

Estimation of carcass composition and cut composition from computed tomography images of live growing pigs of different genotypes

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The aim of the present work was (1) to study the relationship between cross-sectional computed tomography (CT) images obtained in live growing pigs of different genotypes and dissection measurements and (2) to estimate carcass composition and cut composition from CT measurements. Sixty gilts from three genotypes (Duroc × (Landrace × Large White), Pietrain × (Landrace × Large White), and Landrace × Large White) were CT scanned and slaughtered at 30 kg (n = 15), 70 kg (n = 15), 100 kg (n = 12) or 120 kg (n = 18). Carcasses were cut and the four main cuts were dissected. The distribution of density volumes on the Hounsfield scale (HU) were obtained from CT images and classified into fat (HU between –149 and –1), muscle (HU between 0 and 140) or bone (HU between 141 and 1400). Moreover, physical measurements were obtained on an image of the loin and an image of the ham. Four different regression approaches were studied to predict carcass and cut composition: linear regression, quadratic regression and allometric equations using volumes as predictors, and linear regression using volumes and physical measurements as predictors. Results show that measurements from whole animal taken in vivo with CT allow accurate estimation of carcass and cut composition. The prediction accuracy varied across genotypes, BW and variable to be predicted. In general, linear models, allometric models and linear models, which included also physical measurements at the loin and the ham, produced the lowest prediction errors.

Keywords: computed tomography, live growing pig, carcass composition, cut composition, prediction equations

Implications

Pig carcass composition and cut composition are determinant parameters in the optimization of the production chain. The knowledge of these characteristics with the minimum error in live pigs during growth would be useful in breeding and nutritional programmes to improve the overall economic performance of pig carcasses and to ensure an optimized production to obtain the desired product according to producers and industry demands. This paper shows the feasibility of computed tomography for this purpose and the errors of prediction obtained according to different statistical approaches.

Introduction

Pig growth and body composition are essential components of pig profitability. Weight and composition of gain change in

pigs during the growing and finishing periods is determined mainly by the genetic potential and supply of nutrients (Kolstad *et al.*, 1996; Kouba *et al.*, 1999; Lambe *et al.*, 2013). Knowledge of changes in tissue composition at whole-carcass and primal-cut levels would allow the optimization of slaughter, satisfy niche market demands, and improve the economic efficiency of pork production systems. This information is required as the economic value of pig carcasses is related to the composition of primal cuts rather than the composition of the whole carcass (Marcoux *et al.*, 2007).

Serial slaughtering has traditionally been used to study the body composition of pigs during growth (Fisher *et al.*, 2003; Landgraf *et al.*, 2006). However, another method used X-ray computed tomography (CT), a non-destructive and non-invasive technique that measures the density of the tissues and allows the body composition of animals to be estimated easily (Kolstad and Vangen, 1996; Barchia *et al.*, 2010; Lambe *et al.*, 2013); thus, serial slaughtering may no longer be required. Nevertheless, serial slaughtering is still needed

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to establish the relationship between CT data (i.e. total lean volume) and cutting (joints separation) and dissection (separation of the different tissues of the joints) data and to obtain prediction equations for the weight and composition of main cuts. Several studies have demonstrated that CT is an excellent tool for estimating carcass tissue composition in either pig carcasses (Judas *et al.*, 2006; Font i Furnols *et al.*, 2009; Picouet *et al.*, 2010) or live animals (Kolstad *et al.*, 1996; Kolstad, 2001). However, to the best of our knowledge, the estimation of cut composition from the CT scanning of live animal has not yet been reported.

The aim of the present work was (1) to study the relationship between cross-sectional CT images obtained in live growing female pigs of different genotypes and dissection measurements and (2) to estimate carcass composition and cut composition from CT measurements.

Material and methods

Animals and scanning procedure

The study used a total of 60 female pigs from three different genotypes, namely Duroc × (Landrace × Large White), Pietrain × (Landrace × Large White), and Landrace × Large White, respectively, referred to in this paper as DU × (LD × LW), PI × (LD × LW) and LD × LW. Within each genotype, the pigs were assigned randomly to a target live weight of 30 kg

($n = 15$, five per genotype), 70 kg ($n = 15$, five per genotype), 100 kg ($n = 12$, four per genotype) or 120 kg ($n = 18$, six per genotype). The animals were reared individually on the IRTA experimental farm in Monells, Spain. Feeds were provided *ad libitum* according to a two-phase feeding programme and contained 10.24 and 10.08 MJ net energy/kg, 18.00% and 17.02% crude protein and 0.91% and 0.90% digestible lysine fed basis during the first and second phases, respectively. The feeds were formulated to satisfy or exceed the *a priori* estimated animal requirements using commercial standards. The second feeding phase started at ~25 kg BW.

When the pigs reached their assigned target weight, they were fasted for a minimum of 8 h and then transported to the IRTA New Technologies Centre (Monells, Spain), where they were anaesthetized (by intramuscular injection of azaperone at 0.1 mg/kg, ketamine at 0.2 mg/kg, and if necessary, propofol at 0.22 mg/kg), placed in a PVC cradle, and scanned using a CT device (HiSpeed Zx/i; GE Healthcare, Madrid, Spain). Scanning was done following the protocol used in carcass evaluation (Font i Furnols *et al.*, 2009) with some modifications: axial acquisition, 140 kV and 145 mA, 512 × 512 matrix, 7-mm thickness (at the 30 kg target weight) or 10-mm thickness (at the 70, 100 and 120 kg target weights) and displayed field of view (DFOV) between 300 and 460 mm, adapted to the size of the pig. The experiment was approved by the IRTA Ethics Committee.

Table 1 Mean and standard deviation (s.d.) of weights and computed tomography tissue volumes^a obtained in live pigs of three genotypes at different live weight

	30 kg ($n = 15$)		70 kg ($n = 15$)		100 kg ($n = 12$)		120 kg ($n = 18$)	
	Mean	s.d.	Mean	s.d.	Mean	s.d.	Mean	s.d.
LD × LW								
Live wgt. (kg)	30.0	2.2	71.3	2.5	98.5	1.8	122.0	1.8
VolLean (dm ³)	18.5	1.3	41.1	2.1	50.3	5.5	60.1	4.1
VolFat (dm ³)	6.0	0.7	20.0	1.9	35.3	8.1	45.7	5.5
VolBone (dm ³)	2.2	0.1	4.4	0.2	5.7	0.3	6.7	0.4
TotalVol (dm ³)	28.7	1.9	69.5	3.1	96.2	2.2	117.2	4.3
PLean (%)	64.5	1.1	59.2	2.5	52.4	6.6	51.4	4.0
PI × (LD × LW)								
Live wgt. (kg)	31.4	2.2	67.7	1.8	100.5	1.5	123.1	3.8
VolLean (dm ³)	19.2	1.4	43.1	1.3	57.2	1.5	66.6	5.1
VolFat (dm ³)	6.1	0.6	14.3	2.2	30.0	3.1	39.5	4.0
VolBone (dm ³)	2.1	0.1	4.27	0.1	5.55	0.3	6.6	0.4
TotalVol (dm ³)	29.2	2.1	65.1	2.5	96.8	1.5	117.2	2.0
PLean (%)	65.7	1.5	66.2	2.3	59.1	1.9	56.8	3.5
DU × (LD × LW)								
Live wgt. (kg)	29.3	2.3	68.8	2.2	100.8	3.2	123.8	3.4
VolLean (dm ³)	18.3	0.6	41.0	3.6	53.3	2.9	63.3	2.4
VolFat (dm ³)	5.4	1.3	17.4	2.9	33.0	2.5	43.4	3.8
VolBone (dm ³)	2.2	0.1	4.6	0.2	6.2	0.2	7.2	0.3
TotalVol (dm ³)	27.8	2.1	66.4	2.1	96.6	2.2	118.7	2.2
PLean (%)	66.1	2.8	61.7	4.3	55.1	2.6	53.4	2.5

LD: Landrace; LW: Large White; PI: Pietrain; DU: Duroc.

^aVolLean: volume between 0 Hounsfield units (HU) and 140 HU; VolFat: volume between -149 and -1 HU; VolBone: volume between 141 and 1400 HU; TotalVol: volume between -1000 and 1400 HU; PLean: $100 \times \text{VolLean} / \text{TotalVol}$.

Slaughter and dissection

After scanning, the pigs were transported to the experimental abattoir and slaughtered following standard procedures (after having been previously stunning with CO₂ when animals had recovered consciousness from scanning). The carcasses were kept refrigerated at 2°C until they were processed. The left side of each carcass was prepared and cut following the European Union reference method (Walstra and Merkus, 1995) between 24 and 48 h after slaughter. Thereafter, four primal cuts (ham, shoulder, belly and loin) plus tenderloin were weighed and manually dissected. Lean, subcutaneous fat including the skin, intermuscular fat and bone were separated with a knife by trained technicians, and the weights of all these tissues were recorded. All tenderloin

weight was considered as lean. Descriptive statistics of these variables are provided as Supplementary Table S1. Carcass lean meat percentage was calculated by dividing the overall amount of dissected meat from each primal cut plus tenderloin by the total weight of these five cuts. A factor of 0.89 was applied in accordance with Commission Regulation (EC) No. 1249/2008 to obtain the lean meat percentage of the whole carcass from the four main cuts.

Image processing

The distribution of density volumes based on the Hounsfield scale (in Hounsfield units (HU)) was obtained from CT images using the VisualPork software package, which was developed for that purpose by the University of Girona and the

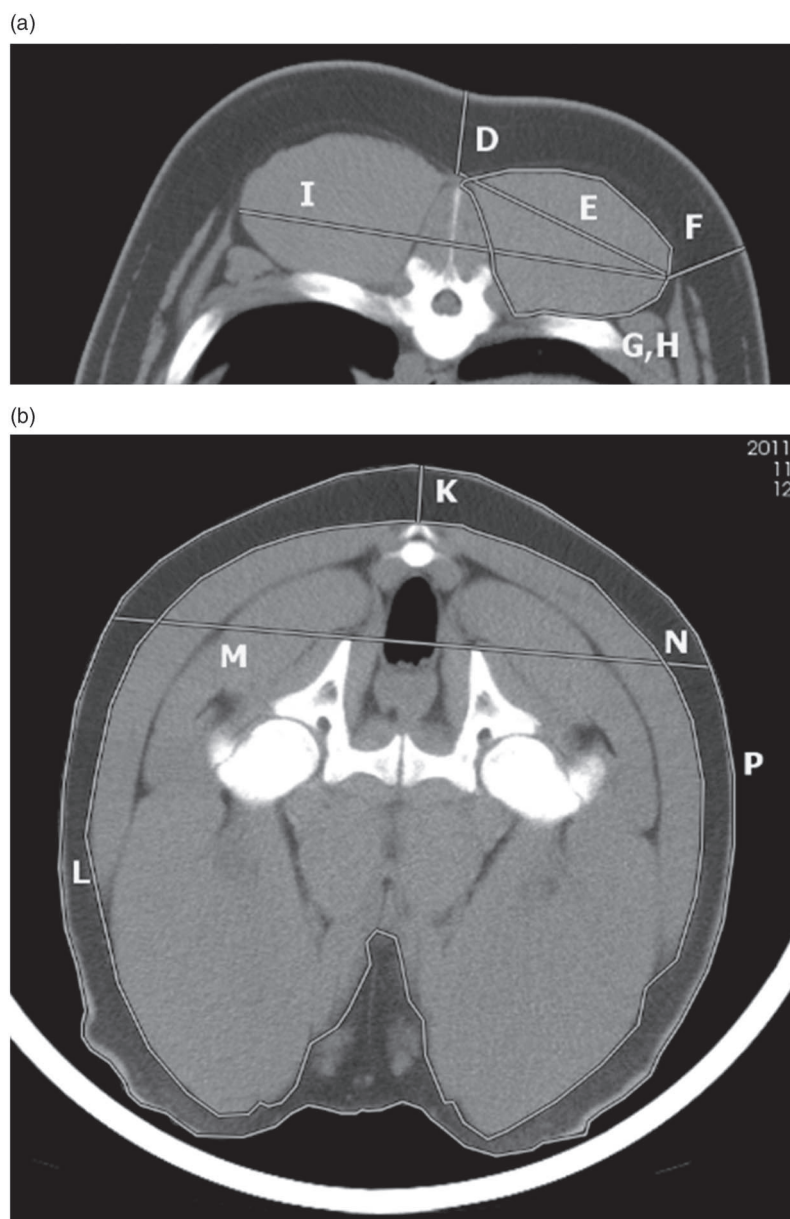


Figure 1 Physical direct measurements taken at the loin (a) and ham (b). (D: superior subcutaneous fat thickness; E: diagonal of the *longissimus thoracis* muscle; F: lateral subcutaneous fat thickness; G: loin area; H: loin perimeter; I: maximum loin width; K: subcutaneous fat thickness at the centre; L: area of the subcutaneous fat; M: width of the ham; N: lateral subcutaneous fat thickness; P: perimeter of the whole ham).

IRTA (Boada *et al.*, 2009; Bardera *et al.*, 2012). The cradle was removed from all the images, but the viscera was left. The frequencies of voxels between -1000 and $+1400$ HU were converted into volumes by means of the DFOV value, the matrix size and the image thickness value, as follows: $\text{volume} = \text{number of voxels} \times \text{thickness} \times (\text{DFOV}/512)^2$. Hounsfield volume distributions were studied further to determine the limits for fat, muscle and bone tissues. From the volume distribution averaged by the target weight in the border region between muscle and bone, the change in slope after the high decrease in the lean area was selected as a cut-off and was set at an HU value of 140 (Supplementary Figure S1). The HU value of 0 was selected as a separation between

muscle and fat. Finally, because viscera were included in the images, a cut-off was necessary to separate fat from tissues with less density, such as the lungs or parts of the intestines. In this case, a change in the curve was found at -149 HU because of the inner methodology of the programme for determining the contour of the body, and this value was used as the cut-off. Thus, the partial volumes estimated between -149 and -1 , between 0 and 140 and between 141 and 1400 were associated with fat, muscle and bone volumes, respectively, and were used afterwards as independent variables in the regression analysis. Volumes between -1000 and -150 HU, which belong mainly to the less dense parts of the viscera, were considered only in the total volume.

Table 2 Descriptive statistics (mean and standard deviation (s.d.)) of direct physical measurements (in mm or mm²) taken from whole-animal CT images^a

	30 kg (n = 15)		70 kg (n = 15)		100 kg (n = 12)		120 kg (n = 18)	
	Mean	s.d.	Mean	s.d.	Mean	s.d.	Mean	s.d.
LD × LW								
Loin area	1307	58	3348	337	3932	481	4613	434
Loin perimeter	157	7	225	4	248	11	259	13
Loin width	140	3	181	2	195	8	209	10
Loin sup. subc. fat	7	1	16	1	30	3	34	5
Loin lat. subc. fat	7	1	13	3	22	8	24	4
Loin diagonal	75	2	98	1	102	6	110	5
Ham perimeter	255	10	245	6	275	11	305	10
Ham sup. subc. fat	5	1	8	3	17	4	24	6
Ham lat. subc. fat	5	1	11	2	18	3	27	5
Ham perimeter	750	26	925	32	1097	22	1226	55
Ham fat area	4037	771	9788	791	14539	4104	20993	3451
PI × (LD × LW)								
Loin area	1526	146	3669	375	5090	353	5158	693
Loin perimeter	161	9	234	10	270	8	266	17
Loin width	132	5	183	4	209	5	207	10
Loin sup. subc. fat	10	1	16	3	29	6	30	4
Loin lat. subc. fat	6	1	11	2	19	4	25	6
Loin diagonal	70	3	99	3	107	2	105	7
Ham perimeter	253	13	258	12	294	5	311	9
Ham sup. subc. fat	6	2	8	3	16	4	20	4
Ham lat. subc. fat	6	1	9	1	17	2	19	3
Ham perimeter	784	46	1025	39	1191	26	1258	51
Ham fat area	3709	767	8462	1141	12024	1643	16144	2646
DU × (LD × LW)								
Loin area	1527	86	3119	406	5026	273	5057	464
Loin perimeter	157	4	220	9	267	7	269	11
Loin width	132	1	175	8	208	6	214	7
Loin sup. subc. fat	9	1	16	2	29	3	35	4
Loin lat. subc. fat	6	2	13	3	18	3	25	6
Loin diagonal	71	2	92	5	108	3	109	5
Ham perimeter	252	9	242	6	282	13	299	10
Ham sup. subc. fat	6	2	9	1	14	1	23	5
Ham lat. subc. fat	7	2	10	2	18	2	20	3
Ham perimeter	742	73	992	18	1151	81	1217	75
Ham fat area	3735	372	8396	715	13286	1838	17334	1886

LD = Landrace; LW = Large White; PI = Pietrain; DU = Duroc; sup. = superior; subc. = subcutaneous; lat. = lateral.

^aDirect physical measurements used as dependent variables (predictors) in the prediction equations.

The relative carcass lean meat volume (PLean) was calculated as the ratio between the carcass lean meat volume and total carcass volume. Descriptive statistics of the volumes by slaughter weight and genotype are presented in Table 1.

Some physical measurements have been obtained to see whether their inclusion as predictors could reduce prediction error. From an image of the loin between the third and fourth last ribs (Figure 1a – without considering the fals rib), the following six direct physical measurements were obtained: (i) maximum loin width (*I*); (ii) loin area (*G*); (iii) loin perimeter (*H*); (iv) diagonal (maximum length) of the *longissimus thoracis* muscle (*E*); (v) lateral subcutaneous fat thickness at the lateral extreme of the *longissimus thoracis* muscle and perpendicular to the skin (*F*); and (vi) superior subcutaneous fat thickness at the centre of the dorsal part of the body and perpendicular to the skin (*D*). From an image of the ham (at the junction of the femur and pubis bones – Figure 1b), the following five direct physical measurements were obtained: (i) width of the ham above the pubis bone (*M*); (ii) area of the subcutaneous fat of the whole image (*L*); (iii) lateral subcutaneous fat thickness at the same level as the width measurement (*N*); (iv) subcutaneous fat thickness

at the centre of the dorsal part of the body and perpendicular to the skin (*K*); and (v) perimeter of the whole image of the ham (*P*). All lengths and areas were measured in millimetres and square millimetres, respectively. Descriptive statistics of these loin and ham direct physical measurements are presented in Table 2.

Statistical analysis

The relationships between dependent variables (carcass and cuts composition) and independent variables (the CT-measured volume, ratio of volumes or physical measurements) were studied within genotypes, as preliminary analyses showed large differences between the studied genetic lines.

Four different regression approaches, all performed with the REG procedure of the SAS software package (version 9.2; SAS Institute Inc., Cary, NC, USA), were studied. Because of the heteroscedasticity of variances related to the differences in slaughter weight, all the analyses were performed by weighting at each target weight the dependent variables by the inverse of the standard deviation of the residuals (weighted least squares approach). This practice allowed better estimation of the variables of interest within weight

Table 3 Predictors used for each approach

	Linear/allometric ^a	Quadratic ^a	Combination ^b
Lean meat (%)	PLean	PLean, PLean2	Ham perimeter, ham superior subcutaneous fat, loin superior subcutaneous fat thickness, loin lateral subcutaneous fat thickness, diagonal muscle thickness, loin area
Main cuts			
Lean	VolLean	VolLean, VolLean2	Ham width, loin width
Fat	VolFat	VolFat, VolFat2	Loin superior subcutaneous fat, ham superior subcutaneous fat
Bone	VolBone	VolBone, VolBone2	–
Ham			
Weight	TotalVol	Total, Total2	Loin superior subcutaneous fat
Lean	VolLean	VolLean, VolLean2	Loin superior subcutaneous fat
Fat	VolFat	VolFat, VolFat2	Ham superior subcutaneous fat and fat area of the ham
Bone	VolBone	VolBone, VolBone2	Ham lateral fat
Loin			
Weight	TotalVol	Total, Total2	Loin superior subcutaneous fat
Lean	VolLean	VolLean, VolLean2	Loin area, loin perimeter
Fat	VolFat	VolFat, VolFat2	Ham superior subcutaneous fat, loin superior subcutaneous fat
Bone	VolBone	VolBone, VolBone2	Ham perimeter, loin superior subcutaneous fat
Shoulder			
Weight	TotalVol	Total, Total2	Loin superior subcutaneous fat
Lean	VolLean	VolLean, VolLean2	Ham subcutaneous fat area
Fat	VolFat	VolFat, VolFat2	Ham perimeter
Bone	VolBone	VolBone, VolBone2	Ham superior subcutaneous fat, loin lateral fat
Belly			
Weight	TotalVol	Total, Total2	Loin superior subcutaneous fat
Lean	VolLean	VolLean, VolLean2	Loin area, PLean
Fat	VolFat	VolFat, VolFat2	Ham lateral subcutaneous fat
Bone	VolBone	VolBone, VolBone2	Loin superior subcutaneous fat
Filet	VolLean	VolLean, VolLean2	Loin superior subcutaneous fat

^aVolLean: volume between 0 Hounsfield units (HU) and 140 HU; VolFat: volume between –149 and –1 HU; VolBone: volume between 141 and 1400 HU; TotalVol: volume between –1000 and 1400 HU; PLean: $100 \times \text{VolLean}/\text{TotalVol}$.

^bPredictors were the volume or ratio used in the linear approach plus the variables in this column. These variables are not always the same for all the genotypes.

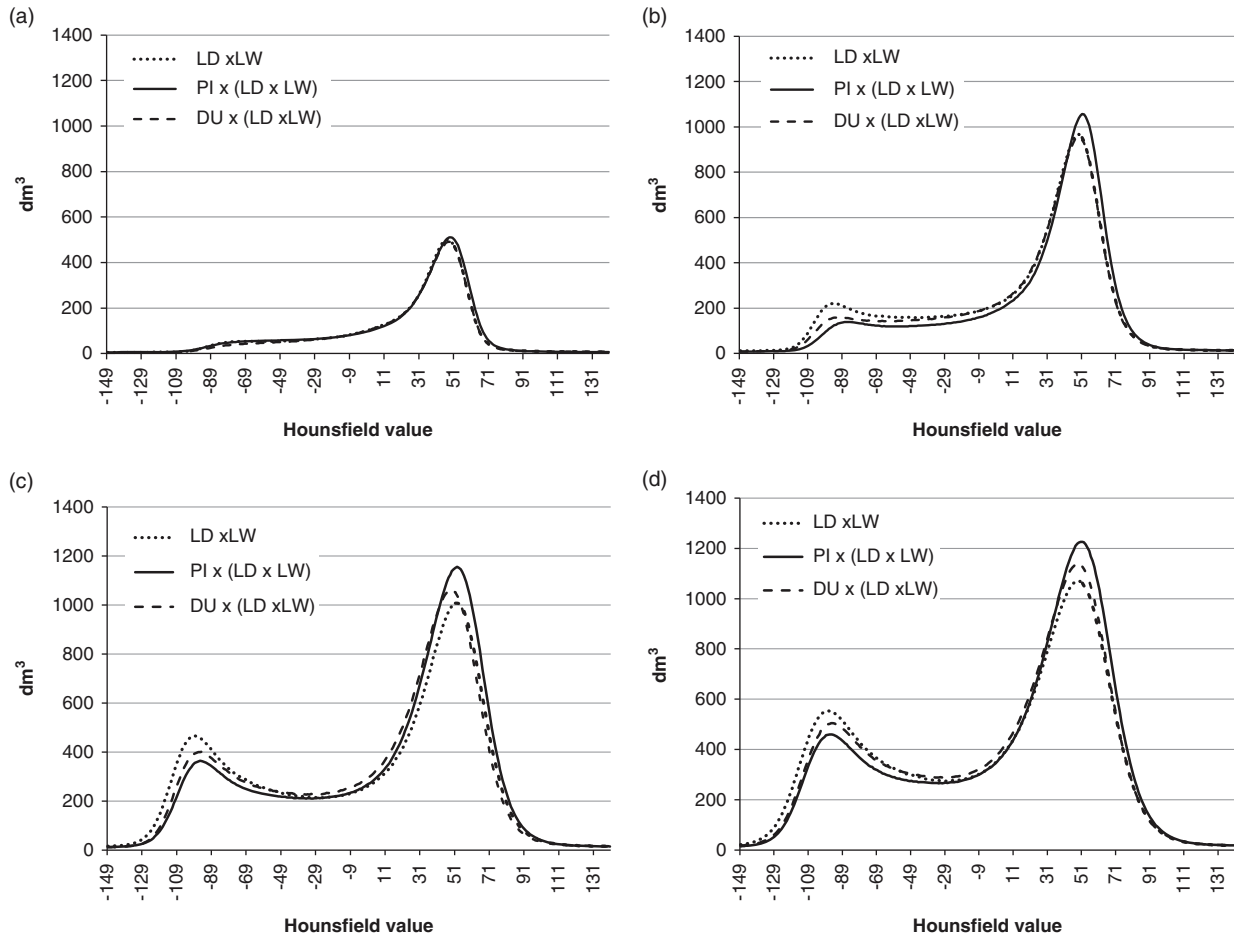


Figure 2 Average volumes associated with each Hounsfield value (in Hounsfield units (HU)) in the fat and lean area by genotype at target weights of (a) 30, (b) 70, (c) 100 and (d) 120 kg. Genotypes: LD x LW = Landrace x Large White; PI x (LD x LW) = Pietrain x (Landrace x Large White); DU x (LD x LW) = Duroc x (Landrace x Large White).

groups and adequate distribution of the residuals. The regression approaches studied were the following (see Table 3 for a detailed description of predictors):

- Linear regressions using CT volumes or CT ratios of volumes as predictors.
- Quadratic regressions using the previous CT volumes or CT ratios of volumes and their squared value as predictors.
- Allometric equations ($y = ax^b$ linearized as $\log y = \log a + b \cdot \log x$), in which CT predictors were chosen as for the previous regression models.
- Linear regression using CT volumes, CT ratios of volumes and direct physical measurements recorded on loin and ham images as predictors. Predictors were selected using the stepwise procedure of SAS (selected criteria: $P < 0.15$) and subjective criteria maximizing the coefficient of determination (R^2) and minimizing root mean square error (RMSE).

The use of prediction models with only direct physical image measurements was not considered, as previous results showed lower prediction accuracy compared with the use of volumes. Transformation of CT volumes to weight by means of voxel densities was also considered, but preliminary

results also showed low prediction accuracy and it was decided to work directly with volumes.

Parameters used to quantify the predictive ability of the equations were the R^2 and the RMSE. Furthermore, the coefficient of variation (CV_d), that is, the percentage of RMSE with respect to the mean, was calculated to make errors comparable.

A cross-validation leave-one-out was used to determine the RMSE of prediction (RMSEPCV) by means of a SAS macro adapted from those presented by Causeur *et al.* (2003). The CV_p computed as the percentage of RMSEPCV with respect to the mean, was calculated. Equations with the lowest CV_p for each trait and genotype were selected.

Results and discussion

The distribution of volumes associated with each HU value by genotype and target live weight is presented in Figure 2. As expected, the volume of live pigs increases as they grow older and heavier. Furthermore, from 70 kg live weight (Figure 2b), a fat peak start appearing (−80 to −120 HU) and grows as pigs gets heavier (Figure 2c and d) indicating that fat is deposited at late stages of life. This is corroborated

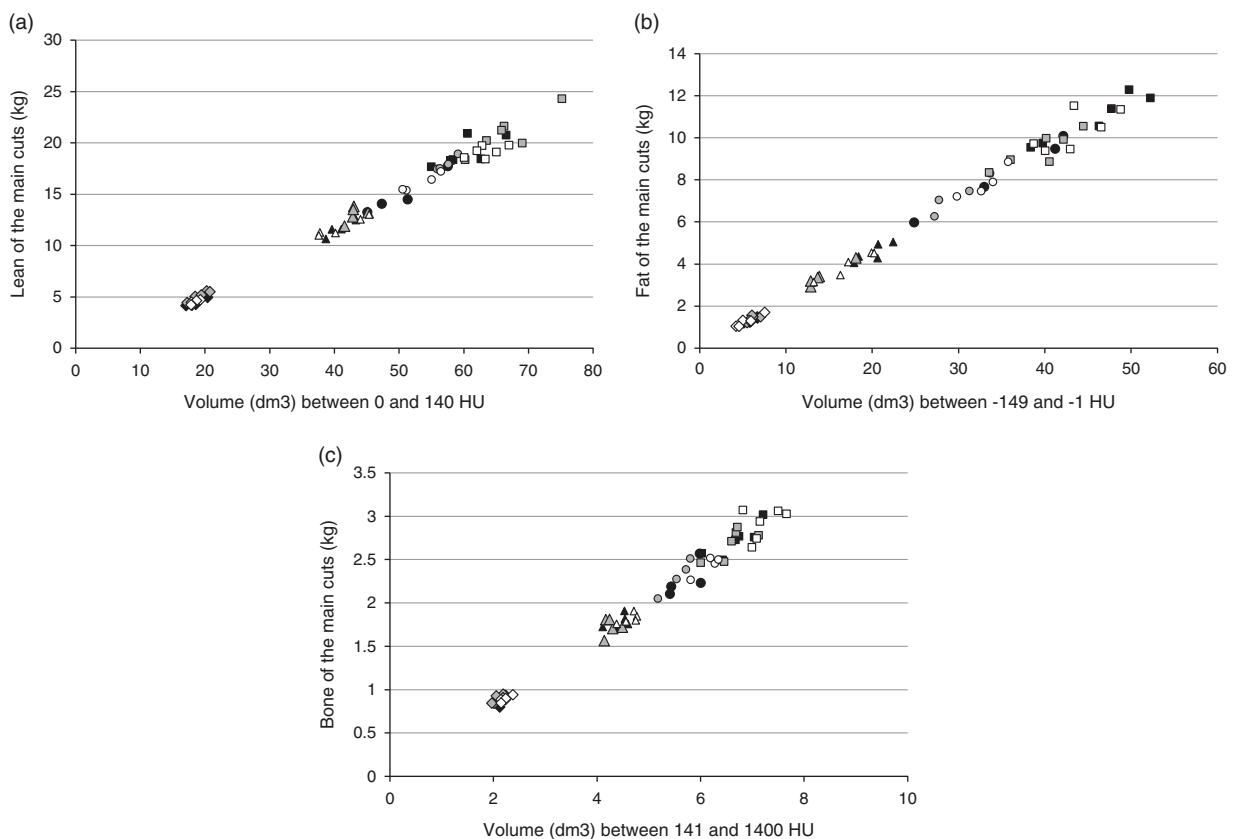


Figure 3 Relationship between the lean (a), fat (b) and bone (c) of the main cuts with the volume of different tissues obtained from the scan of live pigs of different genotypes and weights ($r > 0.99$ for each genotype). Genotypes: black symbols, Landrace \times Large White; grey symbols, Pietrain \times (Landrace \times Large White); white symbols, Duroc \times (Landrace \times Large White). Target weights: \blacklozenge , 30 kg; \blacktriangle , 70 kg; \bullet , 100 kg; \blacksquare , 120 kg. HU = Hounsfield units.

by the fact that some works have shown that the allometric coefficient of fat tissue is higher than one indicating that fat is a late mature tissue (Kouba *et al.*, 1999; Landgraf *et al.*, 2006). It is possible to see differences in the shape of the curve depending on the genotype at each target weight, pointing out that tissue growth patterns differ among genotypes: LD \times LW pigs are fatter, PI \times (LD \times LW) are leaner and DU \times (LD \times LW) are in between.

Fat and muscle thicknesses, perimeters and areas obtained from CT images taken at specific anatomical positions in live pigs have been proven to be good predictors of carcass traits in young pigs (30 kg live weight; Carabús *et al.*, 2011). Nevertheless, the use of images from the whole animal provides more complete information on its composition and allows more accurate determination of fat, lean and bone volumes for the whole carcass. However, the combination of both types of CT information improve prediction equations of some parameters such as lean meat %, loin weight and loin composition. The relationship between lean, fat and bone of the carcass, obtained by dissection of the four main cuts, and volumes associated with these tissues in CT images of whole live pigs (viscera included) of different genotypes and live weights is very strong ($r > 0.99$ for all the tissues and genotypes, Figure 3). It is strong although: (1) volumes are obtained from live pigs and dissection variables are obtained

from cutting and dissection of half carcasses; (2) the live pig images also included white viscera and organs that do not belong in the carcass (some of them are closer to the muscle signal, some others are closer to the fat signal and some others closer to the air signal). The relative weight of white viscera and organs decreases or is kept constant with increasing animal weight, with the exception of flare fat (Font-i-Furnols *et al.*, 2012). Landgraf *et al.* (2006) also found a constant percentage of heart, spleen and kidney, and a decrease in the percentage of lungs and liver during growth. In the present study, the lungs were almost completely removed from the analysis because of their low density (HU values lower than -140), and in agreement with Landgraf *et al.* (2006) and Font-i-Furnols *et al.* (2012), the proportion of the liver with respect to live body weight was low (2.3% to 2.9% at 30 kg and 1.4% to 1.5% at 120 kg). As explained before, the weights of the lean, fat and bone tissues of the main cuts are strongly related to the corresponding volumes obtained from the CT images of whole live pigs (Figure 3). Further research might be done to determine if scanning of live young piglet could be a good predictor of slaughter performances.

The accuracy of the prediction in terms of relative RMSE (CV_d) and RMSEPCV (CV_p) of carcass composition and cut composition traits for each evaluated model is presented in

Table 4 Coefficients of variation of calibration^a and validation^b (%) of different prediction models and coefficient of determination (R^2) of overall carcass lean meat and cut weights and tissue weights for Landrace \times Large White pigs

	Mean	Lineal		Quadratic		Allometric		Lineal + measurements	
		CV_c	CV_p	CV_c	CV_p	CV_c	CV_p	CV_c	CV_p
Lean meat (%)	55.46	2.39	2.60	2.14	2.46	2.34	2.55	1.42	1.81 ^e
Main cuts ^c (kg)									
Lean	12803	5.45	5.85	5.03	5.80	5.01	5.41 ^e	4.25	5.57
Fat	6402	4.28	4.65	4.29	5.08	4.30	4.68	3.44	3.89 ^e
Bones	1934	4.42	4.77 ^e	4.40	4.99	4.41	4.77	4.42	4.77
Ham (kg)									
Weight	7642	5.40	5.80	5.24	5.90	5.28	5.66 ^e	5.27	5.92
Lean	5097	5.58	6.08	5.00	5.86	5.10	5.54 ^e	5.03	5.58
Fat	1921	6.73	7.35	6.76	8.31	6.81	7.45	5.42	5.97 ^e
Bones	623	5.59	5.96	5.53	6.36	5.64	5.99	5.08	5.65 ^e
Loin (kg)									
Weight	5205	6.04	6.56	5.83	6.77	5.90	6.44	5.17	6.22 ^e
Lean	2780	6.65	7.06	6.66	7.51	7.80	8.33	5.40	6.85 ^e
Fat	1774	8.50	9.23	8.14	9.31	8.16	8.99	8.42	8.44 ^e
Bones	651	8.60	9.36	8.41	9.57	8.59	9.32	7.86	9.17 ^e
Shoulder (kg)									
Weight	4340	8.84	9.48 ^e	8.75	9.63	8.84	9.48	8.85	10.29
Lean	2716	8.06	8.63	7.64	8.90	7.84	8.38	7.87	8.36 ^e
Fat	1200	10.01	10.84	8.53	9.73 ^e	8.96	9.77	9.19	10.63
Bones	425	5.57	5.90 ^e	5.49	6.09	5.63	5.99	5.03	6.62
Belly (kg)									
Weight	3503	9.27	10.08	8.67	9.84	8.76	9.53 ^e	9.30	11.18
Lean	1760	10.49	11.32	9.84	11.02	9.99	10.79 ^e	8.91	11.84
Fat	1507	10.43	11.53 ^e	10.46	12.24	10.86	11.90	9.35	11.57
Bones	236	8.50	9.23	8.36	9.25	8.48	9.21	7.06	8.78 ^e
Tenderloin (kg)	450	5.13	5.61 ^e	5.14	6.20	5.45	5.96	9.43	11.14
R^2 lean%		0.92		0.94		0.93		0.97	
R^{2d}		>0.94		>0.94		>0.94		>0.89	

RMSE = root mean square error.

^a CV_c : $100 \times (\text{RMSE}/\text{mean})$.^b CV_p : $100 \times (\text{RMSE of prediction by cross-validation} - \text{RMSEPCV}/\text{mean})$.^cHam, loin, shoulder, belly and tenderloin (tenderloin only for lean).^dRest of the variables.^eEquation with the lowest CV_p .

Table 4 for LD \times LW, in Table 5 for PI \times (LD \times LW) and in Table 6 for DU \times (LD \times LW). These predictions have been done for each genotype across the different weight categories. The large BW range is responsible for the high R^2 values ($0.89 < R^2 < 0.95$) observed in all the prediction models. These R^2 values were somewhat lower when estimating lean meat percentage but were always higher than 0.86. In general, differences in CV_c between the quadratic and linear models are small, indicating that the inclusion of the quadratic term can seldom improve prediction accuracy. However, when CV_p is considered, in general, linear models produced lowest values in comparison with quadratic model, thus, they appear to be more robust. The allometric model yield the lowest CV_p values for some of the variables of interest, especially for PI \times (LD \times LW), although in some cases differences in comparison with the other models do not seem as large. Allometric model is useful for predicting tissue growth with respect to weight (Davies and

Pryor, 1977; Kempster and Evans, 1979; Kouba and Bonneau, 2009), and also in some traits it can be the best choice for estimating tissue growth from tissue volume obtained by CT. Combining the information provided by tissue volumes and measurements (thicknesses, areas or perimeters) improves the accuracy of some of the predictions, especially for LD \times LW and DU \times (LD \times LW). This effect is very important in the estimation of lean meat percentage of the carcass. This importance makes sense, since in carcass classification, linear measurements of fat and muscle depth is well documented (Font i Furnols and Gispert, 2009; Engel *et al.*, 2012). Including physical measurements in the estimation of carcass fat content also reduced the error (CV_p between 3.89% and 6.70%). In a recent study, Lambe *et al.* (2013) showed correlation coefficients of 0.53, 0.14 and -0.28 between fat thicknesses measured in CT images and, respectively, fat, muscle and bone weights obtained by dissection. Bone weights for the four main cuts have

Table 5 Coefficients of variation of calibration^a and validation^b (%) of different prediction models and coefficient of determination (R^2) of overall carcass lean meat and cut weights and tissue weights for Pietrain \times (Landrace \times Large White) pigs

	Mean	Lineal		Quadratic		Allometric		Lineal + measurements	
		CV_c	CV_p	CV_c	CV_p	CV_c	CV_p	CV_c	CV_p
Lean meat (%)	60.03	2.16	2.41	2.08	2.50	2.14	2.39	1.41	1.73 ^e
Main cuts ^c (kg)									
Lean	14532	2.68	2.83	2.57	2.85	2.59	2.73 ^e	2.39	2.85
Fat	5489	5.13	5.70	4.80	5.57	5.38	5.98	4.72	5.50 ^e
Bones	1922	4.52	4.91 ^e	4.49	5.34	4.57	4.95	4.52	4.91
Ham (kg)									
Weight	8212	4.27	4.66 ^e	4.06	4.70	4.48	4.89	4.15	4.69
Lean	5964	4.48	4.86	4.48	5.10	4.56	4.96	4.14	4.63 ^e
Fat	1628	5.23	5.83	5.06	5.97	5.21	5.81 ^e	5.22	6.37
Bones	620	4.21	4.51 ^e	4.17	4.93	4.28	4.58	4.05	4.70
Loin (kg)									
Weight	5362	4.33	4.73 ^e	4.26	4.95	4.38	4.80	4.25	5.32
Lean	3232	4.96	5.41 ^e	4.84	5.67	4.97	5.40	3.11	3.91
Fat	1485	10.39	11.43 ^e	10.29	11.88	10.38	11.50	7.85	11.59
Bones	645	8.66	9.43 ^e	8.33	9.74	8.70	9.44	8.62	9.83
Shoulder (kg)									
Weight	4571	4.84	5.19	4.45	5.12 ^e	4.99	5.36	4.81	5.46
Lean	3001	4.14	4.44	4.09	4.64	4.09	4.40 ^e	4.14	4.61
Fat	1138	9.51	10.34	6.91	8.61 ^e	8.81	9.59	8.31	9.24
Bones	431	7.19	7.60	6.91	7.97	7.17	7.57 ^e	6.55	8.31
Belly (kg)									
Weight	3311	8.47	9.14	8.11	9.29	8.21	8.88 ^e	7.96	8.94
Lean	1848	9.85	10.61	9.46	10.67	9.64	10.40 ^e	9.67	12.10
Fat	1238	9.40	10.49 ^e	9.04	10.85	9.67	10.78	9.07	10.60
Bones	225	8.38	9.07	8.35	9.44	8.34	9.02 ^e	6.64	9.60
Tenderloin (kg)	487	6.67	7.17	6.55	7.38	6.57	7.01 ^e	10.44	11.86
R^2 lean%		0.87		0.88		0.88		0.95	
R^{2d}		>0.94		>0.94		>0.94		>0.95	

RMSE = root mean square error.

^a CV_c : $100 \times (\text{RMSE}/\text{mean})$.

^b CV_p : $100 \times (\text{RMSE of predicción by cross-validation} - \text{RMSEPCV}/\text{mean})$.

^cHam, loin, shoulder, belly and tenderloin (tenderloin only for lean).

^dRest of the variables.

^eEquation with the lowest CV_p .

been estimated with similar accuracy in all the studied models.

Regarding ham weight, the lowest CV_p was obtained with the allometric model for LD \times LW genetic line (RMSEP = 432 g), with the linear model for the PI \times (LD \times LW) genetic line (RMSEPCV = 382 g) and with the lineal model plus measurements for the DU \times (LD \times LW) genetic line (RMSEPCV = 296 g). Linear, allometric and linear models using physical measurements obtained the lowest CV_p for ham lean, fat and bone contents. The CV_p of the different ham tissues and ham weight were similar. However, in almost all cases, these errors were slightly higher than those obtained when estimating the tissue composition of the four cuts together. This difference makes sense, because these estimates were obtained from whole-animal images, which are closer to the composition of the four main cuts than to that of only the ham. Nevertheless, different ham tissue weights are highly correlated with the

tissue volumes of the whole body of the live pig ($r \geq 0.99$ for the weight and all the tissues for each genotype; Figure 4). However, the errors for loin weight (CV_p between 4.73% and 6.65%) were lower than those observed for fat (CV_p : 8.44% to 16.59%) and bone (CV_p : 8.57% to 9.43%) tissues. In general, CV_p was higher for loin than those observed for ham parameters. This difference indicates that whole-body composition presents a higher correlation with ham composition than with loin composition. It also shows that ham composition is a good predictor of whole-body volumes.

It has been reported that cutting error is greater in the separation of the shoulder from the carcass than in the separation of the ham (Nissen *et al.*, 2006). This can probably explain higher CV_p in the shoulder parameters compared with the ham ones. Thus, the obtained CV_p values were between 4.48% and 9.48% (RMSE between 184 and 384 g) for shoulder weight, between 4.40% and 8.36% for lean

Table 6 Coefficients of variation of calibration^a and validation^b (%) of different prediction models and coefficient of determination (R^2) of overall carcass lean meat and cut weights and tissue weights for Duroc × (Landrace × Large White) pigs

	Mean	Lineal		Quadratic		Allometric		Lineal + measurements	
		CV_c	CV_p	CV_c	CV_p	CV_c	CV_p	CV_c	CV_p
Lean meat (%)	56.59	2.61	2.93	2.54	2.93	2.59	2.91	1.53	2.02 ^e
Main cuts ^c (kg)									
Lean	13053	3.14	3.37	3.04	3.51	3.92	4.16	2.79	3.28 ^e
Fat	5984	6.75	7.34	6.81	7.87	6.79	7.36	5.45	6.70 ^e
Bones	2040	4.82	5.08 ^e	4.69	5.11	4.87	5.13	4.82	5.08
Ham (kg)									
Weight	7461	3.78	4.03	3.43	4.03	3.99	4.25	3.44	3.97 ^e
Lean	5128	4.55	4.85	4.07	4.58 ^e	5.41	5.82	4.50	4.95
Fat	1695	7.58	8.35	7.57	8.95	7.68	8.41	6.82	7.72 ^e
Bones	638	3.00	3.25 ^e	2.98	3.39	3.06	3.33	2.99	3.51
Loin (kg)									
Weight	5344	8.99	9.60	8.34	9.57	8.33	8.98	5.78	6.65 ^e
Lean	2982	8.95	9.36	8.78	9.98	8.70	9.20	5.71	7.23 ^e
Fat	1689	16.44	17.29	16.35	18.42	15.84	16.59 ^e	9.18	17.74
Bones	673	9.90	10.70	9.56	10.70	10.00	10.78	7.32	8.57 ^e
Shoulder (kg)									
Weight	4445	4.14	4.48 ^e	4.11	4.62	4.23	4.58	4.10	4.65
Lean	2738	5.61	6.00	5.61	6.30	5.72	6.15	4.61	5.19 ^e
Fat	1237	7.20	7.86	7.03	7.90	6.92	7.49	6.05	6.95 ^e
Bones	470	7.74	8.35 ^e	7.74	8.68	7.75	8.37	6.53	8.64
Belly (kg)									
Weight	3387	6.04	6.58 ^e	5.78	6.73	6.90	7.52	5.84	6.85
Lean	1766	5.48	5.93	4.84	5.66 ^e	6.53	7.09	5.33	6.42
Fat	1362	10.67	11.64	9.73	11.24 ^e	11.13	12.17	10.47	11.68
Bones	259	7.51	8.14 ^e	7.50	8.71	7.51	8.14	6.93	9.23
Tenderloin (kg)	440	10.21	10.69	10.25	11.25	10.32	10.85	7.49	8.08 ^e
R^2 lean%		0.86		0.87		0.86		0.95	
R^{2d}		>0.94		>0.95		>0.94		>0.95	

RMSE = root mean square error.

^a CV_c : $100 \times (\text{RMSE}/\text{mean})$.

^b CV_p : $100 \times (\text{RMSE of prediction by cross-validation} - \text{RMSEPCV}/\text{mean})$.

^cHam, loin, shoulder, belly and tenderloin (tenderloin only for lean).

^dRest of the variables.

^eEquation with the lowest CV_p .

weight, between 6.95% and 9.73% for fat weight and between 5.90% and 8.35% for bone weight.

The belly was the cut for which weight and composition were predicted the worst from whole-pig CT images, although the relative errors were not considerably higher than those for the shoulder. According to Nissen *et al.* (2006), the belly produces a high dissection error because of the thin layers of fat and muscle, which are difficult to separate by knife. Additionally, these thin layers may also be the reason for the large number of voxels with partial volume effects, which makes it difficult to assign a given tissue type. There are some differences in the relationship between the lean volume and the weight of the lean of the different cuts. In accordance with the prediction results, this relationship is less precise in the belly than in the other cuts ($r > 0.98$ to 0.99 in the loin and shoulder and $r \geq 0.97$ to 0.99 in the belly, depending on the genotype; Supplementary Figure S2 to S4).

Tenderloin weight error was quite different across the genetic lines. These differences are probably due to the fact that the proportion of fillet in the whole pig body is very small, and consequently, its correlation with whole-body tissue composition is low.

Because of the observed differences in tissue growth patterns, prediction equations were obtained by genotype between 30 and 120 kg BW, and it is worthwhile to see if they fit within weight group. Using the models that yielded the lowest error within genotypes (Tables 4, 5 and 6), prediction equations by weight group was drawn and their respective accuracy was assessed (Table 7 – selected prediction equations are presented in Supplementary Table S2). Lean meat percentage is estimated well for all the slaughter weights, although the accuracy is lower for the lightest carcasses. CT resolution in 30 kg animals probably is better than in heavier pigs, because of lower DFOV applied and lower thickness of the image, thus, this difference in

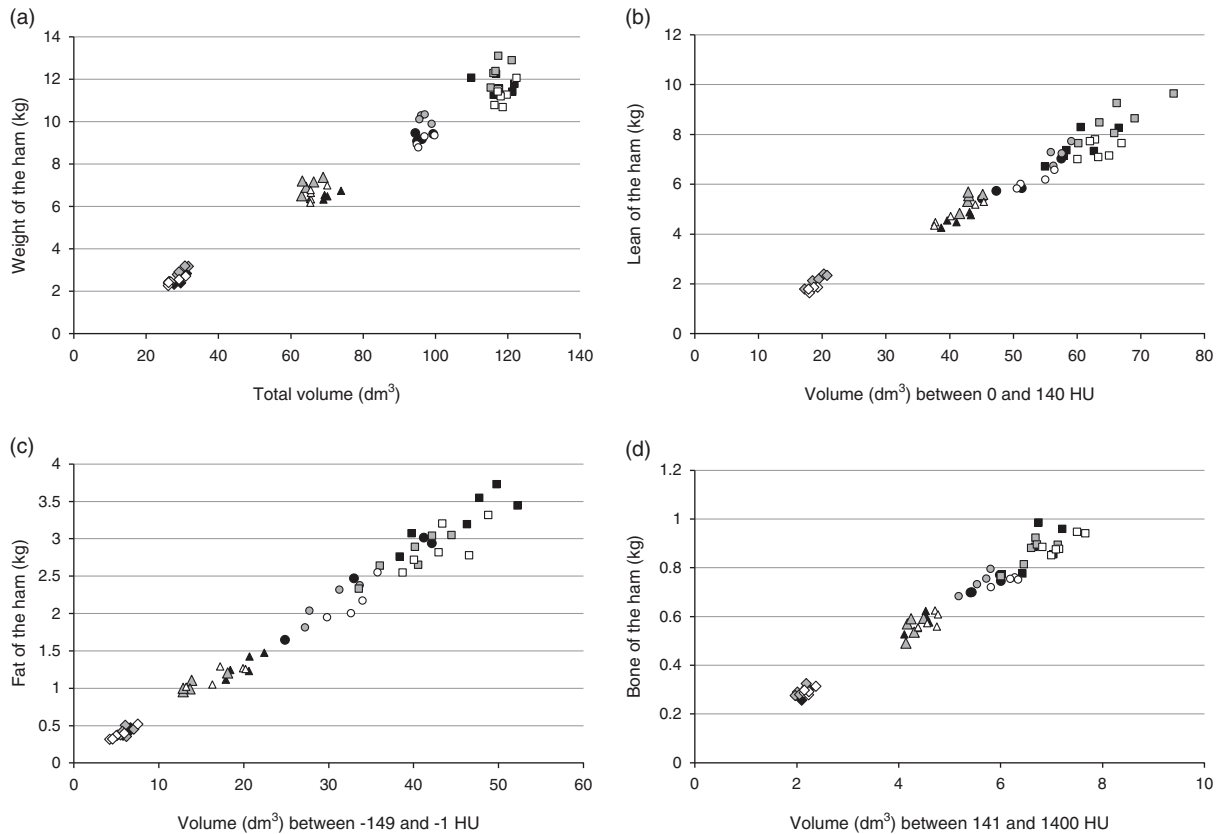


Figure 4 Relationship between (a) the weight (kg), (b) the lean (kg), (c) the fat (kg) and (d) the bone (kg) of the ham with the total, lean, fat and bone volume, respectively, obtained from the scan of live pigs of different genotypes and weights ($r > 0.989$ for each genotype). Genotypes: black symbols, Landrace \times Large White; grey symbols, Pietrain \times (Landrace \times Large White); white symbols, Duroc \times (Landrace \times Large White). Target weights: \blacklozenge , 30 kg; \blacktriangle , 70 kg; \bullet , 100 kg; \blacksquare , 120 kg. HU = Hounsfield units.

accuracy is probably mainly due to difficulties in applying the European cutting procedures (Walstra and Merkus, 1995) and separating the tissues, in particular fat, in 30 kg carcasses. In fact, for the four main cuts, fat tissue shows the highest relative error, with values ranging from 6.42% in the shoulder to 13.97% in the loin. It is also important to note that loin bone and belly bone predictions had lower accuracy in terms of R^2 that was not significant ($P > 0.05$). Loin and belly fat estimates for the 70 kg animals also had a low accuracy, with CV values around 12%. However, in terms of R^2 , the lowest accuracy was for loin bone and belly lean weight ($R^2 < 0.16$). At the 100-kg target weight, models predicting loin and belly weights did not explain a significant portion of the observed variances. At the 120-kg target slaughter weight, loin weight and bone were predicted much better than at the other target weights, while loin fat was predicted with worst accuracy than at 100 kg. Lambe *et al.* (2013) also found higher R^2 values between dissected and CT-predicted fat, muscle and bone of the carcass side with increasing live weight (60, 85 and 115 kg). Moreover, all the belly parameters estimated in the present study also had higher or similar accuracy at the 120 kg target weight than at the others. This can be explained with the fact that heavier carcasses are easier to cut and dissect, although that does not seem to be the case with the shoulder. In fact, only the models predicting

shoulder weights at the 120-kg target slaughter weight did not explain a significant portion of the observed variation, probably because, as reported by Nissen *et al.* (2006), cutting error is the highest for this cut.

Conclusions

There is a very good relationship between cross-sectional CT images obtained in live growing female pigs of different genotypes (including viscera) and carcass and main cuts composition. For this reason, measurements taken on CT images of whole-animal *in vivo* allow accurate estimation of carcass lean meat percentage as well as the weight and tissue composition of cuts in pigs from 30 to 120 kg BW, using empirical regression equations. The prediction accuracy varied across genotypes, variables of interest and BW. Linear models using CT tissue volumes as predictors, allometric models or linear models using CT tissue volumes and physical measurements at specific anatomical positions of the animal body, were in general more robust than quadratic models. However, further work is needed to allow the accurate prediction of pig cut weights and composition using reconstructed 3D pig CT images from which the whole animal body is virtually cut and dissected, thus mimicking dissection by the butcher, in an attempt to render prediction accuracy independent of pig genetic traits and BW.

Table 7 Coefficient of variation (CV)^a and coefficient of determination (R²)^b calculated within weight groups using prediction equations yielding the lower prediction error within genotypes and across weight groups

	30 kg	70 kg	100 kg	120 kg
	CV(%) / R ²	CV(%) / R ²	CV(%) / R ²	CV(%) / R ²
Lean meat (%)	1.52/0.64	1.31/0.95	1.40/0.97	1.55/0.94
Main cuts ^c				
Lean	2.97/0.91	3.32/0.81	2.19/0.97	2.20/0.87
Fat	6.72/0.79	4.30/0.94	3.50/0.95	3.98/0.88
Bone	2.76/0.68	4.39/0.29	3.94/0.71	4.56/0.58
Ham				
Weight	4.56/0.83	3.53/0.61	2.42/0.81	4.63/0.36
Lean	4.88/0.85	3.01/0.90	2.94/0.94	4.75/0.79
Fat	9.44/0.60	5.32/0.88	4.55/0.94	5.10/0.84
Bone	3.25/0.69	4.63/0.50	2.33/0.80	4.29/0.66
Loin				
Weight	5.63/0.76	5.43/0.65	5.27/0.29	3.98/0.58
Lean	6.44/0.67	5.81/0.76	3.75/0.92	4.98/0.74
Fat	13.97/0.58	12.87/0.83	7.04/0.92	10.67/0.54
Bone	7.93/0.06	8.08/0.01	8.69/0.34	6.45/0.48
Shoulder				
Weight	3.25/0.85	3.62/0.51	5.84/0.36	6.04/0.03
Lean	3.34/0.85	4.05/0.74	3.48/0.93	6.23/0.60
Fat	6.42/0.76	6.31/0.78	6.40/0.66	6.91/0.48
Bone	4.68/0.78	5.36/0.38	5.03/0.69	7.58/0.41
Belly				
Weight	7.37/0.58	8.26/0.31	8.01/0.03	6.16/0.20
Lean	7.65/0.56	8.05/0.15	7.69/0.67	7.71/0.46
Fat	13.62/0.55	11.53/0.78	9.90/0.66	7.38/0.67
Bone	6.81/0.26	6.35/0.44	9.07/0.63	6.54/0.58
Tenderloin	5.73/0.51	4.14/0.69	6.57/0.71	7.84/0.41

^a100 × root mean square error/mean.

^bR²-values in italics are not significant (*P* > 0.05).

^cHam, loin, shoulder, belly and tenderloin (tenderloin only lean).

Also more work is needed to build growth models that would allow relate live young piglet CT images with carcass composition at slaughter.

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Supplementary material

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S1751731114002237>

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