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1 **Active Packaging Applications for Food**

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38

39 **ABSTRACT**

40 The traditional role of food packaging is continuing to evolve in response to changing market  
41 needs. Current drivers such as consumer's demand for safer, "healthier," and higher-quality  
42 foods, ideally with a long shelf-life; the demand for convenient and transparent packaging,  
43 and the preference for more sustainable packaging materials, have led to the development  
44 of new packaging technologies, such as active packaging (AP). As defined in the European  
45 regulation (EC) No 450/2009, AP systems are designed to "*deliberately incorporate*  
46 *components that would release or absorb substances into or from the packaged food or the*  
47 *environment surrounding the food.*" Active packaging materials are thereby "*intended to*  
48 *extend the shelf-life or to maintain or improve the condition of packaged food*". Although  
49 extensive research on AP technologies is being undertaken, many of these technologies have  
50 not yet been implemented successfully in commercial food packaging systems. Broad  
51 communication of their benefits in food product applications will facilitate the successful  
52 development and market introduction. In this review, an overview of AP technologies, such  
53 as antimicrobial, antioxidant or carbon dioxide-releasing systems, and systems absorbing  
54 oxygen, moisture or ethylene, is provided, and, in particular, scientific publications  
55 illustrating the benefits of such technologies for specific food products are reviewed.  
56 Furthermore, the challenges in applying such AP technologies to food systems and the  
57 anticipated direction of future developments are discussed. This review will provide food  
58 and packaging scientists with a thorough understanding of the benefits of AP technologies  
59 when applied to specific foods and hence can assist in accelerating commercial adoption.

60

61 **Keywords:** active packaging, oxygen scavenger, antimicrobial packaging, antioxidant  
62 releaser, ethylene absorber.

63

64 END PAGE 2

65

66 **Nomenclature**

|    |      |                                 |
|----|------|---------------------------------|
| 67 | AA   | ascorbic acid                   |
| 68 | AITC | allyl isothiocyanate            |
| 69 | AnV  | <i>p</i> -anisidine value       |
| 70 | AP   | active packaging                |
| 71 | BEO  | basil leaf essential oil        |
| 72 | BHA  | butylated hydroxyanisole        |
| 73 | BHT  | butylated hydroxytoluene        |
| 74 | BI   | browning index                  |
| 75 | CEO  | cinnamon essential oil          |
| 76 | CFU  | colony-forming units            |
| 77 | CPP  | cast polypropylene              |
| 78 | CSP  | catalytic system with palladium |
| 79 | EDTA | ethylenediaminetetraacetic acid |
| 80 | EO   | essential oil                   |
| 81 | EVOH | ethylene vinyl alcohol          |
| 82 | EFSA | European Food Safety Authority  |
| 83 | FFA  | free fatty acids                |
| 84 | GC   | gas chromatography              |
| 85 | GRAS | generally regarded as safe      |
| 86 | HDPE | high-density polyethylene       |
| 87 | IC   | inhibitory concentration        |
| 88 | IU   | international units             |

|     |       |  |
|-----|-------|--|
| 89  | LAB   | lactic acid bacteria   |
| 90  | LAE   | ethyl-N <sup>α</sup> -dodecanoyl-L-arginate or lauric arginate ester |
| 91  | LDPE  | low-density polyethylene   |
| 92  | LLDPE | linear low-density polyethylene                                      |
| 93  | MA    | modified atmosphere  |
| 94  | MAP   | modified atmosphere packaging  |
| 95  | MDA   | malonaldehyde  |
| 96  | MO    | microorganism  |
| 97  | MRE   | meal-ready-to-eat  |
| 98  | OPET  | oriented polyester   |
| 99  | OPP   | oriented polypropylene   |
| 100 | OS    | oxygen scavenger   |
| 101 | OTR   | oxygen transmission rate   |
| 102 | PBAT  | poly(butylendipate terephthalate)                                    |
| 103 | PCL   | polycaprolactone   |
| 104 | PE    | polyethylene   |
| 105 | PET   | poly(ethylene terephthalate)   |
| 106 | PFO   | polyfuryloxirane   |
| 107 | PLA   | polylactic acid  |
| 108 | PP    | polypropylene  |
| 109 | PS    | polystyrene  |
| 110 | PV    | peroxide value   |
| 111 | PVC   | poly(vinyl chloride)   |

|     |                  |  |
|-----|------------------|--|
| 112 | PVDC             | poly(vinylidene chloride)                    |
| 113 | RH               | relative humidity                            |
| 114 | SAP              | super-absorbent polymer                      |
| 115 | SiO <sub>x</sub> | silicon oxide                                |
| 116 | SP/IP            | smart and intelligent packaging              |
| 117 | TBA              | thiobarbituric acid                          |
| 118 | TBARS            | thiobarbituric acid-reactive substances      |
| 119 | TCC              | total coliform counts                        |
| 120 | TiO <sub>2</sub> | titanium dioxide                             |
| 121 | TMA              | trimethylamine                               |
| 122 | TVC              | total viable counts                          |
| 123 | (U.S.)FDA        | (United States) Food and Drug Administration |
| 124 | WVTR             | water vapor transmission rate                |
| 125 | ZnO              | zinc oxide                                   |
| 126 |                  |  |

127

128 **Introduction**

129 Packaging plays critical role in the food supply chain. The primary function of packaging is to  
130 serve as a container for the food enabling efficient transport within the whole supply chain,  
131 preventing any physical damage, and protecting against manipulation and theft. Packaging  
132 also meets the fundamental need to maintain food quality and safety from production to  
133 final consumption by preventing any unwanted chemical and biological changes. Hence, the  
134 packaging acts as a barrier to protect the food from environmental influences such as  
135 oxygen, moisture, light, dust, pests, volatiles, and both chemical and microbiological  
136 contamination (Coles and others 2003; Yildirim 2011; Arvanitoyannis and Oikonomou 2012;  
137 Pereira de Abreu and others 2012). The protective role of the packaging is primarily passive,  
138 acting as a barrier between the food, the atmosphere surrounding the food, and the  
139 external environment. However, there are some exceptions, such as fresh produce, for  
140 which highly gas permeable or perforated packaging materials are used to allow gas  
141 exchange through the packaging (Lee and Paik 1997; Hussein and others 2015). Such  
142 packaging systems, however, are limited in their ability to further extend the shelf-life of the  
143 packaged food. Over recent decades, consumer concern about the safety and additive  
144 content of food has received much attention. There is an increasing trend to natural high-  
145 quality foods, which are non-processed or minimally processed, do not contain  
146 preservatives, but offer an acceptable shelf-life (Singh and others 2011; Gerez and others  
147 2013). In response, the protective function of packaging has been refined and improved  
148 leading to the development of new packaging technologies, such as modified atmosphere  
149 packaging (MAP) (Ohlsson and Bengtsson 2002; Rodriguez-Aguilera and Oliveira 2009;

150 Sandhya 2010; Cooksey 2014; Zhuang and others 2014), active packaging (AP) (Singh and  
151 others 2011; Yildirim 2011; Arvanitoyannis and Oikonomou 2012; Pereira de Abreu and  
152 others 2012; Dobrucka and Cierpiszewski 2014; Realini and Marcos 2014; Kuorwel and  
153 others 2015; Brockgreitens and Abbas 2016), smart and intelligent packaging (SP/IP) (Kerry  
154 and Butler 2008; Lee and Rahman 2014; Realini and Marcos 2014; Biji and others 2015;  
155 Brockgreitens and Abbas 2016), and the application of nanomaterials (Imran and others  
156 2010, Llorens and others 2012, Rhim and others 2013, Reig and others 2014, Rhim and Kim  
157 2014, Bumbudsanpharoke and others 2015). The emphasis of this review is on active  
158 packaging.

159

160 Active packaging is an innovative approach to maintain or prolong the shelf-life of food  
161 products while ensuring their quality, safety, and integrity. As defined in the European  
162 regulation (EC) No 450/2009, active packaging comprises packaging systems that interact  
163 with the food in such a way as to *“deliberately incorporate components that would release or*  
164 *absorb substances into or from the packaged food or the environment surrounding the food”*  
165 (European Commission 2009). Active packaging systems can be divided into active  
166 scavenging systems (absorbers) and active-releasing systems (emitters). Whereas the former  
167 remove undesired compounds from the food or its environment, for example, moisture,  
168 carbon dioxide, oxygen, ethylene, or odor, the latter add compounds to the packaged food  
169 or into the headspace, such as antimicrobial compounds, carbon dioxide, antioxidants,  
170 flavors, ethylene or ethanol. Table 1 provides an overview of the primary active packaging  
171 technologies and their potential benefits in food applications.

172

173 The addition of active substances, such as antimicrobials and antioxidants, through the use  
174 of AP instead of direct addition to the food, may decrease the amount of such substances  
175 required. Traditionally, active substances are added to the bulk of the food, whereas for  
176 most fresh and processed food, food degradation or microbial growth occurs at the surface  
177 of the food. Furthermore, the activity of the active substances when directly added to food  
178 may be reduced or inhibited as a result of interaction between the active substances and the  
179 food components, and/or during the processing of the food. Therefore, the addition of  
180 active substances via active packaging may be more effective than their addition to the bulk  
181 of the food.

182

183 A large variety of active packaging systems has been developed and, to date, numerous  
184 reviews have emphasized the potential of active packaging technologies to supply safer,  
185 “healthier”, and higher-quality foods to the consumer (Kuorwel and others 2015;  
186 Brockgreitens and Abbas 2016). However, the number of reviews presenting the benefits of  
187 active packaging technologies applied to specific food products is limited (Llorens and others  
188 2012; Pereira de Abreu and others 2012; Cichello 2015). Active packaging technologies that  
189 have been evaluated in model systems may not always behave in the same way in real food  
190 applications. The complex structure of the food may influence the activity of the packaging.  
191 For example, the release rates, absorption rates, or diffusion rates of active substances may  
192 be affected. Moreover, active substances or carriers may react with food components or  
193 bind to them, thereby inhibiting the desired activity. It is therefore important to critically  
194 review active packaging studies pertaining to specific foods thereby enabling food and

195 packaging scientists to better understand the benefits of such systems and potentially  
196 accelerate the adoption of the technologies in commercial applications.

197

198 The primary focus of this review is to provide an overview of those active packaging  
199 technologies that have already been successfully applied to food, thereby highlighting the  
200 benefits for the particular food products. Specific emphasis is given to antimicrobial and  
201 antioxidant packaging systems. Furthermore, packaging systems that emit carbon dioxide or  
202 absorb oxygen, moisture or ethylene, and have been successfully implemented are discussed  
203 in depth.

204

### 205 **Oxygen Scavengers**

206 The application of oxygen scavengers (OS) is one of the main active packaging technologies  
207 that aims to remove any residual oxygen present in the food package (Solovyov 2010;  
208 Arvanitoyannis and Oikonomou 2012; Realini and Marcos 2014) or improve barrier  
209 properties by acting as an active barrier (Sängerlaub and others 2013a). Several food  
210 products are sensitive to oxygen, thus, the food industry seeks to exclude oxygen from food  
211 packaging. This is mainly achieved using gas flushing or modified atmosphere packaging  
212 processes. However, the residual oxygen-concentration in the package often remains  
213 between 0.5-5% (Solovyov 2010; Gibis and Rieblinger 2011; Pereira de Abreu and others  
214 2012) and may further increase during storage. This can be due to insufficient evacuation  
215 during the packaging process, oxygen permeation through the packaging material or poor  
216 sealing (Pereira de Abreu and others 2012), or due to oxygen dissolved in the food itself

217 being released into the headspace of the package to reach equilibrium with the gas phase  
218 (Pénicaud and others 2012).

219

220 Oxygen in packaging negatively affects the quality and shelf-life of several foods as it leads to  
221 oxidation of the product (Choe and Min 2006) or promotes the growth of aerobic  
222 microorganisms (Lee 2010; Solovyov 2010) resulting in color modifications (Møller and  
223 others 2000; Nannerup and others 2004; Larsen and others 2006; Gibis and Rieblinger 2011;  
224 Hutter and others 2016), or sensory changes (Jacobsen 1999; Granda-Restrepo and others  
225 2009a; Li and others 2013), or nutritional losses (Chung and others 2004; Lopez-Gomez and  
226 Ros-Chumillas 2010; Van Bree and others 2012). A reduction in the residual oxygen level of a  
227 food packaging can be achieved through the application of oxygen scavengers, in some cases  
228 down to <0.01 vol.-%. The oxygen-scavenging mechanism is mostly chemical. Most common  
229 are iron-based scavengers (Miltz and Perry 2005; Galotto and others 2009; Braga and others  
230 2010; Gibis and Rieblinger 2011; Polyakov and Miltz 2016), of which the OS activity is  
231 triggered by moisture so that the reduced iron is irreversibly oxidized to a stable ferric oxide  
232 trihydrate complex (Solovyov 2010). In contrast, other applied metals, such as cobalt (Galdi  
233 and others 2008; Damaj and others 2014), act as a catalyst for the oxidation of polymers, or  
234 palladium (Yildirim and others 2015), that catalyzes the oxidation of hydrogen into water.

235 Other systems that scavenge oxygen chemically include photosensitive dyes (Maloba and  
236 others 1996; Miller and others 2003; Zerdin and others 2003; Perkins and others 2007),  
237 ascorbic acid (Matche and others 2011; Pereira de Abreu and others 2012), gallic acid  
238 (Wanner 2010; Ahn and others 2016), or unsaturated fatty acids (Arvanitoyannis and  
239 Oikonomou 2012; Pereira de Abreu and others 2012). Moreover, there are also biochemical

240 mechanisms that work through the use of enzymes (Andersson and others 2002; Fernández  
241 and others 2008; Kothapalli and others 2008; Nestorson and others 2008; Gohil and Wysock  
242 2014), or biological approaches using bacterial spores (Anthierens and others 2011), or  
243 yeasts that are immobilized in a solid material (Edens and others 1992). Some commercially  
244 available oxygen-scavenging solutions are SHELFPLUS® O2 (OS-masterbatch, Albis Plastic  
245 GmbH, DE), AMOSORB™ ColorMatrix™ (different OS-solutions, PolyOne™, Europe Ltd., UK),  
246 Cryovac® (OS-film, Sealed Air Corporation, USA), AGELESS OMAC® (OS-film, Mitsubishi Gas  
247 Chemical Inc., USA), OxyRx® (OS-containers, Mullinix Packages Inc., USA), or Aegis® OXCE  
248 (OS-masterbatch, Honeywell International Inc., USA).

249  
250 Much published work about oxygen-scavenging technologies suggests they have great  
251 potential in food applications. However, such research has mainly been performed using  
252 oxygen-scavenging sachets containing iron powder (Charles and others 2003; Solovyov 2010;  
253 Antunez and others 2012; Cruz and others 2012; Kartal and others 2012; Chounou and  
254 others 2013; Cichello 2015). In contrast to Asia or the USA, sachet-based applications are not  
255 well accepted by consumers in European countries (Restuccia and others 2010), as they are  
256 recognized as foreign bodies in food containers. In fact, the risk of accidental breakage,  
257 which can lead to involuntary consumption of the content, is only one of the disadvantages  
258 of such sachet-based active packaging technologies. Further drawbacks include the  
259 requirement of an additional packaging operation step or their unsuitability in combination  
260 with beverages or moist foods due to moisture sensitivity (Suppakul and others 2003; Day  
261 2008; Pereira de Abreu and others 2012). Alternatively, several new oxygen-scavenging  
262 technologies have been developed over the last decade, such as incorporating active

263 substances directly into packaging films or containers. However, only few of them have been  
264 successfully implemented in real food systems. Consequently, studies demonstrating the  
265 benefits of alternative oxygen-scavenging systems to particular food products are rather  
266 rare. Table 2 provides an overview of several oxygen-scavenging technologies which have  
267 already been applied to food products.

268

269 **Iron.** Shin and others (2009) applied an iron-based OS packaging to extend the shelf-life of  
270 processed meat products. Meatballs were packed in active PP-based multilayer trays  
271 containing OS materials (40, 80 and 100% w/w) in the middle layer. During a storage time of  
272 up to 9 months at 23 and 30 °C, oxidative-induced color and flavor changes of the meatballs  
273 packaged in the active OS containers (100% w/w) were significantly lower compared to  
274 those in the passive packages. This was also confirmed by TBA values (thiobarbituric acid)  
275 indicating less lipid peroxidation of meatballs in these OS packages.

276

277 Military rations constitute a range of products in which a long shelf-life is of particular  
278 interest. These food products are critical because the military requires good stability for a  
279 minimum of 3 years without refrigeration (Gomes and others 2009) which presents a  
280 challenge, especially with components with high oil content that are susceptible to oxidative  
281 deterioration. Within this context, Gomes and others (2009) investigated the influence of an  
282 iron-based OS-containing laminate material for its ability to extend the shelf-life of a hot-  
283 filled meal-ready-to-eat (MRE) cheese spread, a component of military operational rations.  
284 The authors demonstrated that the proposed O<sub>2</sub>-absorbing laminate reduced the initial  
285 headspace oxygen concentration in MRE pouches from 20.4 to 6.82 vol.-% (67.44 vol.-%

286 decrease) within the first 24 h, was further reduced to below 1 vol.-% within 11 days of  
287 storage, and it remained below this level throughout the whole storage period (1 year). The  
288 oxygen concentration in the regular MRE pouches also decreased by 50% during the first  
289 15 days and remained constant at a concentration of 5 vol.-%. This oxygen decrease,  
290 however, was assigned to the oxidative degrading reactions of the food. After 1 year of  
291 storage, the positive effect of the O<sub>2</sub>-absorbing laminate was illustrated through MRE cheese  
292 spread with significantly lower rancidity and higher sensory acceptance compared to MRE  
293 stored in packages without OS. Furthermore, the vitamin C content of the samples in the O<sub>2</sub>-  
294 absorbing laminate could be better preserved resulting in an almost 1.5 times higher  
295 vitamin C content compared to the control samples. Thus, the authors clearly demonstrated  
296 that the O<sub>2</sub>-absorbing laminate removed oxygen before it was otherwise available for  
297 degrading reactions in the food product.

298

299 A novel iron-based oxygen scavenger using iron nanoparticles, blended with activated  
300 carbon, sodium chloride, and calcium chloride, was produced and evaluated by Mu and  
301 others (2013). Nano-sized iron-based oxygen scavenger (110 nm average particle size)  
302 exhibited a higher oxygen-scavenging rate (estimated 13.5 mL/d) compared to the oxygen  
303 scavenger-containing microscale pure iron powder (about 20 µm; estimated 1.8 mL/d).  
304 Moreover, the scavenger capacity of the nano-sized scavenger was found to be almost 1.4  
305 times higher than that of its micro-sized counterpart. This advantage was successfully  
306 utilized to inhibit lipid oxidation in lipid-containing foods. Storage of roasted sunflower seeds  
307 and walnuts over a period of 120 days showed that the iron nanoparticles were an effective  
308 means of inhibiting lipid oxidation. While the peroxide values (PV) of the control samples

309 without OS increased (sunflower seeds: from 4.32 to 46.89 meq O<sub>2</sub>/kg oil, walnuts: from  
310 2.41 to 21.85 meq O<sub>2</sub>/kg), the PV of the samples containing the nano-sized oxygen scavenger  
311 were shown to be significantly lower (sunflower seeds: 19.82 meq O<sub>2</sub>/kg, walnuts: 8.84  
312 meq O<sub>2</sub>/kg) after 120 days of storage. Similar results were obtained for secondary lipid  
313 oxidation. Although significant differences in the *p*-anisidine values (AnV) between the  
314 samples were only observed after 40 and 80 days of storage of sunflower seed and walnut  
315 samples, respectively; the AnV of the samples with the scavenger were about half that  
316 without OS after 120 days. Nevertheless, the use of nanosized iron powder may further  
317 reduce consumer acceptance due to possible leakage and unintentional consumption.

318

319 In some cases, preservation of food quality can be affected even if high barrier and modified  
320 atmosphere packaging are applied, for example, if the sealing layer of packages exhibits  
321 defects, particularly when these defects have a critical size that is below the detection limit  
322 of standard leak testers of 10 µm (Sasaki and Kamimura 1997; Sänglerlaub and others  
323 2013a). In this context, Sänglerlaub and others (2013a) simulated food packages with pinhole  
324 defect sizes of 10 µm. They performed long-term storage experiments (300 days) to  
325 compare O<sub>2</sub> absorption with a snack food product, salami in a baked bread roll, in packages  
326 with and without an iron-based multilayer OS film. Although reactions of the food with  
327 oxygen could not be fully prevented, oxidation reactions were significantly reduced by the  
328 application of the OS film. Salami samples packed with OS showed less difference in color  
329 ( $\Delta E$  of 5.4) compared to those packed without OS ( $\Delta E$  of 6.9). Additionally, the use of OS film  
330 led to less lipid oxidation in the product resulting in hexanal concentrations almost 4 times  
331 lower than those in the control samples without OS. Hence, the applicability of an oxygen

332 scavenger layer in the barrier film structures to provide extended protection against oxygen  
333 penetration through such seal defects was confirmed.

334

335 **Ascorbic Acid.** Matche and others (2011) modified LLDPE films by blending them with iron  
336 and ascorbic acid (Fe/AA) or zinc and ascorbic acid (Zn/AA). Thereby, the ascorbic acid (AA)  
337 acted as a reducing agent and the transition metals were used to catalyze the oxidation  
338 reaction (Graf 1994). The incorporation of these reactive chemicals led to active films with  
339 an OS performance of 47.6 and 37.4 mL, respectively, in 750 hours. With the application of  
340 the OS films in the form of sealed bags to bakery products, an overall shelf-life extension of  
341 packaged buns and bread slices could be demonstrated. In particular, the study of Matche  
342 and others (2011) showed that microbial growth was retarded from 2 to 5 days in samples  
343 packaged with the OS film. Instrumental texture analysis and moisture analysis revealed that  
344 both bun and bread samples without OS film were significantly firmer and dryer,  
345 respectively, after 4 days. This was explained by the lower water vapor transmission rates  
346 (WVTR) of both Fe/AA (17.2) as well as Zn/AA films (17.4) compared to the pure LLDPE film  
347 (20 g/m<sup>2</sup> 100 gauge/day), which was used as a control. Moreover, both modified variations  
348 showed lower oxygen transmission rates (OTR). Sensory testing additionally supported the  
349 obtained measured data as the bread slices and buns packed in OS films were sensorially  
350 acceptable (softness and taste) up to 5 and 6 days, respectively, whereas the control  
351 samples were not acceptable anymore on the second day. Despite these positive results, the  
352 study lacks information about the package size and volume as well as the evolution of the  
353 headspace oxygen concentration of the packaged bakery products. This hinders the

354 understanding of the capacity of the scavengers and the correlation between the OS activity  
355 and the extension of the mold-free shelf life of the product.

356

357 **Photosensitive dyes.** Maloba and others (1996) used a photosensitive OS film to improve  
358 oxidative stability of sunflower oil. The applied ethyl cellulose polymer film contained the  
359 common organic dyes eosin and curcumin and the synthesized polyether polyfuryloxirane  
360 (PFO). When exposed to light, this OS film uses energy transfer to convert triplet oxygen  
361 ( $^3\text{O}_2$ ) into highly reactive singlet oxygen ( $^1\text{O}_2$ ), which is absorbed irreversibly by the PFO. The  
362 method and mechanism behind such systems are explained by Rooney (1995). The  
363 sunflower oil was stored in the presence of the OS film and oxidative stability was evaluated  
364 by determination of peroxide values (PV) and gas-chromatographic (GC) measurement of  
365 headspace hexanal. To evaluate the influence of illumination, an initial irradiation period of 2  
366 days at 2000 lux was followed by continuous illumination at normal room light (500 lux), the  
367 latter imitating the light level commonly encountered on retailer's shelves. Control samples  
368 were additionally stored under light exclusion. It was shown that sunflower oil stored under  
369 illumination, at 23 °C and 12 weeks of storage, in the presence of the OS film exhibited  
370 significantly higher oxidative stability compared to all control samples. PVs were significantly  
371 lower with about 20 meq/kg (OS film, illuminated) compared to about 65 meq/kg (no OS  
372 film, dark), about 75 meq/kg (no OS film, illuminated + antioxidant) and about 100 meq/kg  
373 for the sunflower oil alone (illuminated). The same trend was observed for the amount of  
374 headspace hexanal indicating a high correlation ( $r > 0.95$ ) between the 2 methods for  
375 measuring rancidity. Thus, the OS film showed the potential to be applied with a wide range  
376 of foods that contain polyunsaturated oils and that are stored under illumination at the

377 point of sale for long periods. However, industrial application of such an OS system might be  
378 challenging, as the initial irradiation has to be optimized to ensure that the oxygen  
379 concentration in the headspace is reduced below the critical oxygen concentrations to the  
380 specific food products.

381

382 A similar OS film has also been successfully implemented with orange juice to protect juice  
383 from oxidative degradation. Zerdin and others (2003) performed a storage study (1 year)  
384 with orange juice filled in vacuum-sealed OS pouches. The OS activity of these pouches was  
385 triggered by UV illumination just prior to packaging. The authors demonstrated that at 25  
386 and 4 °C, the initial dissolved oxygen concentration of 2.7 ppm in the orange juice samples  
387 packed in OS pouches was reduced to below 0.04 ppm within 3 and 7 days, respectively. In  
388 contrast, for the control pouches without OS, dissolved oxygen in the orange juice was  
389 above 0.04 ppm up to at least 35 and 77 days, respectively, clearly reflecting the impact of  
390 the competition for the oxygen between the OS film and the ascorbic acid as well as the  
391 impact of temperature. This was confirmed by the correlating ascorbic acid retention which  
392 was significantly higher for the samples using the OS pouches with 30.0% (25 °C) and 73.2%  
393 (4 °C) compared to the control pouches showing an ascorbic acid retention of 7.29 (25 °C)  
394 and 51.3% (4 °C) after one year of storage. Furthermore, it was shown that the loss in  
395 ascorbic acid also correlated with an increase in the non-enzymatic browning of the juice.  
396 Samples in the OS pouches stored at 4 °C had a browning index below 0.15 during the entire  
397 storage period, indicating freshly pressed orange juice (Johnson and others 1995). Storage at  
398 25 °C led to an increased browning, however, the browning index of samples in the OS  
399 pouches was significantly lower with about 0.34 compared to that of the control which was

400 about 0.44. Hence, the rapid removal of oxygen was found to be an important factor in  
401 sustaining a higher concentration of ascorbic acid and color preservation in orange juice over  
402 long storage. Inclusion of such an OS film to juice packaging might enable juice producers to  
403 reduce or omit the use of antioxidant substances, however, the additional step of UV  
404 illumination on the production line needs to be taken into consideration.

405

406 In dairy products such as probiotic yogurt, the application of oxygen scavengers can be of  
407 particular interest since dissolved oxygen has a negative effect on the viability of probiotic  
408 bacteria such as *Lactobacillus acidophilus* and *Bifidobacterium* spp. which are essential for  
409 yogurt production (Shah and others 1995). To control the amount of dissolved oxygen, Miller  
410 and others (2003) applied an OS film containing a reducible organic compound such as a  
411 substituted anthraquinone (Rooney 1999). Unlike the OS film of Maloba and others (1996),  
412 this film did not require a constant source of light. It only required UV light exposure to  
413 trigger the scavenging process. Miller and others (2003) tested different manufacturing  
414 methods of probiotic yogurt as well as different packaging systems, and they evaluated their  
415 effect on the dissolved oxygen content during a normal shelf-life for yogurt (42 days). Best  
416 results were obtained by fermenting set-type yogurt in an oxygen-barrier container lined  
417 with an OS film. The initial dissolved oxygen concentration of 16 ppm was decreased to  
418 3 ppm (normal container) and 1.7 ppm (OS-container) after the first day. It further  
419 decreased to 0.2 ppm and 0 ppm, respectively, after 42 days. The rapid oxygen reduction  
420 observed within the first day was highlighted by the authors to be of particular importance  
421 regarding the probiotic bacteria which require low oxygen concentration for post-  
422 fermentation, thereby leading to a product with increased health benefit. In practical

423 applications, it is important to note that OS-lidding films can only be successfully applied  
424 with high oxygen-barrier containers.

425

426 In ultra-high temperature (UHT) milk, oxygen scavengers have been shown to prevent the  
427 development of stale flavor. Perkins and others (2007) packaged indirectly processed UHT  
428 milk using packaging films containing a prototype ZerO<sub>2</sub><sup>TM</sup> OS film laminate. Thereby, the OS  
429 film significantly reduced the initial dissolved oxygen content of ~7 mg/L to ~3.8 mg/L during  
430 14 weeks of room temperature storage (26 °C), compared to the control with ~5 mg/L.

431 Significant reductions of 23-41% were also observed for stale flavor volatiles such as methyl  
432 ketones and aldehydes. Regarding lipid oxidation, a gradual increase in total free fatty acid  
433 levels was observed during the 14-week storage period of samples without OS (data of OS  
434 not published). However, the levels remained far below threshold values, indicating low  
435 lipolytic rancidity. As a consequence, the consumer panel failed to detect a significant  
436 difference in odor between the samples with and without OS. The authors concluded that  
437 sensory analysis would have better reflected the rancid off-flavor, however, a lack in  
438 regulatory approval for the OS prototype used in their study precluded taste-testing.

439

440 **Unsaturated hydrocarbon dienes.** Baiano and others (2004) incorporated an oxygen-  
441 scavenging copolyester-based polymer (Amosorb DFC 4020, ColorMatrix Europe Ltd,  
442 Knowsley, UK) into PET bottles. The oxygen-scavenging principle was based on a transition  
443 metal, such as cobalt salt, which catalyzed the reaction between oxygen and unsaturated  
444 hydrocarbon dienes that are linked to polyester (Cahill and Chen 2000). The authors  
445 evaluated the influence of the OS PET bottle on ascorbic acid degradation and browning in a

446 model system simulating citrus juice. With a 16-week storage period at 5 and 35 °C, the  
447 authors demonstrated that the use of OS bottles could significantly slow down the  
448 degradation kinetics of ascorbic acid and the browning reactions. Compared to glass jars and  
449 conventional PET bottles, the vitamin C loss in the OS bottles was half at 35 °C storage and  
450 even 3-4 times lower at the usual refrigerated storage of 5 °C. The authors reasoned the  
451 inferior results with PET and glass were due to the oxygen permeability of PET and the  
452 presence of pro-oxidant substances and catalysts in glass containers. The results  
453 demonstrated that glass containers could be advantageously replaced with polymeric  
454 bottles including an oxygen scavenger, particularly in the case of beverages containing  
455 ascorbic acid. However, the application of a real fruit juice instead of a model system would  
456 have been preferable. Using the above-mentioned OS polymer in the form of a cast-  
457 extruded monolayer-PET film, Galdi and Incarnato (2011) demonstrated the prevention of  
458 the browning of bananas. Fresh-cut banana slices were shown to have significantly less  
459 (~50%) color difference ( $\Delta E$ ) after three days wrapped in the OS film compared with the  
460 conventional PET film. Later optimizations of these OS PET films resulted in co-extruded  
461 multilayer films where the internal active layer was protected from fast oxidation by 2  
462 external layers of pure PET (PET/OS-PET/PET) in order to increase the reaction time (Di Maio  
463 and others 2015).

464

465 **Palladium.** Many of the oxygen-scavenging systems that have been developed are still too  
466 slow for several food applications, such as boiled meat products, especially if they are  
467 packaged in slices. For such oxidation-sensitive products, removal of oxygen by conventional  
468 means is not generally achievable since light-induced discoloration in meat occurs within

469 hours, even at very low oxygen levels (0.5 until 0.1 vol.-%), depending on the  
470 product/headspace ratio (Andersen and Rasmussen 1992; Møller and others 2000;  
471 Nannerup and others 2004; Larsen and others 2006; Gibis and Rieblinger 2011; Böhner and  
472 others 2014; Hutter and others 2016); and most OS systems require several days to remove  
473 initial headspace oxygen (Matche and others 2011). Recently, a rapid OS system based on a  
474 catalytic system with palladium (CSP) has been developed. Palladium was coated on  
475 PET/SiO<sub>x</sub>-films using magnetron sputtering technology (Lohwasser and Wanner 2005;  
476 Yildirim and others 2010, 2015). This OS film was able to remove up to 2.5 vol.-% residual  
477 oxygen in food packages if hydrogen was included in the modified atmosphere (MA) of the  
478 packaging (Yildirim and others 2015). Due to its high efficiency, this film is particularly  
479 suitable for food products which are very susceptible to oxygen and where the oxidation  
480 reactions are very fast. Hutter and others (2016) showed that an implementation of this OS  
481 film in packaging prevented the discoloration of cooked cured ham. The OS film removed the  
482 2 vol.-% initial oxygen in the headspace (160 cm<sup>3</sup>) of MA-packed ham within 35 min after  
483 packaging. In this way, the color of the ham could be preserved and discoloration prevented  
484 for 21 days of storage, although packages were exposed to light 24 h/day. In contrast,  
485 samples packaged without OS film and stored in light significantly lost their redness within 2  
486 hours of storage. The same OS film was applied to bakery products. For packaged breads,  
487 mold growth is the key factor limiting shelf-life. In this context, Rüegg and others (2016)  
488 applied the palladium-based OS film in MA packages of partially baked buns, toast bread  
489 slices, and gluten-free bread slices. At normal and modified atmosphere without OS film,  
490 visible mold growth was detected in all samples after 2 days with a simultaneous decrease in

491 headspace oxygen concentration. In contrast, in MA packages with OS film, mold growth was  
492 retarded up to 8-10 days, resulting in a 3-4 fold longer shelf-life for all types of bread tested.

493

494 Apart from its high efficiency, the applied CSP also has some limitations. First of all,  
495 hydrogen is required, therefore, the application of the CSP is limited to products packed  
496 under modified atmosphere. As hydrogen concentrations >5.7 vol.-% in nitrogen are  
497 considered as flammable, a maximum amount of oxygen to be removed was suggested to be  
498 2.5 vol.-% (Yildirim and others 2015). Another drawback is that the catalytic activity of the  
499 palladium-based system may be inhibited or even inactivated by volatile sulfur compounds  
500 present in the headspace of certain packaged food products (Röcker and others 2017). With  
501 regard to the safety of palladium, the EFSA published a scientific opinion on the safety  
502 assessment of the palladium metal and hydrogen gas for use in active food contact  
503 materials. The EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing  
504 Aids (CEF) concluded that *“the active substances palladium and hydrogen do not raise a*  
505 *safety concern for the consumer when used as an oxygen scavenger in packages for foods*  
506 *and beverages at room temperatures or below. Palladium should not be in direct contact*  
507 *with food and should be incorporated in a passive structure impermeable to liquids which*  
508 *prevents the migration at detectable levels”* (EFSA CEF Panel 2014).

509

510 Implementation of OS technologies into food packaging operations is often challenging as OS  
511 systems are sensitive to environmental conditions, such as temperature and pH (Galdi and  
512 others 2008; Solovyov 2010; Damaj and others 2014), or require humidity (Solovyov 2010) or  
513 UV-light (Rooney 1999; Miller and others 2003; Zerdin and others 2003) to trigger the

514 oxidative reaction, or negatively interact with food volatiles (Röcker and others 2017). The  
515 incorporation of OS materials into polymer matrices may change the film properties, such as  
516 OTR or WVTR values (Matche and others 2011). This should be considered as a change in  
517 such properties may result in decreased quality of the food. Additionally, not only the active  
518 substances, but also all other materials used to incorporate active substances into the  
519 packaging should be safe. Finally, OS systems should not have any negative influence on the  
520 sensory properties of the food.

521

522 For the selection of a suitable OS system for a specific food application, factors influencing  
523 the oxidation kinetics, such as storage temperature, humidity, pH of the food and possible  
524 light exposure, should be considered. Information about initial headspace oxygen  
525 concentration, headspace volume, barrier properties of the packaging, as well as the  
526 minimum shelf-life, is also essential to determine the required OS capacity and OS rate.  
527 Finally, the cost of the OS packaging has to correspond with the benefit provided to the  
528 particular food product.

529

### 530 **Moisture Scavengers**

531 Moisture content and water activity are critical factors affecting the quality and safety of  
532 various types of foods (Labuza and Hyman 1998). For instance, many dry products are  
533 sensitive to humidity during storage, and even low relative humidity (RH) levels inside the  
534 packages may cause significant quality deterioration. Increase in moisture makes the  
535 products more prone to microbial spoilage and may cause alterations in texture and  
536 appearance, consequently reducing shelf-life (Labuza and Hyman 1998, Day 2008). For other  
537 products, such as fresh fish, meat, and fruit/vegetables, keeping a controlled high RH level

538 inside the package is beneficial in preventing drying. In addition, some excess liquid caused  
539 by drip loss is common for such fresh products like fish and meat. Consumers perceive liquid  
540 in a package as reducing the attractiveness of a product making it less desirable (Droval and  
541 others 2012).

542

543 Moisture control strategies in packaging can be divided into categories, such as moisture  
544 reduction (for example, by MAP through replacing the humid air in the headspace with dry  
545 modified atmosphere gas, or vacuum-packaging through the removal of humid air in the  
546 headspace), moisture prevention (by barrier packaging), and moisture elimination (by  
547 applying a desiccant/absorber). Only the latter category can be considered active, whereas  
548 moisture reduction and prevention are more passive strategies. Passive systems can include  
549 those that lower the humidity without any active ingredients, such as micro-perforated films  
550 (for example, Xtend® films, Israel) (Suppakul 2015), and other packaging materials that are  
551 inherently hygroscopic, such as those that are 100% cellulose-based (as defined in (EC) No  
552 450/2009 (European Commission 2009)). Humidity levels inside packages can also be  
553 controlled through the appropriate selection of packaging materials with a high barrier  
554 against water vapor.

555

556 Active moisture scavengers can be further distinguished into 2 main types: RH controllers  
557 that scavenge humidity in the headspace, such as desiccants, and moisture removers that  
558 absorb liquids (Brody and others 2001). The latter can be applied in the form of pads, sheets,  
559 or blankets, and are typically placed underneath fresh products in different packaging  
560 concepts (MAP, vacuum, skin pack, and so on). They are applied for foods of high water

561 activity: fish, meat, poultry, fruits, and vegetables (especially cut products) (Vermeiren and  
562 others 1999, Day 2008). Thereby, drip loss increases with storage time and by increasing the  
563 exposed surface area, and longitudinal cutting of the muscle fibers (McMillin 2008). Such  
564 pads are mostly composed of porous materials, polymers (PP or PE), foamed and perforated  
565 PS sheets, or cellulose, combined with superabsorbent polymers/minerals/salts  
566 (polyacrylate salts, carboxymethyl cellulose, starch copolymers, silica/silicates) (Ščetar and  
567 others 2010). Moisture absorbing pads are not often considered to be active packaging.  
568 According to the EU Guidance to the Commission Regulation (EC) No 450/2009, "*Materials*  
569 *and articles functioning on the basis of the natural constituents only, such as pads composed*  
570 *of 100% cellulose, do not fall under the definition of active materials because they are not*  
571 *designed to deliberately incorporate components that would release or absorb substance.*"  
572 However, moisture absorbing pads containing components that "*are intentionally designed*  
573 *to absorb moisture from the food*" and can be considered as active packaging (European  
574 Commission 2009). Absorbing pads can also be used in combination with antimicrobials (for  
575 example, Dri-Fresh® Fresh-Hold™ ABM, Sirane, USA), pH control agents and/or carbon  
576 dioxide generators/oxygen scavengers, to avoid certain shortcomings, such as odor  
577 generation or leakage.

578

579 **Desiccants** are used to control humidity in the packaging headspace. Examples of desiccants  
580 are: silica gel, clays, molecular sieves (synthetic crystalline version, such as from zeolite,  
581 sodium, potassium, calcium alumina silicate), humectant salts (such as sodium chloride,  
582 magnesium chloride, calcium sulfate), and other humectant compounds (such as sorbitol); as  
583 well as calcium oxide (Müller 2013, Day 2008). The absorption capacity of desiccants

584 depends on its water vapor sorption isotherm (Sängerlaub and others 2013b). Desiccants are  
585 commonly placed into packages in the form of sachets, micro-porous bags, or are integrated  
586 in pads. Some examples applied to food products are given in Table 3, however, pads are  
587 excluded as they are already successfully applied on the market.

588

589 **Zeolites.** Zeolites have a significant tendency to attract moisture and can also release the  
590 absorbed water without any change in the crystalline structure and moisture absorption  
591 properties (Julkapli and Bagheri 2016). By virtue of these properties, natural nanozeolite can  
592 be added to pulp/paper and plastic films to regulate the amount of moisture absorbed in  
593 packaging (Julkapli and Bagheri 2016), however, its application is still limited to non-food  
594 packaging (Wu and others 2010).

595

596 **Bentonite/sorbitol/calcium chloride.** One of the potential application of moisture  
597 scavengers is packaging of mushrooms since the quality of the mushrooms is strongly  
598 influence by the RH in the headspace. Humidity below 86% RH promotes moisture and  
599 hence weight loss, mushroom senescence, and textural changes; on the contrary, 100% RH  
600 promotes psychrophilic bacteria growth and also causes discoloration of mushrooms. The  
601 optimal humidity in the headspace of packaging for mushrooms was found to be 96%  
602 (Mahajan and others 2008). Mahajan and others (2008) investigated new packaging  
603 concepts for fresh mushrooms (*Agaricus bisporus*) - a produce with a high moisture content  
604 and short shelf-life (1-3 days at ambient temperature). In this study, combinations of fast-  
605 absorbing (CaCl<sub>2</sub>, KCl, and sorbitol, with moisture holding capacity 0.91 ±0.01 [g H<sub>2</sub>O/g] in  
606 120 h) and slow-absorbing moisture absorbers (bentonite/sorbitol, with moisture-holding

607 capacity  $0.34 \pm 0.02$  [g H<sub>2</sub>O/g] in 120 h), were tested (Table 3). The best combination of  
608 absorbers was found to be bentonite/sorbitol/CaCl<sub>2</sub> in proportions of 0.55:0.25:0.2 g/g  
609 desiccant, respectively. The study revealed that the moisture holding capacity of the  
610 scavenger is dependent on the relative humidity; it increased from 0.51 to 0.94 g water per g  
611 desiccant when the relative humidity was increased from 76% to 96%. A change in  
612 temperature (from 4 to 16 °C) did not have a significant influence on the moisture-holding  
613 capacity. A positive impact of desiccant usage was observed, namely a decrease in the  
614 moisture condensation inside the packaging, and improved transparency of the packaging  
615 film. Mushrooms (250 g) packed with 5 g of bentonite/sorbitol/CaCl<sub>2</sub> desiccants resulted in a  
616 lower browning index (BI 14.8) compared to those packed without desiccants (BI 18) after 5  
617 days of storage at 10°C. However, for packages with higher levels of desiccants (10 or 15 g),  
618 browning indices were greater due to the browning and excessive moisture loss. The  
619 appearance of mushrooms packed with 5 g desiccants was also better since higher amounts  
620 resulted in excessive moisture loss. After 5 days, the mushrooms packed with 5 g of  
621 desiccants were still sellable on the market. The results show that the capacity of the  
622 moisture absorbers had to be precisely tuned and product-adjusted to achieve an optimal  
623 effect, as too high absorption capacities could result in a decrease in quality. Similar  
624 experiments were performed by Azevedo and others (2011). A mixture of desiccants  
625 consisting of calcium oxide/calcium chloride/sorbitol in a ratio of 0.5:0.26:0.24, respectively,  
626 resulted in a moisture-holding capacity of 0.81 g water per g of desiccant. However, no food  
627 application was evaluated in this study.

628

629 **Poly(acrylic acid) sodium salt.** Mbuge and others (2016) investigated the use of a food grade  
630 super-absorbent polymer (SAP) as desiccant for the drying of maize in order to reduce mold  
631 growth, and, consequently, aflatoxin contamination. The applied SAP, a cross-linked  
632 poly(acrylic acid) sodium salt powder, was placed into a porous “tea bag” membrane and  
633 integrated into sealed containers (material not stated) containing fresh maize with a water  
634 content of about 32%. After drying the maize to the optimal water content of 13 % at 40°C  
635 drying temperature, the lowest aflatoxin contamination was observed for the applications  
636 with SAP-to-maize-ratios of 1:5 and 1:1 resulting in aflatoxin contents of 33 or <3 ng/g,  
637 respectively. Aflatoxin contamination could be reduced to <4 ng/g, even at 20, 30 and 40 °C  
638 drying temperatures, with a 24 h frequency change of the SAP (1:5), complying with current  
639 Kenyan and European legislation that limit aflatoxin content to 10 and 4 ng/g, respectively.  
640 The application of poly(acrylic acid) sodium salt therefore shows potential for grain drying in  
641 reducing aflatoxin contamination, particularly in developing countries as it is a cheap and  
642 reusable solution.

643

644 **Sodium chloride and hygroscopic ionomer.** Langowski and others (2006) and Sangerlaub  
645 and others (2013b) developed salt-embedded, humidity-regulating trays consisting of a 3-  
646 layer structure: barrier layer, active layer with NaCl and sealing layer, to control the humidity  
647 in food packages. Thereby, the active layer was foamed and stretched to form cavities  
648 around the salt particles. Such humidity-regulating trays, made of a thermoformed  
649 multilayer structure containing a foamed hygroscopic ionomer Entira™ AS SD100 as an active  
650 layer, were used by Rux and others (2016) to pack tomatoes and strawberries (Table 3).  
651 Thereby, the active layer contained 0 or 12 wt% NaCl. When just water was packed, the

652 amount of water absorbed by the trays containing 0 and 12 wt% NaCl was 7.6 and 13.2 g,  
653 respectively. In the presence of tomatoes or strawberries, the humidity produced by these  
654 products was efficiently absorbed by the trays and no condensation effect was observed.  
655 The trays containing 12 wt% NaCl best regulated the in-package RH below 97%. A slightly  
656 higher product weight loss (2-3 wt% for strawberry, 1 wt% for tomatoes) compared to the  
657 control PP trays (0.3-0.6 wt%) was observed. However, in this study, no other quality  
658 parameters of the strawberries and tomatoes were evaluated. Similar weight loss has been  
659 observed in the study of Rux and others (2015). They reported a water loss of 11.4 g for  
660 packaged mushrooms (250 g) in similar humidity-regulating trays (PP/foamed and stretched  
661 PP -NaCl/EVOH/PE) containing 18 wt% NaCl, compared to 6.7 g water loss for those packed  
662 in standard PP trays, during a storage of 6 days at 7°C. In-package RH remained stable at 93%  
663 during storage. After 6 days, mushrooms packed in humidity-regulating trays had a better  
664 color appearance and gill exposure as well as less incidence of decay, compared to those in  
665 the control PP trays. Singh and others (2010a, 2010b) also confirmed the water loss for  
666 packed mushrooms with such humidity-regulating trays. An increase in the amount of NaCl  
667 integrated in the trays from 6 to 18 wt% resulted in a weight loss within the range of 1.3 to  
668 4.5 g for packaged mushrooms at 5 °C. Differences in moisture loss with the same type of  
669 moisture-absorbing packaging may occur due to the different physiological state of the  
670 product, storage temperatures, and packaging films used.

671

672 **Tamarind seed galactoxyloglucan.** Polysaccharides, such as galactoxyloglucan from tamarind  
673 seeds, can be used as moisture-absorbing aerogels. Such aerogel-based packaging systems,  
674 in combination with enzyme-based (galactose oxidase) oxygen-scavenger systems, were

675 shown to have capacities to absorb water and saline solution 40 times their weight  
676 (Gracanac 2015). With the absorption of drip, however, the absorption capacity was reduced  
677 to 20 times the initial weight. Nevertheless, galactoxyloglucan aerogels have been shown to  
678 have potential to be employed in moisture-absorbent materials for meat packaging  
679 applications.

680

681 Commercial moisture absorbers can be found in a variety of formats including as absorbing  
682 pads, such as Cryovac® Dri-Loc® (Sealed Air Corporation, USA), Thermasorb (Thermasorb PVT  
683 Ltd., Australia), and MeatGuard® or MeatPad® (McAirlaid's Inc., USA); absorbing films, such  
684 as Pichitto/Pichit (MTC Kitchen, Japan), MoistCatch™ (Kyodo Printing Co., Ltd., Japan), and  
685 Active Film™ (CSP Technologies, USA); absorbing paper, such as Onyx Desiccant Paper (Onyx  
686 Specialty Papers, Inc., USA); absorbing pouches, such as Humidor Bag (Boveda Inc., USA),  
687 and trays, such as Fresh-R-Pax® (Maxwell Chase Technologies, LCC, USA).

688

689 As described in this section, various moisture scavenging systems can be applied to preserve  
690 quality and prolong the shelf life of food. The examples presented above underline the  
691 importance of product-adjusted moisture scavenging systems for food applications. The  
692 most well-established and commercially well-recognized technologies incorporate the use of  
693 sachets, pouches (including desiccants) or pads – devices that do not interfere with the  
694 structure of external packaging materials. Implementation of moisture absorbers/controllers  
695 in other forms, such as in the structure of packaging or as a coating, for commercial food  
696 product applications (fresh fruits, vegetables, fish and meat) is still under development and  
697 future research is expected to be focused in this area (Restuccia and others 2010).

698

699 **Ethylene Absorbers**

700 Ethylene (C<sub>2</sub>H<sub>4</sub>) is a growth-stimulating hormone (plant growth regulator) accelerating  
701 ripening and senescence through increasing the respiration rate of fresh and minimally  
702 processed climacteric produce and shortening the shelf-life during postharvest storage.  
703 Ethylene also accelerates chlorophyll degradation rates, especially in leafy products, and  
704 enhances excessive softening of fruits (Saltveit 1999; Ozdemir and Floros 2004). For these  
705 reasons, the removal of ethylene from the product environment by application of ethylene  
706 scavengers slows ripening and senescence, thereby enhancing quality and prolonging shelf-  
707 life.

708

709 **Potassium permanganate.** Ethylene scavenger systems involve either inclusion of a small  
710 sachet containing an appropriate scavenger in the packaging or incorporation of an ethylene  
711 absorber in the film structure. The sachet material should be highly permeable to ethylene,  
712 allowing diffusion through it. The most commonly used active component of the sachet is  
713 potassium permanganate (KMnO<sub>4</sub>) in order to oxidize/inactivate ethylene (Floros and others  
714 1997; Ayhan 2011; Llorens and others 2012) However, KMnO<sub>4</sub> is never used in direct food  
715 contact due to its high toxicity (Martínez-Romero and others 2007).

716

717 **Minerals.** Another ethylene-scavenging system is based on the use of finely dispersed  
718 minerals, such as zeolite, active carbon, or pumice. These minerals could be incorporated  
719 into a plastic film structure commonly used in fresh produce packaging (De Kruijf and others  
720 2002). Such minerals are intended to scavenge ethylene and also modify the gas

721 permeability of the film so that carbon dioxide can diffuse faster and oxygen can enter more  
722 readily than through pure polyethylene to obtain an equilibrium atmosphere (De Kruijf and  
723 others 2002; Esturk and others 2014).

724

725 **Metals and metal oxides** are also good candidates for ethylene removal. Photoactive TiO<sub>2</sub> is  
726 reported to oxidize ethylene into water and carbon dioxide. Since metal oxides are activated  
727 by either UV light, visible light or both, the negative effect of UV exposure on food quality  
728 should be considered. Nano-silver is also claimed as an ethylene blocker; however, it has  
729 been tested in absorbent pads which were placed in trays of fresh-cut melon and not in the  
730 packaging structure (Hu and Fu 2003; Fernández and others 2010). Palladium-based  
731 scavengers are shown to have good ethylene adsorption capacity, but they are mostly tested  
732 as sachets in packages (Abe and Watada 1991; Bailén and others 2006, 2007; Cao and others  
733 2015) or in a storage room (Martínez-Romero and others 2009), not in the structure of  
734 packaging films. The high cost of palladium has been assumed to limit its industrial  
735 application (Martínez-Romero and others 2007). Abe and Watada (1991) reported that  
736 charcoal with palladium chloride as an absorbent, present in paper sachets and not in the  
737 packaging structure, was effective in preventing ethylene accumulation, reducing the  
738 softening in fresh-cut kiwifruit and bananas, and chlorophyll loss in spinach leaves, but not  
739 effective in broccoli pieces. It was also effective in absorbing most of the ethylene during 3  
740 days of storage for kiwifruit slices and banana sections at 20 °C. An ethylene concentration  
741 of 0.4 ppm in the trays of broccoli and spinach was effectively absorbed by the ethylene  
742 absorbent. In this study, 10 g paper packets containing ethylene absorbent were placed in

743 metal trays with a glass cover, however, it was acknowledged that this type of high barrier  
744 packaging is not suitable for products that respire.

745

746 Sothornvit and Sampoompuang (2012) incorporated activated carbon (30%) with 0.3% of the  
747 polysaccharide glucomannan into paper made of rice straw. The active material adsorbed  
748 0.69  $\mu\text{L/L}$  ethylene with a scavenging capacity of 77%. The ethylene adsorption capacity per  
749 surface area was calculated as 34.2  $\mu\text{L/L/m}$ . Therefore, it was suggested that a separate bag  
750 or wrapper or a laminate inside a carton might have the potential to extend the shelf life of  
751 ethylene-sensitive products, such as banana, mango, tomato, and apple. However, no food  
752 application was evaluated in this study. There has also been a further study on cardboard  
753 coated with polylactic acid and ethylene scavengers (clinoptilolite, sepiolite, sepiolite  
754 permanganate) designed as an active packaging for fresh fruits and vegetables. However, in  
755 this study, there was no application to prove the effect in a real food system (Taboada-  
756 Rodríguez and others 2013).

757

758 The incorporation of scavengers in packaging films may be a better option to solve sachet-  
759 related problems. Ethylene scavengers could either be embedded into a solid, dispersed in  
760 plastic, or incorporated into various layers of the packaging (Ozdemir and Floros 2004).

761 However, there has been only limited research into the application of ethylene absorbers in  
762 the structure of packaging films. The main focus of the following section is the application of  
763 ethylene scavengers incorporated into the actual packaging material, rather than in sachet  
764 format, for fresh produce. Ethylene scavenger can prolong the shelf life of climacteric fruits,  
765 such as apples, kiwifruit, apricot, bananas, mango, cucumber, tomato, and avocados, and

766 vegetables, such as carrots, potatoes, and asparagus (De Kruijf and others 2002). The list of  
767 different produce packaged with different packaging film and ethylene absorber and the  
768 benefits of such systems are presented in Table 4.

769

770 **Nano-particles.** Nano-TiO<sub>2</sub> is reported to oxidize ethylene into H<sub>2</sub>O and CO<sub>2</sub> (Han and Nie  
771 2004). Yang and others (2010) tested PE blended with nano powders of Ag, TiO<sub>2</sub>, and kaolin  
772 for preservation of fresh strawberries at 4 °C for 12 days. Results showed that active PE with  
773 nano-powders maintained physicochemical and physiological quality and sensory attributes  
774 of strawberry better than the control (PE). Active packaging decreased the rate of fruit decay  
775 (to 16.7% for nano-packaging and 26.8% for normal packaging), maintained the content of  
776 total soluble solids, preserved ascorbic acid, and reduced the malondialdehyde content ( to  
777 66.3 μmol/g for nano-packaging and 75.4 μmol/g for normal packaging), and enzyme activity  
778 of polyphenol oxidase and pyrogallol peroxidase. However, the gas composition including  
779 ethylene in the headspace was not monitored in this study and there is no indication of shelf  
780 life.

781

782 Chinese bayberries were packaged by Wang and others (2010) with active PE including 30%  
783 nano powder of Ag, TiO<sub>2</sub>, and kaolin-clay or treated with hot air or the combination of hot air  
784 treatment and nano-packaging, and stored at 1 °C and 80-90% RH for 8 days. Results showed  
785 that the application of hot air (48 °C for 3 h) and/or active packaging reduced the incidence  
786 of green mold decay (from 75.5% to 34.6% for active packaging and to 22.7% for hot air  
787 treatment and to 14.8% for the combined treatment), fruit respiration, and ethylene  
788 production, and maintained fruit firmness compared to the control (fruit directly packed in

789 PE with no heat treatment) for 8 days of storage. The respiration rate and ethylene  
790 production of the combined treatment of hot air and active packaging were 49.6% and  
791 25.9%, respectively, which were lower than the control. This study suggested that the  
792 combined treatment was more effective in maintaining the quality of Chinese bayberries  
793 than heat treatment or nano-packaging alone.

794

795 Li and others (2009) studied the effect of active packaging produced by blending nano  
796 powders of nano-Ag, nano-TiO<sub>2</sub>, and kaolin with polyethylene for the preservation of  
797 Chinese jujube. The active packaging improved the physicochemical and sensory quality of  
798 the product compared to polyethylene without nano-powder (control). Application of active  
799 packaging significantly reduced fruit softening, weight loss, browning, and climactic  
800 evolution during 12 days of storage. An important index of rate of browning of the product  
801 was reduced from 0.7 to a lower level of 0.6 on day 12. The ethylene production rate  
802 increased initially and then declined for all treatments. The maximum ethylene content was  
803 reported as 17.6 μL/kg h for the control on the third day and 9.2 μL/ kg h for nano-packaging  
804 on the sixth day of storage. The active packaging is recommended for Chinese jujube to  
805 improve quality, however, a specific shelf life was not indicated.

806

807 Hu and others (2011) studied the effect of PE blended with nano-Ag, nano-TiO<sub>2</sub>, and  
808 montmorillonite on the quality of ethylene-treated mature kiwifruit at 4 °C for 42 days.  
809 Weight loss, softening, color variation, and Brix degrees (°Brix) of kiwifruit were significantly  
810 reduced by 22.7%, 124.8%, 23.5% and 14.4%, respectively. Ethylene concentrations in the  
811 headspace were 39.5 μL/L and 16.8 μL/L for the control and nano-packaging, respectively,

812 on storage day 42. For kiwifruit, 30  $\mu\text{L/L}$  was reported to cause unacceptable softening. The  
813 researchers stated that nanocomposite packaging was effective in inhibiting ethylene  
814 production (57.4% of lower headspace ethylene in active packaging), preventing  
815 physiological changes, and delaying ripening, however, no specific shelf life was indicated. A  
816 lower level of ethylene production was related to the synergistic effect of nanoparticles  
817 which decompose or oxidize ethylene into water and carbon dioxide (Li and others 2009).  
818 Li and others (2011) tested poly(vinyl chloride) (PVC) film coated with nano-ZnO powder on  
819 fresh-cut 'Fuji' apple at 4 °C for 12 days. Nano-coated PVC film reduced fruit decay rate and  
820 enzyme activity, retarded ethylene production, maintained °Brix and titratable acidity  
821 compared to uncoated PVC (control). Maximum ethylene content was reported as 40  $\mu\text{L/kg}$   
822 day for nano-packaging on the ninth day and, 70  $\mu\text{L/kg}$  day for the control on the sixth day of  
823 storage. The browning index was significantly reduced from 31.7 to 23.9 on day 12,  
824 maintaining the initial appearance. The activity of polyphenol oxidase was 9.6 U/g min in  
825 active packaging and 21.5 U/g min in normal packaging on day 9. The authors reported that  
826 the nano-packages had more oxygen and less carbon dioxide in the headspace compared to  
827 the control, indicating the lower respiration in the nano-coated PVC. Nano ZnO is reported  
828 to have similar physical properties to  $\text{TiO}_2$  which oxidizes ethylene into water and carbon  
829 dioxide under UV irradiation (Han and Nie 2004).

830

831 **Zeolite-based minerals.** Zeolite-based ethylene absorbers are good candidates for  
832 commercial use. The most characteristic property of zeolites is their porous three-  
833 dimensional structure with cation exchange, adsorption, and molecular sieving properties.  
834 Therefore, zeolites have been used in many industrial and agricultural applications, including

835 as an ethylene-absorbing additive incorporated into packaging films. There are various  
836 reports that incorporation of zeolites increases gas permeability of packaging films by means  
837 of their crystalline porous-three-dimensional framework structure (Süer and others 1994;  
838 Kittur and others 2005; Zhao and others 2011). Esturk and others (2014) successfully applied  
839 low-density polyethylene (LDPE) bags with ethylene absorber (8% Tazetut® master batch, an  
840 inorganic product containing 50% of various alumino-silicate minerals (zeolite)) to broccoli  
841 florets under passive modified atmosphere and stored at 4 °C for 20 days. The authors  
842 stated that spoilage occurred quickly in unpackaged broccoli (control) illustrated by  
843 chlorophyll degradation, stem-hardening, and mass loss of 41.5% on day 20, which was less  
844 than 1% for packaged applications. The product (control) was unacceptable for the sensory  
845 panel after 5 days. However, the quality loss was significantly reduced in active LDPE bags  
846 with an ethylene absorber. Ethylene concentration was 61.8 ppm in the control LDPE and  
847 0.33 ppm in active LDPE at the end of the storage. Thus, packaging with zeolite-based active  
848 films extended the shelf life of broccoli up to 20 days, compared to a 5-day shelf-life for the  
849 unpackaged product.

850

851 The quality of kiwifruit packaged with HDPE bags including a sachet with  $\text{KMnO}_4$   
852 impregnated zeolites at 4 °C for 31 days was reported by Küçük (2006). 0.2 mL  $\text{KMnO}_4/\text{g}$   
853 zeolite was impregnated with zeolites of 1-3 mm in size.  $\text{KMnO}_4$  impregnated zeolites were  
854 added into the HDPE bags as 5 and 10% of the amount of kiwifruit. Fruits were firmer and  
855 had a higher vitamin C content in zeolite-containing HDPE films compared to the control  
856 (60.67 mg/100 mL of vitamin C for fruits packaged using HDPE with 5% zeolite and 47.37  
857 mg/100 mL for the control on day 31). There was no significant difference reported for color

858 values  $L^*$ ,  $a^*$ , and  $b^*$  ( $L^*$  indicating lightness,  $a^*$  chromacity on a green (-) to red (+) axis, and  
859  $b^*$  chromacity on a blue (-) to yellow (+) axis) between HDPE bags with and without zeolite.  
860 The ethylene measurements for each treatment and the shelf life were not published in this  
861 publication.

862

863 Boonruang and others (2012) tested 4 different packaging films for mango at 12 °C. The  
864 tested materials were non-perforated, highly gas-permeable film, non-perforated highly gas-  
865 permeable film with ethylene-absorbing property, micro-perforated highly gas-permeable  
866 film and common low-density polyethylene film. The non-perforated film with ethylene  
867 absorber extended the shelf life of mango to 40 days at 12 °C, compared to 35 days with the  
868 non-perforated, highly gas-permeable film, 30 days with the microperforated film , and 5  
869 days with the common low-density polyethylene. The film with ethylene absorber reduced  
870 weight loss, maintained firmness, and there was no sign of decay during storage. Low  
871 ethylene concentrations ( $<3.5 \mu\text{L/L}$ ) were reported in mangoes with the different packaging  
872 films. Ethylene production in the microperforated packages was higher than that in the non-  
873 perforated and non-perforated with ethylene absorber packaging. The ethylene-absorbing  
874 characteristics of the material delayed the ripening process of mangoes. A further study  
875 shows that zeolite-added LDPE bags were applied to ripe kiwifruits successfully establishing  
876 an equilibrium atmosphere in the headspace in 5 days, however, the control material with  
877 no ethylene absorber did not reach equilibrium (steady state oxygen and carbon dioxide)  
878 during 20 days of cold storage at 4 °C. A minimum shelf life of 20 days was suggested for  
879 kiwifruits using zeolite-incorporated LDPE bags. Ethylene measurement is not reported in  
880 this study (Ayhan 2016).

881

882 Jacobsson and others (2004) tested 4 different materials on fresh broccoli at 2 storage  
883 temperatures (4 and 10 °C). Among the materials tested, one material was LDPE-based with  
884 pouches containing a commercial sachet (Ryan Instruments, The Netherlands) to absorb  
885 ethylene, and the other material was commercial LDPE film impregnated with a natural  
886 hydroscopic mineral produced by PEAKfresh® (USA). Results showed that LDPE pouches with  
887 a sachet provided 11 days and LDPE incorporated with ethylene absorber provided 12 days  
888 of shelf-life at 4 °C. But at 10 °C, the commercial LDPE bags incorporated with ethylene  
889 absorber provided the longest shelf-life of 9 days compared to 6 days for sachet application  
890 for broccoli. LDPE film impregnated with a natural hydroscopic mineral resulted in a lower  
891 loss in weight, better color and texture, while the chlorophyll content was maintained. This  
892 publication does not report any ethylene measurements.

893

894 Commercially-available ethylene-scavenging films in the market are mostly zeolite-based  
895 such as Evert-Fresh® (Evert-Fresh Corporation, USA), PEAKfresh® (PEAKfresh®, USA),  
896 Profresh® (E-I-A Warenhandels GmbH, Austria) and Bio-Fresh™ (Grofit Plastics, Israel). The  
897 main limitation of these films is opacity and, thus, these plastics are mostly colored  
898 (Martínez-Romero and others 2007; Ayhan 2013). PEAKfresh® is a polyethylene bag  
899 impregnated with minerals to absorb ethylene and moisture. Evert-Fresh® absorbs ethylene,  
900 ammonia and carbon dioxide. An additive made with low-density polyethylene (LDPE) to  
901 absorb ethylene, ethanol, ethyl acetate, ammonia, and hydrogen sulfide is incorporated in  
902 Profresh®. Bio-Fresh™ is a film used in combination with modified atmosphere packaging to  
903 absorb various substances arising from the ripening process. Active and intelligent systems

904 are governed in Europe by regulations (EC) No 1935/2004 and 450/2009 (European  
905 Commission 2004, 2009). The use of permanganate as active agent in contact with food is  
906 not permitted in Europe (Pereira de Abreu and others, 2012).

907

908 Packaging materials integrated with ethylene-removers in the packaging structure are still  
909 limited in commercial applications. The use of these materials for a broad spectrum of fresh  
910 products is also limited compared with other active packaging applications reported in the  
911 literature. The main principle for the successful packaging of fresh and fresh-cut produce is  
912 that the gas permeability (oxygen and carbon dioxide) of the packaging film and respiration  
913 rate of the produce should correspond allowing gas equilibrium in the headspace, as well as  
914 removal of ethylene from the package environment (Ayhan 2013). The application of  
915 adequate oxygen and low carbon dioxide-modified atmosphere packaging combined with  
916 ethylene absorber could provide further benefits to control the product metabolism and  
917 increase the shelf life of fruits and vegetables compared to the application of solely MAP.  
918 However, it should be noted that packaging parameters should be designed to be produce-  
919 specific, since each produce varies in respiration rate, ethylene production rate, and  
920 ethylene sensitivity, and hence the requirements for packaging and storage vary.

921

## 922 **Antioxidant Releasers**

923 There has been increased activity in the development of antioxidant-releasing packaging for  
924 food applications during recent years. Synthetic antioxidants, such as butylated  
925 hydroxytoluene (BHT) and butylated hydroxyanisole (BHA) have been widely used in food  
926 packaging to prevent lipid oxidation. There is now growing interest in the inclusion of natural

927 antioxidants such as polyphenols, tocopherols, plant extracts, and essential oils to active  
928 packaging materials (Nerín and others 2006; Park and others 2012; Barbosa-Pereira and  
929 others 2014; Marcos and others 2014). Some recent developments in this field have been  
930 summarized in Table 5.

931

932 Torres-Arreola and others (2007) reported a delay in lipid oxidation and protein  
933 denaturation in fresh sierra fish fillets through the incorporation of BHT into LDPE packaging.  
934 Compared to LDPE films, samples packed in BHT-LDPE films demonstrated lower lipid  
935 oxidation, expressed as thiobarbituric acid-reactive substances (TBARS) values ( $4.20 \pm 0.52$   
936 vs  $11.95 \pm 1.06$  mg malonaldehyde (MDA)/kg), peroxide index values ( $7.20 \pm 1.38$  vs  $15.15$   
937  $\pm 1.48$  meq/kg), and free fatty acid contents ( $7.98 \pm 0.43$  vs  $11.83 \pm 1.26$  of oleic acid). Fillets  
938 packed in BHT-LDPE packaging films also inhibited less tissue damage and better retained  
939 firmness than fillets packed in LDPE. However, the presence of synthetic antioxidants in food  
940 is being questioned, therefore, the alternative approach that is being widely studied is the  
941 use of natural antioxidants such as  $\alpha$ -tocopherol. Poly(lactide-co-glycolide) films loaded with  
942 2%  $\alpha$ -tocopherol, or a combination of 1% BHT and 1% BHA, were used to evaluate the  
943 stability of dry whole milk and dry buttermilk (Van Aardt and others 2007). BHT and BHA  
944 have a higher volatility than  $\alpha$ -tocopherol, therefore it was expected that they would be  
945 more suitable for dry food applications. However, in this study,  $\alpha$ -tocopherol offered the  
946 same antioxidant protection for whole milk powder exposed to light and oxygen. In a further  
947 study, sealable multilayer films (HDPE/EVOH/LDPE) manufactured with an inner LDPE layer  
948 containing 4% of  $\alpha$ -tocopherol delayed the lipid oxidation of whole milk powder at 30 and 40  
949 °C (Granda-Restrepo and others 2009b). Similarly, sealable LDPE films containing 1.9 and 3%

950 of  $\alpha$ -tocopherol maintained the oxidation stability (hexanal content) of corn oil for 16 weeks  
951 at 30 °C, compared to 12 weeks for the oil in a control bag without antioxidant (Graciano-  
952 Verdugo and others 2010). Manzanarez-López and others (2011) reported that poly (lactic  
953 acid) films containing 2.58% of  $\alpha$ -tocopherol were also able to delay the induction of the  
954 oxidation, measured as peroxide value, of soybean oil at 20°C (max. values of 9.9 vs 19.5  
955 meq/kg), 30°C (max. values <10 vs 27.5 meq/kg), and 40°C (max. values of 13.5 vs 33.9  
956 meq/kg). Meanwhile, Torrieri and others (2011) observed that with the combined use of  
957 MAP and LDPE-embedded  $\alpha$ -tocopherol packaging, lipid and fat oxidation in fresh bluefin  
958 tuna fillets could be reduced. Several natural antioxidant products (TOCOBIOL®-PV,  
959 NUTRABIOL®-T90, NUTRABIOL®-T50 PV) containing tocopherols incorporated into LDPE films  
960 inhibited lipid oxidation of salmon muscle up to 40% during storage, hence being suitable for  
961 use in extending the shelf life of salmon (Barbosa-Pereira and others 2013).

962

963 Active antioxidant food packaging films produced by incorporating 4.6% of the natural  
964 flavonoid quercetin into EVOH matrix showed enhanced lipid oxidative stability, as  
965 demonstrated by a lower peroxide index (12 vs 27 meq/kg) and a reduction in TBARS values  
966 by 25% during storage time (López-de-Dicastillo and others 2012b). Catechin, an active  
967 component of green tea with properties similar to those of quercetin, was shown to be an  
968 effective antioxidant ingredient to retard the oxidation of sunflower oil and fried peanuts  
969 (López-de-Dicastillo and others 2012a). Fried peanuts stored in sealed bags manufactured  
970 with active films containing 0.33 and 1.34% of catechin at 37 °C for 40 days resulted in a  
971 strong reduction in hexanal content released into the headspace up to 25 days, after which  
972 the hexanal increased at the same rate as in the control sample. Additionally, the peroxide

973 index was used to monitor the effect of the films on the oxidation of sunflower oil over 5  
974 months. On exposing sunflower oil to the films, the peroxide values demonstrated that the  
975 films actively protected the oil. Moreover, the films with quercetin (0.76 and 4.01%) were  
976 more effective compared to those with catechin due to the higher solubility of quercetin in  
977 this product, as well as its higher antioxidant capacity. The reported results using accelerated  
978 shelf life testing (37 °C) to estimate the antioxidant potential of the films. However, the  
979 antioxidant performance under real storage conditions needs to be evaluated before  
980 commercial implementation. Another issue to be addressed would be the inclusion of EVOH  
981 in a multilayer system in order to protect the system from water and to reduce costs, while  
982 maintaining antioxidant activity. Other nonvolatile antioxidants such as ascorbic, ferulic, and  
983 citric acids incorporated into polymers such as sealable EVOH and sealable cornstarch/linear  
984 LDPE films demonstrated antioxidant activity when in contact with brined sardines and  
985 ground beef (López-de-Dicastillo and others 2012b; Júnior and others 2015).

986

987 Antioxidant packaging systems containing volatile extracts, essential oils, or active  
988 components of plants or spices have been developed to improve quality and to extend the  
989 shelf life of various food products. Thymol, carvacrol, and eugenol incorporated into sealable  
990 corn-zein-laminated linear LLDPE films were used for fresh ground beef packaging and  
991 effectively inhibited lipid oxidation and had a positive effect on the stability of beef patties  
992 during storage (Park and others 2012). The active linear LDPE composite film containing 1  
993 wt% of the phenolic antioxidant resveratrol, which is naturally produced by plants under  
994 stress conditions, showed strong antioxidant activity, reduced lipid oxidation by 34.7%, and  
995 extended the shelf life of fresh meat stored at 4 °C by a few days (Busolo and Lagaron 2015).

996

997 Antioxidant films obtained from biomaterials containing green tea extract have been  
998 demonstrated to improve oxidative stability of pork meat products (Siripatrawan and Noipha  
999 2012; Yang and others 2016). Yang and others (2016) reported a stronger antioxidant  
1000 capacity of films obtained with 0.5% of green tea extract compared to those obtained with  
1001 oolong and black tea extracts. Specifically, at the end of 10 days storage of pork meat, the  
1002 TBARS value of the control sample was 1.64 mg MDA/kg, whereas the TBARS values of the  
1003 samples wrapped with film containing green tea extract, oolong tea extract, and black tea  
1004 extract were 0.93, 1.16, and 1.27 mg MDA/kg sample, respectively. In another study,  
1005 Lorenzo and others (2014) reported that multilayer barrier films containing oregano  
1006 essential oil (2%) were more effective in preventing lipid oxidation of foal meat packed in  
1007 MAP (80:20, O<sub>2</sub>:CO<sub>2</sub>) than those with green tea extract (1%). An interesting investigation by  
1008 Carrizo and others (2016) reported radical-scavenging capacity of sealable multilayer films,  
1009 in which green tea extract was added to the laminating adhesive and thus not in direct  
1010 contact with the packaged food (peanuts and cereals covered with chocolate). According to  
1011 the authors, this packaging system was able to protect food against oxidation during a long-  
1012 term period of 16 months. It is important to highlight the industrial relevance of this study,  
1013 since the use of commercial packaging materials would facilitate industrial implementation  
1014 of this technology.

1015

1016 Other authors have explored the application of packaging materials containing rosemary and  
1017 oregano extracts in direct contact with muscle foods as active packaging systems.  
1018 Antioxidant films containing rosemary and oregano extracts showed improved oxidative

1019 stability of lamb and beef meat (Nerín and others 2006; Camo and others 2008, 2011).  
1020 Oregano extracts included in the films were more efficient in preventing oxidation of lamb  
1021 meat than rosemary extracts, they extended fresh odor and color from 8 to 13 days  
1022 compared to the control (Camo and others 2008). Some authors have studied the impact of  
1023 antioxidant packaging in preventing high-pressure-induced lipid oxidation. Bolumar and  
1024 others (2011, 2016) developed LDPE films coated with rosemary extract that were able to  
1025 protect meat patties from high-pressure-processing-induced lipid oxidation and  
1026 consequently extend the shelf life. More specifically, the lipid oxidation of chicken breast  
1027 patties submitted to high-pressure treatment and stored at 5 °C was higher in the surface  
1028 part of samples and the active packaging delayed oxidation it up to 25 days demonstrated by  
1029 lower peroxide values ( $7.2\pm 1.38$  vs  $15.15\pm 1.48$  meq/kg), FFA ( $7.98\pm 0.43$  vs  $11.83\pm 1.26\%$  oleic  
1030 acid), and TBARS ( $4.20\pm 0.52$  vs  $11.95\pm 1.06$  mg MDA/kg) (Bolumar and others 2011).

1031

1032 The use of by-products from the food industry as a source of antioxidants for food packaging  
1033 has also been explored as a means of providing added value to these residues. Packaging of  
1034 beef with LDPE film coated with a brewery residual waste extract was able to reduce lipid  
1035 oxidation by up to 80% during cold storage (Barbosa-Pereira and others 2014). Barley husk,  
1036 another waste product obtained from the brewery industry, also proved to be effective in  
1037 slowing down lipid hydrolysis and improving the oxidative stability in blue shark muscle  
1038 (Pereira de Abreu and others 2011). Meanwhile, anthocyanins from wine grape pomace,  
1039 beet root residue powder, and mango and acerola pulp incorporated into sealable  
1040 biodegradable films had a protective effect on sunflower and palm oil oxidation (Souza and  
1041 others 2011; Oliveira and others 2016; Stoll and others 2016). For example, a sunflower oil

1042 control sample directly exposed to the air and light reached a peroxide index of 65.8 meq/kg  
1043 after 3 days, while the samples stored in cassava starch film bags prepared with  
1044 encapsulated anthocyanins presented lower values (4.7-28.7 meq/kg) (Stoll and others  
1045 2016). Similarly, a lower peroxide index, which was significantly different from that of the  
1046 control (oil with no packaging), was detected in palm oil packed in cassava starch films with  
1047 high concentrations of mango and acerola pulp additives (Souza and others 2011). However,  
1048 it was found that vitamin C in acerola pulp acted as a pro-oxidant agent, which suggests that  
1049 the use of components rich in vitamin C should be avoided.

1050

1051 Extensive research on the use of antioxidant packaging systems to prevent food oxidation  
1052 has been conducted. However, most of the reported studies fail to validate the efficiency of  
1053 the antioxidant packaging systems in real commercial food applications and do not consider  
1054 their target market and consequent legal status. Therefore, to favor the industrial  
1055 implementation of this technology, it is essential to study real food packaging systems.  
1056 Research efforts should focus on the use of packaging materials obtained through scalable  
1057 film processing techniques (such as extrusion or coating vs solvent-casting), packaging  
1058 materials with suitable barrier properties and formats for the studied food product,  
1059 industrial packaging techniques (such as MAP or vacuum vs wrapping), effect on sensory  
1060 properties of food, and validation using real storage conditions.

1061

## 1062 **Carbon Dioxide Emitters**

1063 The antimicrobial effect of CO<sub>2</sub> is thoroughly documented in the literature (Kolbe 1882;  
1064 Valley 1928; Haas and others 1989; Debs-Louka and others 1999). CO<sub>2</sub> is soluble in the

1065 aqueous and fatty phases of food products and the antimicrobial effect is highly dependent  
1066 on the rate of solubility and amount of CO<sub>2</sub> dissolved in the food product. The solubility of  
1067 carbon dioxide increases with decreasing temperature (Devlieghere and others 1998;  
1068 Devlieghere and Debevere 2000) and also varies for different food products depending on  
1069 the properties of the food such as surface area, pH and composition (water, fat, protein)  
1070 (Chaix and others 2015). The antimicrobial effect has been found to be proportional to the  
1071 partial pressure of the gas (Blickstad and others 1981). In terms of food packaging, this  
1072 implies that the total amount of CO<sub>2</sub> present in the headspace of the package is crucial for  
1073 the effect. The concept of a CO<sub>2</sub>-releasing device to be implemented in modified  
1074 atmosphere packages (MAP) to maintain high headspace levels of CO<sub>2</sub> during storage, and  
1075 thereby, facilitate smaller package volumes (lower gas to product (g/p) volume ratio), and  
1076 prolonged shelf life was introduced in the 1990s.

1077

1078 The implementation of CO<sub>2</sub> emitters in MA-packages may allow for increased filling degree,  
1079 reduced package sizes, improved transport efficiency, and a net reduction in environmental  
1080 impact. The release of carbon dioxide from a tuned emitter system may also prevent  
1081 packaging deformation as it compensates for CO<sub>2</sub> absorption into the food product in the  
1082 initial stages of storage. In this way it counteracts formation of negative pressure in MA-  
1083 packages that increase the drip loss of the product, which may give the packages an  
1084 unattractive appearance to the consumer (Holck and others 2014). Further, inhibition of  
1085 growth of spoilage bacteria and prolonged shelf life for fresh food products at sustained high  
1086 CO<sub>2</sub> levels in the packages will have a knock-on effect in the form of a reduction in food  
1087 waste, an issue gaining increasing attention and priority in western parts of the world.

1088

1089 The emitters usually come in the form of a pad or sachet, in many cases as a combined liquid  
1090 absorber. The active ingredients inside the absorbent pad react when the pad absorbs liquid  
1091 that is seeping out of the product, resulting in the release of CO<sub>2</sub>. Over the last decade, the  
1092 field of CO<sub>2</sub> emitters has advanced significantly, reflected in increased research activity and  
1093 sale of commercial CO<sub>2</sub> emitters. Table 6 lists the CO<sub>2</sub> releaser technologies available for  
1094 food preservation to date, as well as their applications and benefits to specific food  
1095 products. In the literature, there are several reports of the use of ferrous carbonate in  
1096 carbon dioxide emitters (Rooney 1995; Sivertsvik 2003; Restuccia and others 2010).  
1097 However, documentation and descriptions of the technology principle, benefits, and food  
1098 applications are scarce. Other concepts have also emerged, such as the technology applied  
1099 in the emitter Verifrais™ (SARL Codimer, Paris, France). The active ingredients in the  
1100 Verifrais® system are sodium bicarbonate and ascorbic acid (Rooney 1995; Kerry 2014).  
1101 There are also examples of (commercial) combined O<sub>2</sub> scavengers and CO<sub>2</sub> emitters on the  
1102 market, such as Ageless® G (Mitsubishi Gas Chemical Co., Japan) and FreshPax® M  
1103 (Multisorb Technologies Inc, USA). These systems are based on either ferrous carbonate or a  
1104 mixture of ascorbic acid and sodium bicarbonate (Coma 2008).

1105

1106 One of today's most well documented CO<sub>2</sub>-releasing systems is based on a combination of  
1107 the active substances sodium bicarbonate and citric acid. Several scientific publications  
1108 document the effect of this CO<sub>2</sub>-releasing system. Amongst the first reports in the literature  
1109 of such applications is a study from 1995 (Bjerkeng and others 1995). In this study the effect  
1110 of CO<sub>2</sub>-emitter (non-commercial) on, for example, the microbial and sensory shelf life of cod

1111 fillets in MA-packages (70% CO<sub>2</sub>, 30% N<sub>2</sub>) and vacuum was investigated. The CO<sub>2</sub> level in the  
1112 MA-packages (g/p ratio not given) with emitter decreased to approximately 40% after a few  
1113 days of storage and thereafter increased and fluctuated around 50% CO<sub>2</sub> for the remaining  
1114 storage time. For the vacuum-packages with emitter, the CO<sub>2</sub> level increased to 50% after  
1115 one day of storage and reached 80% at day 15 (very limited headspace volume available).  
1116 For the control (MAP and liquid absorber) the CO<sub>2</sub> level rapidly dropped to 20% after one  
1117 day of storage. The cod fillets packaged in modified atmosphere and vacuum with emitter  
1118 were found to have a sensory shelf life, based on ammonia-like odor, of 11 days, compared  
1119 to 7 days for the control (MA-packaged with liquid absorber), supported by measured  
1120 trimethylamine (TMA) levels and microbial analyses (total viable counts (TVC) and H<sub>2</sub>S-  
1121 producing bacteria).

1122

1123 In a study by Hansen and others (2007), the effect of the same emitter system was  
1124 investigated, also in the packaging of cod fillets, looking at the simultaneous effect of high  
1125 CO<sub>2</sub> and O<sub>2</sub> level and different g/p ratio. Packaging in modified atmosphere (60% CO<sub>2</sub>, 40%  
1126 O<sub>2</sub>) with CO<sub>2</sub> emitter at a low g/p ratio (1.3/1.0) resulted in extension of shelf life (14 – 21  
1127 days total) both in terms of sensory properties (odor and appearance assessment) and  
1128 bacterial growth when compared to vacuum-packaging (7 – 14 days total shelf life). The  
1129 shelf-life obtained with the emitter was, however, comparable to that of cod packaged in  
1130 MAP at a high g/p ratio (3.9/1.0) without emitter. For the MA-packages (both g/p 1.3/1.0  
1131 with emitter and g/p 3.9/1.0 without emitter), the dominating bacteria at the end of the  
1132 storage time were different species of *Carnobacterium* and some *Photobacterium*. In the  
1133 MA-packages with a g/p ratio of 1.3/1.0 including an emitter, the level of headspace CO<sub>2</sub>

1134 increased to about 70 – 80% during the storage period, effectively compensating for the CO<sub>2</sub>  
1135 absorbed by the product, whilst for MA-packages with a g/p of 3.9/1.0 without emitter, the  
1136 CO<sub>2</sub> level dropped to 35-40% during the storage time. In a recently published study (Hansen  
1137 and others 2016), cod fillets in vacuum-packages with a CO<sub>2</sub> emitter displayed a shelf-life  
1138 extension of 2 days (9 days total) compared to vacuum-packages with a regular liquid  
1139 absorber. However, the longest shelf life (13 days) was obtained for the combination MAP  
1140 (60% CO<sub>2</sub> and 40% N<sub>2</sub>) and CO<sub>2</sub> emitter, based on sensory (odor and appearance assessment)  
1141 and microbiological evaluation (TVC, H<sub>2</sub>S-producing bacteria counts, lactic acid bacteria  
1142 counts (LAB), and microbiota analysis). With a CO<sub>2</sub> emitter present in the MA-packages, the  
1143 headspace level of CO<sub>2</sub> (g/p 1.6/1) was kept stable at about 35 – 37% once the CO<sub>2</sub>  
1144 absorption into the product had reached equilibrium (after 1 day of storage). For the MA-  
1145 packages (g/p 1.6/1) without emitter, the CO<sub>2</sub> level dropped to 26 – 27% after equilibrium.  
1146 Distinct differences in TVC were measured for the different packaging methods; after 15  
1147 days of storage the cod fillets in vacuum had a TVC of log 7.1 cfu/g, while the number for the  
1148 cod in MA-packages and MAP with emitter was log 6.4 cfu/g and log 5.5 cfu/g, respectively.  
1149 In the studies described above, the laboratory-type emitters were custom-made; the ratio  
1150 between citric acid and sodium bicarbonate was adjusted to the pH of the food product.

1151

1152 The impact of the combined CO<sub>2</sub> emitter and liquid absorber (laboratory-type, custom-  
1153 made) on the quality and shelf life of fresh salmon has been thoroughly documented  
1154 (Hansen and others 2009a, b, c). Hansen and others (2009a) demonstrated the effectiveness  
1155 of a laboratory-type emitter in reducing the g/p ratio for MA-packages (60% CO<sub>2</sub>, 40% N<sub>2</sub>) of  
1156 salmon fillets without compromising the shelf life (based on microbial, sensory, and textural

1157 analysis). For the MA-packages (g/p 3/1) without emitter, the CO<sub>2</sub> level dropped to 40% 4  
1158 days after packaging and then stabilized. For the MA-packages (g/p 1/1) with emitter, the  
1159 CO<sub>2</sub> level displayed an initial drop to about 45% (day 1), but subsequently the level increased  
1160 and reached 65 – 70% during the storage time. The measured TVC levels for the salmon  
1161 packaged in MAP with and without emitter were comparable and the obtained shelf life for  
1162 the two packaging methods was the same. The TVC of the MA-packaged samples (with and  
1163 without emitter) reached a level of log 5 – 6 cfu/g after 15 days of storage, while for the  
1164 vacuum-packaged salmon, the same bacterial counts were measured 7 – 10 days into  
1165 storage. The results illustrate that a CO<sub>2</sub> emitter can allow for a more sustainable packaging  
1166 of fresh fish products with a significant reduction in package sizes and hence amount of  
1167 packaging material, since a comparable shelf life can be obtained at significantly reduced g/p  
1168 ratio.

1169

1170 The emitter system (laboratory-type) has also been studied for different meats. In a study by  
1171 Pettersen and others (2014), the effect of different packaging methods was evaluated for  
1172 fresh reindeer meat. The study documented prolonged sensory shelf life (odor evaluation)  
1173 for meat packaged in modified atmosphere (60% CO<sub>2</sub>, 40% N<sub>2</sub>) with and without CO<sub>2</sub> emitter  
1174 of 21 days, compared to 17 days for vacuum-packaged meat. For reindeer meat packaged in  
1175 MAP with CO<sub>2</sub> emitter (storage temperature 15 °C), lower TVCs were measured after 13 and  
1176 17 days (log 3 – 4 cfu/g) compared to MAP without emitter and vacuum-packaging at the  
1177 same sampling times (log 4 – 5 cfu/g). Samples from the 3 packaging methods reached the  
1178 same level of TVC (log 6 cfu/g) at the end of the storage time (day 21). The CO<sub>2</sub> level in the  
1179 MA-packages with emitter displayed an initial decrease after 1 day of storage to 56%, but

1180 increased to 67% towards the end of the storage time. The packaging strategy with CO<sub>2</sub>  
1181 emitter also resulted in a significantly reduced drip loss for the reindeer meat; 1% for MAP  
1182 with emitter compared to 3% for MAP without emitter. The article concluded that the  
1183 capacity of the custom-made emitter was too low for the product, implying that the  
1184 beneficial effect of an emitter could be expected to be more pronounced.

1185

1186 In a similar study by Holck and others (2014) where the emitter capacity was more carefully  
1187 tuned towards the food product, the emitter compensated well for CO<sub>2</sub> absorbed by chicken  
1188 fillets in MAP (100% CO<sub>2</sub>) and the drip loss was drastically reduced; from a weight loss of  
1189 7.5% for fillets packaged without emitter, to 2.5% for fillets packaged with an emitter in MA-  
1190 packages with the same g/p ratio (2.5). The effect was assumed to be due to a reduction in  
1191 packaging collapse and physical squeeze on the fillet. The emitter maintained the CO<sub>2</sub> level  
1192 close to 100% throughout the storage time, while for the MA-packages without emitter the  
1193 CO<sub>2</sub> level dropped slightly to 90 – 95% depending on the g/p ratio. The microbial shelf life  
1194 was found to be the same for fillets packaged in 100% CO<sub>2</sub> with emitter and in 100% CO<sub>2</sub>  
1195 with regular fluid absorber. For the chicken packaged in 100% CO<sub>2</sub> with emitter, significant  
1196 bacterial growth inhibition was detected; the bacteria required 7 additional days to reach a  
1197 level of 10<sup>7</sup> cfu/cm<sup>2</sup> compared to packaging with commonly applied gas composition of 60%  
1198 CO<sub>2</sub> and 40% N<sub>2</sub>. This study reports that packaging in 100% CO<sub>2</sub> without emitter is not  
1199 possible due to an unacceptably high drip loss.

1200

1201 Trindade and others (2013) evaluated the use of a combined oxygen scavenger and carbon  
1202 dioxide emitter sachet (Didai, technology unknown) for active packaging of lamb cuts. The

1203 study did not draw conclusions on a significantly prolonged shelf life for the product with  
1204 emitter applied in vacuum-packages. The percentage level of CO<sub>2</sub> generated in the vacuum-  
1205 packages including an O<sub>2</sub> scavenger/CO<sub>2</sub> emitter was reported to be 45% CO<sub>2</sub> after about 17  
1206 days of storage. The same CO<sub>2</sub> level was measured in the control vacuum-packages without  
1207 emitter, speculated by the authors to be a result of CO<sub>2</sub> emission by anaerobic  
1208 microorganisms. The CO<sub>2</sub> level is unexpectedly low for vacuum-packages with O<sub>2</sub>  
1209 scavenger/CO<sub>2</sub> emitter, considering the limited available volume and that the O<sub>2</sub> level is  
1210 stated to be zero. No information is provided on the total gas composition and the authors  
1211 do not describe how the headspace gas composition in vacuum-packages was measured.  
1212 Furthermore, it is not stated if the emitter capacity was product-adjusted and the concept of  
1213 the technology is not clarified. In a different study, by Chen and Brody (2013), packaging  
1214 with a CO<sub>2</sub>-releasing packaging structure (CSP Technologies, technology unknown) was  
1215 found to be effective at controlling the proliferation of artificially inoculated *Listeria*  
1216 *monocytogenes*, as well as TVC and Enterobacteriaceae on a vacuum-packaged ready-to-eat  
1217 meat product (cooked ham) stored at different temperatures and monitored over a 4-week  
1218 period. The percentage levels of CO<sub>2</sub> gas in the vacuum-packages were measured as 56%,  
1219 72%, and 69% at 4, 10, and 22 °C, respectively, at the end of the storage period. Inoculation  
1220 of *L. monocytogenes* is not an industrially relevant scenario as the focus is on preventing  
1221 these bacteria from growing on the food product, rather than inhibiting the bacteria already  
1222 present. Furthermore, interpretation of the results for this study is complicated by the lack  
1223 of documented sampling data (bacterial counts) during the storage time. The 2 latter studies  
1224 are not included in Table 6 as the underlying technologies (active substances) of the emitters  
1225 have not been published.

1226

1227 Comparisons of different CO<sub>2</sub> emitter concepts are challenging. Differences in pre-handling  
1228 of the food product, storage conditions and temperature, type of packaging material, g/p  
1229 ratio, package size, and gas composition are all variables that will have an impact on the  
1230 effect that the CO<sub>2</sub> emitter contributes to the shelf life of the food product. Another  
1231 important factor concerning the evaluation of CO<sub>2</sub> emitter that is rarely documented is the  
1232 material and density of the emitter substrate. The type and structure of the substrate in  
1233 which the active ingredients are incorporated is of importance for liquid absorption and the  
1234 amount and rate of CO<sub>2</sub> release. Examples of materials applied in different layers of CO<sub>2</sub>  
1235 emitters/liquid absorbers are cellulosic fiber, or other fiber-based materials, SAP (super-  
1236 absorbent polymer), other hydrogels and perforated plastic films. In addition, the active  
1237 ingredients may be evenly distributed in pores of the substrate material or in a bulk deposit  
1238 in the core of the substrate. These aspects should be taken into consideration when  
1239 comparing the performance of different emitter systems.

1240

1241 The results of earlier scientific studies, as summarized in the previous paragraphs, have  
1242 shown that an optimal effect of CO<sub>2</sub> emitters can only be achieved when the emitter  
1243 capacity is optimized, that is adapted to the physiological properties and weight of the food  
1244 product in question. An emitter with an optimal capacity will ensure an adequate CO<sub>2</sub> level,  
1245 counteract formation of negative pressure within the package, ensure sufficient liquid  
1246 absorption, and extend shelf life. Optimization of the emitter capacity was investigated in a  
1247 study by Hansen and others (2009b) for salmon fillets in MA-packages with different fillet  
1248 sizes and g/p ratios. A model was developed based on the results, making it possible to

1249 calculate the required amounts of sodium bicarbonate and citric acid, based on weight and  
1250 surface area of the salmon fillets, g/p ratio, and tray capacity. Additional research is required  
1251 on this topic focused on product-specific concepts, making CO<sub>2</sub> emitter technologies more  
1252 flexible and suitable for a broader range of food products.

1253

1254 With regard to commercialization, there are different emitters already on the market today.  
1255 Emitters based on sodium bicarbonate and citric acid include CO<sub>2</sub> Freshpads (CO<sub>2</sub>  
1256 Technologies, Urbandale, Iowa, USA)(Kerry 2014), SuperFresh (Vartdal Plastindustri AS,  
1257 Vartdal, Norway), and the Active CO<sub>2</sub> pad (CellComb AB, Säffle, Sweden). In addition,  
1258 UltraZap® XtendaPak (Paper Pak Industries, La Verne, CA, USA), and the CO<sub>2</sub>Pad (McAirlaid's  
1259 GmbH, Steinfurt, Germany) are based on other CO<sub>2</sub>-releasing concepts.

1260

### 1261 **Antimicrobial Packaging Systems**

1262 Antimicrobial food packaging presents a system designed to inhibit the growth of spoilage  
1263 and pathogenic microorganisms. In this review, the most studied antimicrobial food  
1264 packaging systems have been classified according to their active substance/material:  
1265 essential oils (Table 7); enzymes and bacteriocins (Table 8); antimicrobial polymers (Table 9);  
1266 and organic acids, their derivatives and other organic compounds (Table 10). Furthermore,  
1267 antimicrobial nanoparticles are reviewed separately as the nano-size itself either increases  
1268 or enables the antimicrobial activity (Table 11).

1269

### 1270 **Essential Oils (EOs)**

1271 Recent interest in reducing the use of petroleum-based additives as active materials for food  
1272 preservation has led to the application of natural additives both for the benefit of the  
1273 individual as well as for the environment (Alves-Silva and others 2013). Essential oils (EOs)  
1274 are secondary metabolites and play an important role in plant defense, thus, some of them  
1275 possess strong antimicrobial properties. In addition, most of them are classified as GRAS  
1276 (Ruiz-Navajas and others 2013) and, as a result, EOs have been extensively studied as  
1277 additives in bio-based emulsified films and coatings. Many scientific publications are  
1278 connected to the potential interest in this type of active packaging but without any real  
1279 application to food. Some studies, however, have demonstrated the effectiveness of EO-  
1280 enriched packages containing food and these are presented in Table 7.

1281

1282 Cinnamon essential oil (CEO) is among the most studied EOs in active materials. Gherardi  
1283 and others (2016) showed that a multilayer material containing about 18 and 10% of  
1284 cinnamaldehyde as the major compound of the selected EO showed high activity against *E.*  
1285 *coli O157:H7* and *S. cerevisiae*, as the material reduced both microorganisms by 3 log  
1286 CFU/mL. Compared to the results obtained in culture media, *E. coli* showed higher sensitivity  
1287 to active materials in tomato puree. Interestingly, to maintain greater CEO in the film and  
1288 avoid too much loss of volatile substances, an antimicrobial packaging material was  
1289 developed by incorporating a cinnamon essential oil/ $\beta$ -cyclodextrin inclusion complex into  
1290 polylactic acid nanofibers via an electrospinning technique (Wen and others 2016).

1291 Application in the preservation of pork (25 °C) showed that the sample packed with the  
1292 nano-film decayed on the eighth day compared to the third day for the control packed with  
1293 fresh-keeping film. In this study, the initial bacterial load of the pork was  $10^3$ – $10^4$  CFU/g on

1294 the first day and the unpacked control pork reached an excessive number of colonies after 4  
1295 days (above  $1.10^7$  CFU/g). Another option to control the release of major compounds of CEO  
1296 is to reversibly anchor cinnamaldehyde to a polymer, such as chitosan films, via imino-  
1297 covalent bonding (Higueras and others 2015). The antimicrobial properties of chitosan-Schiff  
1298 base films in milk inoculated with *Listeria monocytogenes* led to a growth inhibition for 12  
1299 days under refrigeration conditions.

1300

1301 Carvacrol is another EO regularly used as a bio-based bioactive compound. However, the  
1302 synergistic antimicrobial effect of different EOs on food has been less frequently studied.  
1303 One example is the study by Campos-Requena and others (2015) based on carvacrol and  
1304 thymol, both included in HDPE/modified montmorillonite nanocomposite films. A synergistic  
1305 antimicrobial effect was observed with *Botrytis cinerea*, when the films were applied  
1306 through indirect contact with strawberries. The half maximal inhibitory concentration (IC50)  
1307 of the EOs in the film was reduced from 40.4 mg/g (carvacrol only) to 13.2 mg/g (both EOs  
1308 50:50). Knowing that the major volatile compounds of oregano EO is carvacrol, Rodriguez-  
1309 Garcia and others (2016) evaluated the effect of oregano EO applied within pectin coatings  
1310 on the inhibition of *Alternaria alternata* on tomatoes. The authors showed that 25.9 g/L was  
1311 effective in inhibiting microbial growth.

1312

1313 To enhance safety and shelf life of cooked cured ham, Ruiz-Navajas and others (2015)  
1314 studied 2 Spanish endemic species of thyme, *Thymus piperella* and *Thymus moroderi*. They  
1315 reported that *T. piperella* had a higher effect than *T. moroderi*, probably due to the higher  
1316 concentration of carvacrol in the former (predominant compound, 31,9%). Addition of both

1317 EOs into films (from 1 to 2%) significantly decreased the aerobic mesophilic and lactic acid  
1318 bacteria counts in food samples, with lowest counts for *T. piperella* at 2%. After 21 days, for  
1319 example, the addition of 2% EOs led to a reduction of 0.87 and 0.53 log cycles of aerobic  
1320 mesophilic bacteria compared to the uncoated samples of *T. piperella*- and *T. moroderi*-  
1321 based films, respectively. In another study with thyme EOs, Quesada and others (2016)  
1322 designed an active packaging system for the shelf-life extension of sliced ready-to-eat  
1323 cooked pork during refrigerated storage. Interestingly, the package included an inner surface  
1324 coated with a chitosan film with thyme essential oil (0%, 0.5%, 1%, and 2%) and was not in  
1325 direct contact with the meat to avoid modification of organoleptic properties. The authors  
1326 reported that yeast populations were affected by the presence of thyme EO and the yeast  
1327 counts decreased as a function of the EO dose in the film, especially during the first 21 days  
1328 of storage.

1329

1330 Arfat and others (2015) investigated microbiological and sensory changes of sea bass slices  
1331 wrapped with fish protein/fish gelatin composite films incorporated with basil leaf essential  
1332 oil (BEO) during storage at 4 °C for 12 days. Films were incorporated with 100% BEO (w/w,  
1333 based on protein content). The shelf life was longer for samples wrapped with material  
1334 incorporating BEOs (10-12 days) compared to the control (6 days). *Allium spp.* extract and  
1335 vanillin have also been proposed as bioactive EOs. With the former, Llana-Ruiz-Cabello and  
1336 others (2015) showed an efficiency against molds in lettuce during 7 days of storage (6.5%  
1337 Proallium®). With the latter, Lee and others (2016) demonstrated that crab sticks packed  
1338 with starfish gelatin films containing 0.05% vanillin exhibited antimicrobial activity against *L.*  
1339 *monocytogenes* previously inoculated on the food product.

1340

1341 This brief overview illustrates that essential oil-based packaging has the potential to enhance  
1342 food preservation. However, such packaging have not yet been extensively commercialized.  
1343 Various factors need to be considered, such as the impact of EOs on (1) the organoleptic  
1344 profile of the target food, (2) the physio-chemical properties of the materials, and (3) the  
1345 effectiveness of this packaging system when manufactured under real conditions. In order to  
1346 limit the constraint of their strong odor and taste, EO-based materials can be selectively  
1347 used with compatible foods in terms of flavor. Another option could be the development of  
1348 tasteless, colorless, and odorless EO derivatives (sensory inertness), such as some curcumin  
1349 derivatives (Coma and others 2011; Etxabide and others 2016).

1350

1351 According to the recent scientific publication by Dornic and others (2016), although EOs  
1352 contain compounds naturally produced in the natural environment by higher plants, their  
1353 consumption may nevertheless present a risk to health, given their composition. Indeed,  
1354 their consumption may cause adverse effects when used inappropriately. The recent study  
1355 of Rivaroli and others (2016) showed that higher doses of 3.5 g/animal/day could have a pro-  
1356 oxidant effect in feedlot livestock. As mentioned by Eghbaliferiz and Iranshahi (2016),  
1357 natural antioxidants can act as pro-oxidants, which produce free radicals and cause DNA  
1358 damage and mutagenesis. Consequently, further research is needed to understand the  
1359 potential toxicity of EOs incorporated into packaging materials.

1360 **Enzymes and Bacteriocins**

1361 Incorporation of proteins, particularly enzymes and bacteriocins, into food packaging to control  
1362 spoilage caused by food pathogenic microorganisms has been an area of research for several  
1363 decades (Table 8).

1364 Enzymes can serve as effective antimicrobials in food packaging by being chemically bonded to,  
1365 or physically entrapped in, packaging films. As an antimicrobial enzyme, lysozyme can destroy  
1366 the glycosidic bonds of the Gram-positive bacterial peptidoglycans. Lysozyme incorporated into  
1367 whey protein films (204 mg/g of film) migrated into the food and inhibited the growth of  
1368 *Listeria monocytogenes* to 4.4 log CFU/cm<sup>2</sup>, extending the shelf life of smoked salmon (Min and  
1369 others 2005). Barbiroli and others (2012) reported incorporation of lysozyme and lactoferrin  
1370 into paper containing carboxymethyl cellulose, which allowed non-covalent binding of the  
1371 positively charged proteins to the paper matrix. Tests on thin meat slices laid on paper sheets  
1372 containing either or both antimicrobial proteins indicated that lysozyme was most effective in  
1373 preventing growth of aerobic bacteria in the meat sample, giving almost 1 log cycle reduction  
1374 with respect to the control. Lysozyme is accepted by the U.S. Food and Drug Administration  
1375 (FDA 2001) as an antimicrobial agent in casings for frankfurters, and in Europe the use of  
1376 lysozyme (E1105) falls under Directive 95/2/EC on food additives (European Union 1995).  
1377

1378 Bacteriocins are peptides or small proteins, produced by some species of lactic acid bacteria  
1379 (LAB), which inhibit the growth of food spoilage bacteria, mainly Gram-positive bacteria. The  
1380 bacteriocin nisin has been successfully incorporated into methylcellulose/hydroxypropyl  
1381 methylcellulose coatings (Franklin and others 2004) or PE films (Siragusa and others 1999), and

1382 it has been coated on LDPE films (Mauriello and others 2005; Neetoo and others 2008) or  
1383 paperboard (Lee and others 2004). Subsequently, the effective inhibition of bacterial growth  
1384 was achieved in such foods as hot dogs, beef, milk, cold-smoked salmon, and orange juice. For  
1385 example, packaging films coated with a cellulose derivatives-based solution containing 10,000  
1386 and 7,500 IU/mL nisin significantly decreased *Listeria monocytogenes* populations on the  
1387 surface of hot dogs by greater than 2 log CFU per package after 60 days of refrigerated storage  
1388 (Franklin and others 2004). Similarly, it was established that nisin-coated LDPE films were  
1389 effective in inhibiting the bacterial flora in milk stored at 4 °C for 7 days, and the most  
1390 significant results were observed in raw milk and pasteurized milk with a reduction of 0.9 and  
1391 1.3 log, respectively. Nisin (E234) has been authorized for food preservation in Europe under  
1392 Directive 95/2/EC on food additives (European Union 1995).

1393

1394 The incorporation of nisin with other antimicrobial agents into PE and PE/polyethylene oxide  
1395 films or polyamide coatings effectively inhibited *Brochothrix thermosphacta*, coliform bacteria  
1396 growth and extended the shelf life of beef (Cutter and others 2001; Kim and others 2002) and  
1397 fresh oysters (Kim and others 2002). According to the research of Khan and others (2016), the  
1398 immobilization of nisin and EDTA on the surface of the cellulose nanocrystal/chitosan-films, by  
1399 using genipin as a cross-linking agent, restricted the growth of psychrotrophs, mesophiles and  
1400 *Lactobacillus* spp. in fresh pork loin meats, and increased the microbiological shelf life of the  
1401 meat sample by more than 5 weeks. The films also reduced the counts of *E. coli* and *L.*  
1402 *monocytogenes* in meat samples by 4.4 and 5.7 log CFU/g, respectively, after 35 days of  
1403 storage. Furthermore, through the formation of nisin, citric acid, EDTA, and polyethylene glycol

1404 sorbitan monooleate-coatings on polymeric films of different hydrophobicity (polyvinylchloride,  
1405 nylon or linear LDPE), the shelf life of refrigerated broiler drumsticks was extended by 0.6 to 2.2  
1406 days (Natrajan and Sheldon 2000). In other studies, nisin, in combination with enterocins,  
1407 sakacin, and potassium lactate, was incorporated into interleaves and tested on cooked ham  
1408 and bacterial growth of *L. monocytogenes* (Jofré and others 2007) and *Salmonella* spp. (Jofré  
1409 and others 2008) was successfully inhibited. Other bacteriocins such as enterocins (Marcos and  
1410 others 2007), lactocins (Massani and others 2014), natamycin (De Oliveira and others 2007),  
1411 and pediocin (Santiago-Silva and others 2009) have been incorporated into biopolymer-based  
1412 films or used as coatings on various substrates, and they reduced bacterial (*L. monocytogenes*,  
1413 *Lactobacillus plantarum*, *Listeria innocua*, *Salmonella* spp.) or fungal (*Penicillium roqueforti*)  
1414 growth on cooked ham, Wieners (2.5 log reduction), Gorgonzola cheese, and sliced ham (0.5-2  
1415 log reduction), respectively.

1416

1417 Further research effort is still needed to evaluate the release of enzymes and bacteriocins from  
1418 various films and coatings into packaging, as well as diffusion to the surface of the food.

1419 Moreover, the impact of the packaging on sensory properties of food should be thoroughly  
1420 assessed. To date, only nisin and natamycin have been approved for use as food additives in  
1421 various countries including the USA and the EU. Therefore, legislative issues regarding the use  
1422 of bacteriocins as food preservatives remain the main limitation in their commercial  
1423 exploitation. Nevertheless, the use of enzymes and bacteriocins in combination with other  
1424 preservation techniques can produce synergistic effects in food packaging while maintaining  
1425 the safety and quality of minimally processed and fresh food products.

1426

1427 **Antimicrobial Polymers**

1428 Some polymers like chitosan or  $\epsilon$ -polylysine are inherently antimicrobial and are used in films  
1429 and coatings (Table 9).  $\epsilon$ -Polylysine is a natural antimicrobial polypeptide that is effective  
1430 against Gram-positive and Gram-negative bacteria. However, only a few studies have reported  
1431 on polylysine incorporation into packaging materials. For example, Zinoviadou and others  
1432 (2010) developed  $\epsilon$ -polylysine-containing whey protein films that significantly reduced the  
1433 specific growth rate of total flora and completely inhibited lactic acid bacteria growth in fresh-  
1434 cut beef portions as well as prolonged shelf life.

1435

1436 Chitosan, along with its derivative products (such as chitooligosaccharides), presents  
1437 antimicrobial and antifungal activity against a wide range of target microorganisms, and it has  
1438 also been proven to be beneficial to food packaging. Chitosan has been incorporated as an  
1439 antimicrobial additive into food packaging with synthetic polymers such as LDPE (Park and  
1440 others 2010) and bio-based polymers such as carboxymethylcellulose (Youssef and others 2016)  
1441 or used as a coating on plastic films (Joerger and others 2009). When chitosan-incorporated  
1442 LDPE films were applied on fresh sliced red meats, microorganisms on the meat surface were  
1443 not inhibited but significant extension of red color shelf life was observed in refrigerated  
1444 samples (Park and others 2010). Meanwhile, bio-nanocomposite films containing chitosan had  
1445 an effect on the total bacterial counts, mold and yeast counts, and coliforms in soft white  
1446 cheese during 30 days of storage at 7 °C, and increased its shelf life (Youssef and others 2016).  
1447 Total bacterial counts significantly decreased during storage especially for samples that were

1448 coated with preservative films. Furthermore, the coliform, mold and yeast organisms in soft  
1449 cheese were inhibited by the active films. Moreover, ethylene copolymer film was coated with  
1450 chitosan through attachment of the polymer to the corona-treated surface of the film, and the  
1451 antimicrobial activity of the composite film against *Listeria monocytogenes* Scott A was tested  
1452 on turkey breast and a log reduction of about 1.7 after 10 days and 1.2 after 15 days at 4 °C was  
1453 achieved (Joerger and others 2009).

1454

1455 Ye and others (2008a) determined that chitosan-coated plastic films were not able to control  
1456 the growth of *L. monocytogenes* on ham steaks and, therefore, evaluated the antilisterial  
1457 efficacy of chitosan-coated plastic films incorporating 5 additional GRAS antimicrobials: nisin,  
1458 sodium lactate, sodium acetate, potassium sorbate, and sodium benzoate. The incorporation of  
1459 those antimicrobials into chitosan-coated plastic film retarded or inhibited the growth of *L.*  
1460 *monocytogenes*, while the film containing sodium lactate was the most effective antimicrobial  
1461 film and showed excellent long-term antilisterial effect with the counts of *L. monocytogenes*  
1462 being slightly lower than the initial inoculum. Similarly, the same films inhibited the growth of *L.*  
1463 *monocytogenes* on cold-smoked salmon samples for at least 6 weeks (Ye and others 2008b).  
1464 However, the authors indicated that sensory studies are needed before this technology is  
1465 further developed (Ye and others 2008a).

1466

1467 The antimicrobial activity of chitosan (low molecular weight, 150 kDa, 75-85% deacetylation)  
1468 coating with 5-10% of lauric arginate ester (LAE), 2-20% of sodium lactate, and 0,3-0,6% of  
1469 sorbic acid (alone or in combination) on PLA films was verified using *Listeria innocua* and

1470 *Salmonella typhimurium* (Guo and others 2014). Most effective combinations were 5%  
1471 chitosan/5% LAE/2% sodium lactate/0.3% sorbic acid and 5% chitosan/5% LAE. Both  
1472 combinations reduced *S. typhimurium* to an undetectable level at 0, 24, and 48 h, and  
1473 significantly reduced *L. innocua* (even 6 logs after 48 h). Antimicrobial tests on surface-  
1474 contaminated turkey slices led to the reduction of *L. innocua* growth by 3 log CFU/cm<sup>2</sup> for both  
1475 films. The films also reduced the growth of *L. monocytogenes* on the surface of ready-to-eat  
1476 meat by 2.5-3 log CFU/cm<sup>2</sup> during storage of 3 or 5 weeks at 10 °C. For *S. typhimurium* the  
1477 reduction was 1.5 log CFU/cm<sup>2</sup>.

1478

1479 The effect of chitosan used in combination with nisin, potassium sorbate, or silver-substituted  
1480 zeolite incorporated into LDPE on the physicochemical and microbial quality of chicken  
1481 drumsticks stored at 5 °C for 6 days was also investigated (Soysal and others 2015). Total  
1482 aerobic mesophilic bacteria counts of samples packed in bags containing 2% of chitosan, nisin,  
1483 zeolite, and potassium sorbate in LDPE layer were 1.03, 0.98, 0.51, and 0.17 times lower,  
1484 respectively, than those of samples packed in control bags. Moreover, samples packed in active  
1485 bags had lower TBARS values than those of samples in control bags. The exploitation of GRAS  
1486 antimicrobials nisin and potassium sorbate in food packaging is straightforward, whereas the  
1487 use of silver zeolite as a surface biocide is debatable. Although it is approved by the US FDA as a  
1488 food contact substance, in the EU it is not included in the list of authorized substances, but is in  
1489 the provisional list for use in accordance with national law. In another study, chitosan films  
1490 were developed by incorporating lauric arginate ester (LAE) and their antimicrobial activity  
1491 against mesophiles, psychrophiles, *Pseudomonas spp.*, coliforms, lactic acid bacteria, hydrogen

1492 sulfide-producing bacteria, yeast and fungi was evaluated on chicken breast fillets at 2, 6, and 8  
1493 days (Higuera and others 2013). Chitosan films demonstrated antimicrobial activity in the  
1494 range of 0.47-2.96 log reduction, dependent on time and bacterial group studied, while the  
1495 incorporation of 5% LAE in the film increased antimicrobial activity to 1.78-5.81 log reduction.

1496

1497 It should be noted that chitosan has been given GRAS status by the U.S. FDA (FDA 2002, 2005,  
1498 2011) for agricultural and medicinal purposes, but it is not yet specifically approved as an  
1499 antimicrobial food additive. Meanwhile, the other antimicrobial polymer mentioned above,  
1500 polylysine, was granted GRAS status by the U.S. FDA in 2004 (FDA 2004). Along with excellent  
1501 antimicrobial properties, packaging coatings and films prepared from such biopolymers exhibit  
1502 a variety of other advantages, such as biodegradability, edibility, nontoxicity, biocompatibility,  
1503 an aesthetic appearance, and good barrier properties. However, further studies are needed to  
1504 fully evaluate industrial feasibility and the commercial viability of implementation of the  
1505 proposed technologies. Furthermore, there is a need to evaluate the packaging effects on the  
1506 sensory properties of food as well as to validate already developed packaging by using  
1507 commercial food products held under real storage conditions.

1508

### 1509 **Organic Acids, their Derivatives and other Organic Compounds**

1510 Some organic compounds such as selected organic acids and their derivatives, exhibit  
1511 antimicrobial activity (Table 10) and can be incorporated into packaging films.

1512

1513 **Citric Acid.** Júnior and others (2015) investigated the antimicrobial activity of citric acid on  
1514 packaged minced beef. 30% of a mixture of citric acid/cornstarch/glycerol (ratio: 1.5:68.5:30)  
1515 was incorporated in extruded cornstarch/LLDPE films. Although the microbial population  
1516 increased in all the samples, less growth was observed in minced beef packed with active films  
1517 compared to the control samples at the end of a 10-day evaluation period. The authors  
1518 reported a reduction in total bacteria counts of approximately 1 log CFU/g. The results  
1519 identified the potential of active films containing citric acid to extend the shelf life of minced  
1520 beef. However, further research needs to be conducted to improve the limited antimicrobial  
1521 effect demonstrated in this study.

1522

1523 **Sorbic Acid.** García-Soto and others (2015) incorporated 0.5% and 1% of sorbic acid and 8% of  
1524 algal extract (*Fucus spiralis*) into PLA films to protect the flat fish megrim (*Lepidorhombus*  
1525 *whiffiagonis*) from microbial growth. The authors reported a positive antimicrobial effect  
1526 against psychrotrophs with a reduction level of 0.9 log CFU/g in comparison to PE films and  
1527 lower mean values for aerobes and Enterobacteriaceae after 7 days of storage. Although the  
1528 results obtained do not demonstrate a significant antimicrobial effect at the end of the shelf life  
1529 (11 days), improved sensory properties (external odor, gill appearance, and odor) were  
1530 reported for megrim packed with active films, while control samples were considered  
1531 unacceptable from a sensorial point of view. Limjaroen and others (2005) incorporated sorbic  
1532 acid into solvent cast poly(vinylidene chloride) (PVDC) films. Beef bologna and cheddar cheese,  
1533 inoculated with *L. monocytogenes* ( $10^3$  and  $10^5$  CFU/g each), were wrapped in PVDC films  
1534 containing 1.5 or 3% w/v sorbic acid. After 28 days of storage at 4 °C, lower *L. monocytogenes*

1535 counts were obtained in beef bologna samples packed with active films and inoculated with  $10^5$   
1536 CFU/g (4.4 log lower for both sorbic acid films, compared to the control). In the inoculated  
1537 cheddar cheese samples, the active films did neither significantly affected the growth of the  
1538 inoculated *L. monocytogenes* nor that of mesophilic aerobic bacteria after 35 days of storage at  
1539 4 °C. Beef samples inoculated with  $10^3$  CFU/g, in contrast, demonstrated 6.5 and 7.2 log lower  
1540 *L. monocytogenes* counts for 1.5 and 3% sorbic acid films, respectively, compared to the  
1541 control. Moreover, mesophilic aerobic bacteria and LAB counts in the beef packages with the  
1542 active films were found to be around 4 and 6 log lower than in control samples with initial *L.*  
1543 *monocytogenes* inoculums of  $10^5$  and  $10^3$  CFU/g, respectively. This research, however, has  
1544 some drawbacks, especially because the use of sorbic acid as an additive in meat products is  
1545 restricted according to Commission Regulation (EU) No 1129/2011 (European Commission  
1546 2011). Sorbic acid can only be used for selected applications for meat products, such as aspic,  
1547 pate, and surface treatment of dried meat products, jelly coatings, and collagen-based casings  
1548 of meat products at the maximum level of 1 g/kg or *quantum satis*. Moreover, the reported  
1549 sample preparation, a solution-casting, laboratory-scale method, cannot be applied on an  
1550 industrial scale. This trial should be repeated using melt (extrusion), however, the high  
1551 temperature may influence the sorbic acid activity.

1552

1553 **Potassium sorbate.** Cestari and others (2015) developed thermoplastic starch/PBAT blended  
1554 films with 5% potassium sorbate content to prevent microbial growth in restructured chicken  
1555 steaks during frozen storage. After 30 days of storage, *Escherichia coli* (initial count 1.94 log  
1556 CFU/g) was not detected in samples packed with potassium sorbate films, while *E. coli* was

1557 detected in control samples (1.3 log CFU/g). In chicken steaks packed with the active films, *E.*  
1558 *coli* was kept under the detection limit until completion of frozen storage (150 days). Kaya and  
1559 others (2015) reported the use of potassium sorbate and/or sodium lactate (3% applied alone  
1560 and 1.5% of each, applied in combination) in brine to protect smoked rainbow trout  
1561 (*Oncorhynchus mykiss*) fillets against microbial growth. The trout fillets were kept in 8% NaCl  
1562 brine for 12 h before smoking. After 4 weeks of storage at  $6 \pm 1$  °C, total aerobic mesophilic  
1563 bacteria counts were shown to be about 3, 2.1 and 1.7 log CFU/g lower for trout kept in  
1564 potassium sorbate, sodium lactate and its combination, respectively, compared to the control  
1565 (about 8.2 CFU/g) kept in brine without preservatives. Similar results were observed for yeast  
1566 and molds. Additionally, after 5 weeks of storage an identification of the bacteria species in the  
1567 applied samples was performed. With the exception of the trout kept in potassium sorbate  
1568 brine, *Serratia liquefaciens*, which is considered as one of the main pathogens and spoilage  
1569 bacteria in smoked fish, was the dominating species. This indicates that potassium sorbate was  
1570 effective against this pathogen as it was not present in the corresponding samples.

1571

1572 **Potassium metabisulfite.** Several fruits and vegetables are highly susceptible to enzymatic  
1573 changes, and the application of some antimicrobials can provide additional properties against  
1574 this problem. Foralosso and others (2014) tested PVC films that contained a 0.1, 1, or 2% w/w  
1575 mixture of pure and encapsulated potassium metabisulfite (ratio 1:1) as an active  
1576 (antimicrobial, antioxidant and antibrowning) substance. Cut Gala apples (*Malus domestica*)  
1577 were wrapped in the active PVC films, and stored at 4, 8, 12, 16, and 20 °C and 30% RH.  
1578 Samples wrapped in PVC films with 1 and 2% potassium metabisulfite mixtures resulted in a

1579 lower browning index which was rated to be around 60 and 50%, respectively, compared to the  
1580 control (around 90%), and a shelf-life extension from 4 to 8 days for apples stored at 8 °C was  
1581 reported. Samples wrapped using 2% potassium metabisulfite mixtures and stored at 4 °C  
1582 demonstrated toxicological and microbiological stability (migration of sulfites below 10 mg/kg  
1583 SO<sub>2</sub>, according to Brazilian regulation for plastic materials in contact with food; and microbial  
1584 counts below 10<sup>6</sup> CFU/g, considered as the quality threshold by the authors) throughout the 20-  
1585 day storage period. The active film provided the conditions suitable for apple consumption up  
1586 to 12 days of storage at 8 and 12 °C complying with the microbiological contamination limit of 6  
1587 logs CFU/g.

1588

1589 **Oxidized regenerated cellulose.** Sezer and others (2016) incorporated oxidized regenerated  
1590 cellulose micro-particles (4% w/w) in poly(ε-caprolactone (PCL) films and evaluated their  
1591 antibacterial activity on packed sliced salami inoculated with *L. monocytogenes* (10<sup>4</sup> CFU/g).  
1592 After 14 days of storage at 4 °C in contact with the active PCL films, 50% of total colony-forming  
1593 units (about 8 log CFU/g) of *L. monocytogenes* did not survive. The packaging also led to a  
1594 decrease in the growth of *E. coli* and *S. aureus*. Moreover, active films containing 4% of oxidized  
1595 regenerated cellulose micro-particles reduced the oxygen and water permeability by 93 and  
1596 70%, respectively.

1597

1598 **Allyl isothiocyanate.** A high antibacterial activity is reported for allyl isothiocyanate (AITC)  
1599 against a wide range of bacteria (Kim and others 2015). Pang and others (2013) reported the  
1600 positive effect of using AITC (18 and 36 µg/L) in the vapor phase when applied, alone and in

1601 combination with MAP (49% CO<sub>2</sub>/0.5% O<sub>2</sub>/50.5% N<sub>2</sub>), to catfish fillets stored at different  
1602 temperatures. The authors observed that AITC (alone or in combination with MAP) had an  
1603 antimicrobial effect against *Pseudomonas aeruginosa* and extended the shelf life of fresh  
1604 catfish fillets from 4 to 5 days (18 µg AITC/L), 11 (36 µg AITC /L) and 23 days (MAP combined  
1605 with both concentrations of AITC) at 8 °C. The latter applications maintained the *P. aeruginosa*  
1606 counts at a level of about 3 CFU/g during 23 days, compared to the control without MAP (about  
1607 9 log CFU/g after 7 days) and with MAP (about 7.5 log CFU/g after 12 days). At 15 and 20 °C, the  
1608 combination of both technologies was not as effective as at 8 °C, but still extended the shelf life  
1609 at least 2.6 times compared to the controls. No sensory analysis of catfish fillets was performed  
1610 at the end of storage. However, due to the pungency and strong smell of AITC, a sensory  
1611 analysis of the final product should be performed to assure the acceptability of the product. In  
1612 the context of odor, Kim and others (2015) recommend the application of AITC in vapor phase  
1613 and low concentration (0.02-2500 mg/mL) to avoid negative impact on food.

1614

1615 Commercial packaging solutions containing AITC can be found in a variety of formats (sheets,  
1616 labels and films) on the Japanese market under the trademark Wasaouro™ (Mitsubishi-Kagaku  
1617 Foods Corporation, 2002). However, even though antimicrobial tests with AITC (occurring in  
1618 mustard) were successful on several types of food products (Kim and others 2015) and it has  
1619 been given GRAS status (FDA 2006), it has to be emphasized that the regulations in specific  
1620 countries can differ and data regarding current status can change (such as the approval status  
1621 for AITC in EU and USA). In 2010, the EFSA panel on food additives and nutrient sources added  
1622 to food (ANS) gave its scientific opinion on the safety of allyl isothiocyanate for the proposed

1623 uses as a food additive. Therein, it is stated that AITC is “*an efficient alternative to already*  
1624 *approved preservation techniques for a range of foods,*” such as bakery products (including all  
1625 types of pre-packed bread and fine bakery ware), all types of cheese, fruits, and vegetables  
1626 (EFSA ANS Panel, 2010). To give another example, for sorbic acid, and its derivatives, such as  
1627 potassium sorbate, the EFSA has re-evaluated their status as food additives in 2015 (EFSA ANS  
1628 Panel 2015). The main hurdle to commercialization of active packaging solutions containing  
1629 organic compounds are regulatory requirements. Therefore, research efforts should be focused  
1630 on the development of tailor-made active packaging solutions that comply with the specific  
1631 legislation for each food product.

1632

### 1633 **Nanoparticles**

1634 Antimicrobial nanomaterials represent an increasingly important component of some active  
1635 packaging for food applications (Ayhan 2013). Antimicrobial nanoparticles (particles between 1-  
1636 100 nm in size) are incorporated into a polymer matrix with the aim of prolonging the shelf life  
1637 of packaged food. High surface-to-volume ratio and enhanced surface reactivity of the nano-  
1638 sized antimicrobial agents cause inactivation of microorganisms more effectively than their  
1639 micro or macro-scale counterparts (Radusin and others 2016). The preparation of food  
1640 packaging materials depends on the nature of the nanoparticle, its size, and its specific surface  
1641 area.

1642

1643 Despite the large number of studies reported in the literature in this area, there are only a few  
1644 studies incorporating real food systems. Commonly used or tested antimicrobial nanoparticles

1645 are metal ions (silver, copper, gold, platinum), metal oxide (titanium dioxide, zinc oxide,  
1646 magnesium oxide), and organically modified nano-clays. From ancient times, silver (Ag) has  
1647 been used as an antimicrobial agent. Its ability as an antimicrobial agent increases in nano-  
1648 dimension and, hence, there are now many studies with Ag nanoparticles incorporated in food  
1649 packaging materials as antimicrobial agents (Panea and others 2014; Azlin-Hasim and others  
1650 2016; Li and others 2017) (Table 11). The most recent studies illustrate that addition of Ag  
1651 nanoparticles into different polymer matrices, in combination with other additives or  
1652 nanoparticles, can significantly prolong the shelf life of different foodstuffs. Li and others (2017)  
1653 reported that rice stored in LDPE without Ag/TiO<sub>2</sub> showed a serious mildew condition after one  
1654 month with increased total plate counts (TPC) from 4.84 to 7.15 log cfu/g, while the rice stored  
1655 in a nanocomposite based on LDPE with Ag/TiO<sub>2</sub> had a low TPC of 5.48 log cfu/g. Mihaly  
1656 Cozmuta and others (2015) reported that the microbiological safety of bread stored in Ag/TiO<sub>2</sub>-  
1657 based packaging inhibited the proliferation of yeast/molds, *B. cereus*, and *B. subtilis*. The shelf  
1658 life of bread was extended by reducing the degradation rate of the main nutritional compounds  
1659 compared to the bread stored in an open atmosphere or in a commonly used plastic packaging.  
1660 Azlin-Hasim and others (2016) prepared nanocomposite material based on PVC and silver  
1661 nanoparticles, and they reported that this significantly extended the product shelf life and  
1662 resulted in lower lipid oxidation of chicken breast fillets, while Panea and others (2014)  
1663 reported reduction in MO but with higher lipid oxidation.

1664

1665 Emamifar and others (2010) conducted a study on the antimicrobial activity of LDPE loaded  
1666 with nano-silver and zinc oxide (ZnO) for the packaging of orange juice. This system was very

1667 effective in prolonging the shelf life of orange juice (up to 28 days). ZnO has also been used as  
1668 an antimicrobial agent added to active packaging films for packaging fresh poultry meat by  
1669 Akbar and Anal (2014), and they showed a reduction of the initial bacterial counts (*S. aureus*  
1670 and *S. typhimurium*) by 2 log within 24h of incubation at  $8\pm 1$  °C. After 6 days there were no  
1671 viable cells of *S. aureus*, and no *S. typhimurium* after 8 days of incubation.

1672

1673 Titanium dioxide (TiO<sub>2</sub>) has been studied as antimicrobial nanoparticles in LDPE for the  
1674 packaging of fresh pears, and a decrease in mesophilic bacteria from 3.14 to less than 2 log  
1675 CFU/g for the entire storage period (17 days) was recorded, whereas for neat LDPE cell loads  
1676 increased from 3.19 to 4.02 log CFU/g. Furthermore, yeasts decreased from 2.45 to less than 2  
1677 log CFU/g, whereas those for the control sample increased from 2.1 to 3.37 log CFU/g (Bodaghi  
1678 and others 2013). In addition, copper (Cu) was effective against *Pseudomonas* spp. (isolated  
1679 from spoiled fiordilatte cheese) when incorporated in PLA and used for packaging of fiordilatte  
1680 cheese. A delay in microbial proliferation was recorded when the active films were used (Conte  
1681 and others 2013).

1682

1683 As reported in the previous sections, the use of antimicrobial nanoparticles has great potential  
1684 in preserving the microbial quality of the food systems. In this context, the appropriate  
1685 antimicrobial agent needs to be selected according to the targeted food. Additionally, the  
1686 impact of nanoparticles on the properties of the packaging films, such as barrier properties and  
1687 transparency, should also be considered. However, the safety evaluation and approval for use  
1688 of such nanoparticles in food packaging remains the greatest challenge due to the difficulties in

1689 the evaluation of the safety of nanoparticles in general (Radusin and others 2016) as well as  
1690 constraints associated with the current legislative landscape (Amenta and others 2015, Radusin  
1691 and others 2016, Rauscher and others 2017).

1692

1693 Over the last decade, various studies have been conducted in this area and several scientific  
1694 reviews have been published. However, these have mostly focused on technology and several  
1695 mechanisms, as well as “*in vitro*” studies on culture media. There has been little research  
1696 involving real food packaging systems. Such research, however, is of great importance, since  
1697 the antimicrobial activity of the active agents with culture media does not necessarily correlate  
1698 with the antimicrobial activity in the food. This is mainly due to the complex structure of the  
1699 food as well as the differences in the antimicrobial activity test conditions.

1700

1701 Before an antimicrobial food packaging can be successfully developed, a number of factors  
1702 have to be considered. Firstly, the food system has to be fully understood in terms of its  
1703 components, and physical and chemical characteristics, such as pH, and water activity, as well  
1704 as its microbiological aspects, including identification of those microorganisms that are  
1705 desirable and undesirable. A suitable antimicrobial active agent should be selected with respect  
1706 to all these characteristics. In particular, the antimicrobial spectrum and the efficiency of the  
1707 agent should target the microorganisms that limit the shelf life of the particular food. According  
1708 to the international standard on the measurement of antibacterial activity of plastics and other  
1709 non-porous surfaces (ISO 22196 2011), derived from Japanese Industrial Standard (JIS Z 2801  
1710 2000), a decrease of the number of microorganisms in the magnitude of 2 log colony forming

1711 units (CFU)/cm<sup>2</sup> is required to demonstrate antimicrobial efficacy. In food systems, shelf-life  
1712 tests have to be performed to evaluate the efficiency of the antimicrobial film for the selected  
1713 product. In this context, the maximum permitted level of microorganisms in a food is very  
1714 specific and depends on several factors, such as the type of microorganisms (spoilage or  
1715 pathogenic), the type of food and the regulations in force in the country where the product will  
1716 be marketed. In the EU, for instance, the microbiological criteria for foodstuffs are regulated by  
1717 the Commission Regulation (EC) No 2073/2005) (European Commission 2005). For some food  
1718 systems, such as several bakery products, no visual mold growth should be observed, whereas  
1719 for others, the number of microorganisms should not exceed a certain number. Additionally,  
1720 the influence of the food on the efficiency of the antimicrobial agent should be considered  
1721 since the agent may be entrapped or deactivated by the food component, or the activity of the  
1722 agent may be affected by a low or high pH.

1723

1724 A second consideration is the storage conditions of the packed food since the temperature or  
1725 relative humidity may affect the release and/or the efficiency of the active agent. A third factor  
1726 involves selection of antimicrobial agents that do not cause any undesired changes in the food,  
1727 such as the sensory properties. The last aspect to consider is that the addition of antimicrobial  
1728 agents should not result in undesirable changes in the packaging material, such as barrier,  
1729 sealing and adhesion properties, transparency, or glossiness, and it should not cause any  
1730 increase in the migration of substances from the packaging material to the food.

1731

1732 **Conclusion**

1733 Extensive research on the development of new active packaging technologies has been  
1734 conducted over recent years generating a wide variety of active packaging systems that may be  
1735 applied to extend the shelf life of food products. This review highlights the huge potential of  
1736 active packaging systems and concludes that challenges in the implementation of new  
1737 technologies to real food applications are similar across all the active packaging technology  
1738 categories discussed. Food products are very complex systems and packaging parameters are  
1739 highly product-specific. Thus, to achieve an optimal activity or capacity of the desired active  
1740 packaging system, product-tailored concepts have to be applied. Thereby, it is crucial to  
1741 consider all the influencing factors, such as the physical/chemical/physiological properties of  
1742 the food, packaging size, and storage conditions. Scale up and industrialization of the active  
1743 packaging technologies could be challenging and therefore should be taken into consideration  
1744 at early development state for successful commercialization. The cost of the implementation of  
1745 the technology has to correspond with the benefit gained by the particular food product,  
1746 legislative and regulatory issues must be addressed, and broad consumer acceptance is  
1747 required. A successful collaboration between research institutes and industry, including  
1748 development, legislative and commercial functions, is required to overcome these challenges.  
1749 However, the recent advances discussed in this review can provide food and packaging  
1750 scientists with a better understanding of the potential and the benefits of active packaging  
1751 technologies and, hence, assist in accelerating their commercial adoption.

1752

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1761 The authors made the following contributions to the manuscript:

1762 Selcuk Yildirim: Corresponding author. Development of concept of manuscript and distribution  
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1764 Bettina Röcker: Coordination of authors’ contributions. Abstract, Introduction, Conclusion,  
1765 Oxygen scavengers, Moisture scavengers, References

1766 Marit Kvalvåg Pettersen: Carbon dioxide emitters

1767 Julie Nilsen-Nygaard: Carbon dioxide emitters, Moisture scavengers

1768 Zehra Ayhan: Ethylene absorbers

1769 Ramune Rutkaite: Antioxidant releasers; Antimicrobial packaging systems (enzymes and  
1770 bacteriocins, polymers)

1771 Tanja Radusin: Antimicrobial packaging systems (nanoparticles)

1772 Patrycja Suminska: Moisture scavengers; Antimicrobial packaging systems (organic acids, their  
1773 derivatives and other organic compounds)

1774 Begonya Marcos: Antioxidant releasers, Moisture scavengers, Antimicrobial packaging systems  
1775 (organic acids, their derivatives and other organic compounds)

1776 Veronique Coma: Antimicrobial packaging systems (essential oils)

1777

1778

1779

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2591 **Tables**

2592 See separate file