2009, 60, 525–531. [CrossRef]
- De Andrés, M.T.; Benito, A.; Pérez-Rivera, G.; Ocete, R.; López, M.A.; Gaforio, L.; Muñoz, G.; Cabello, F.; Martínez-Zapater, J.M.; Arroyo-García, R. Genetic diversity of wild grapevine populations in Spain and their genetic relationships with cultivated grapevines. *Mol. Ecol.* 2012, *21*, 800–816. [CrossRef]
- 44. Lacombe, T.; Boursiquot, J.M.; Laucou, V.; Di Vecchi-Staraz, M.; Péros, J.P.; This, P. Large-scale parentage analysis in an extended set of grapevine cultivars (*Vitis vinifera* L.). *Theo. Appl. Genet.* **2013**, *126*, 401–414. [CrossRef]
- Park, S.D.E. Trypanotolerance in West African Cattle and the Population Genetic Effects of Selection. Ph. D. Thesis, University of Dublin, Dublin, Ireland, 2001.
- 46. Peakall, R.; Smouse, P. GENALEX 6: Genetic analysis in Excel. Population genetic software for teaching and research. *Mol. Ecol. Res.* **2006**, *6*, 288–295. [CrossRef]
- 47. Bi 'nkowski, J.; Miks, S. Gene-Calc. 2018. Available online: www.gene-calc.pl. (accessed on 20 February 2023).
- 48. Minch, E.; Ruiz-Linares, A.; Goldstein, D.; Feldman, M.; Cavalli-Sforza, M. *Microsat 1.5d: A Computer Program for Calculating Various Statistics on Microsatellite Allele Date;* Washington State University: Pullman, WA, USA, 1997.
- 49. Felsenstein, J. Phylogeny inference package. *Cladistics* **1989**, *5*, 164–166.
- 50. Kumar, S.; Stecher, G.; Tamura, K. MEGA 7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol. Biol. Evol.* **2015**, *33*, 1870–1874. [CrossRef]
- Fenández-González, M.; Mena, A.; Izquierdo, P.; Martínez, J. Genetic characterization of grapevine (*Vitis vinífera* L.) cultivars from Castilla La Mancha (Spain) using microsatellite markers. *Vitis* 2007, 46, 126–130.
- 52. Žulj Mihaljević, M.; Šimon, S.; Pejić, I.; Carka, F.; Sevo, R.; Kojić, A.; Gaši, F.; Tomić, L.; Jovanović Cvetković, T.; Maletić, E.; et al. Molecular characterization of old local grapevine varieties from South East European countries. *Vitis* **2013**, *52*, 69–76.
- 53. Basheer-Salimia, R.; Lorenzi, S.; Batarseh, F.; Moreno-Sanz, P.; Emanuelli, F.; Grando, M.S. Molecular identification and genetic relationships of Palestinian grapevine cultivars. *Mol. Biotechnol.* **2014**, *56*, 546–556. [CrossRef]
- Gómez-Espín, J.M.; Gil Meseguer, E. La exportación española de uva de mesa en la segunda mitad del siglo XX. Papeles Geografía 1987, 13, 87–104.
- Santiago, J.L.; González, I.; Gago, P.; Alonso-Villaverde, V.; Boso, S.; Martínez, M.C. Identification of and relationships among a number of teinturier grapevines that expanded across Europe in the early 20th century. *Aust. J. Grape and Wine Res.* 2008, 14, 223–229. [CrossRef]
- 56. Jiménez-Cantizano, A.; Amores-Arrocha, A.; Gutiérrez-Escobar, R.; Palacios, V. Identification and relationship of the autochthonous 'Romé' and 'Rome Tinto' grapevine cultivars. *Span. J. Agric. Res.* **2018**, *16*, e07SC02. [CrossRef]

- 57. Jiménez-Cantizano, A.; Muñoz-Martín, A.; Amores-Arrocha, A.; Sancho-Galán, P.; Palacios, V. Identification of Red Grapevine Cultivars (*Vitis vinifera* L.) Preserved in Ancient Vineyards in Axarquía (Andalusia, Spain). *Plants* 2020, 9, 1572. [CrossRef] [PubMed]
- 58. Orden de 4 de Octubre de 2022, Boletín Oficial de la Junta de Andalucía (BOJA). Available online: https://www.juntadeandalucia. es/eboja/2022/195/s54.html (accessed on 3 January 2023).
- 59. Sancho-Galán, P.; Amores-Arrocha, A.; Palacios, V.; Jiménez-Cantizano, A. Genetical, Morphological and Physicochemical Characterization of the Autochthonous Cultivar 'Uva Rey' (*Vitis vinifera* L.). *Agronomy* **2019**, *9*, 563. [CrossRef]
- 60. Carimi, F.; Mercati, F.; Abbate, L.; Sunseri, F. Microsatellite analyses for evaluation of genetic diversity among Sicilian grapevine cultivars. *Genet. Resour. Crop Evol.* **2010**, *57*, 703–719. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.